

CONTENT OF PHENOLIC ACIDS AND TOTAL PHENOLICS IN SEVERAL AQUATIC PLANT SPECIES

L. DJURDJEVIĆ

*Institute for Biological Research "Siniša Stanković",
11060 Belgrade, Yugoslavia*

Abstract - According to the content of free phenolic compounds, the five stagnant water plant species examined in the present work can be arranged in the following order: *Nymphaea alba* > *Lemna minor* > *Salvinia natans* > *Trapa natans* > *Ceratophyllum demersum*. These plants contained more of free *p*-coumaric, ferulic, vanillic and syringic acid comparing to bound forms (ester linked) of these compounds. In addition to these phenolic acids, in the bog sediment *p*-hydroxybenzoic acid was detected mainly in its bound form. Dissolved phenolics and vanillic acid were identified in the bog water, as well.

UDC 581.526.3:547.562

INTRODUCTION

Within the scope of allelopathic investigations performed so far, much attention has been paid to interrelationship of terrestrial plant species. Phenolic compounds identified as the most frequently found allelochemical products were shown to act inhibiting numerous physiological processes leading to a suppression of some plant species within a phytocoenosis (Whittaker 1970; Chou and Muller 1972; Rice 1974; Lodhi 1976, 1978; Colton and Einhellig 1980; Li *et al.* 1992). However, similar data on aquatic plants of stagnant waters are rather scarce in the available literature. It has been reported earlier that some aquatic plants in Yugoslavia are conspicuously dominant forming submerged meadows which cover rather large areas (Micevski 1962, 1963, 1969; Rauš *et al.* 1978). Due to their submerged and emersed vegetation, bogs represent suitable sites for allelopathic examinations. Since aquatic plants are either completely or partially submerged allelochemicals must be transmitted from donor (emitting) plant to acceptor (affected) plant through either their rhizosphere soil or water. In the present work content of phenolic acids (PAs) and total phenolics (TP) in five aquatic plant species, sediment and water samples of a bog ecosystem was determined.

MATERIAL AND METHODS

Plant species

The bog examined throughout the present work is situated along the Danube in the vicinity of Belgrade (opposite shore of Višnjička banja locality). It is inhabited by several emersed (*Lemna minor*, *Nymphaea alba*, *Salvinia natans* and *Trapa natans*) and one submerged (*Ceratophyllum demersum*) plant species. Phenolic compounds were determined either in whole plants (*Lemna minor*, *Salvinia natans* and *Ceratophyllum demersum*) or in the leaves (*Nymphaea alba* and *Trapa natans*). The samples (500 g each) were dried, pulverized and sieved (mesh 35).

Analysis of phenolic compounds

Phenolic acids and total phenolic compounds were extracted from 2 g of dry plant material with 80% boiling ethanol solution followed by ethyl acetate (3 x 30 mL, 8 h). Pooled ethanol and ethyl acetate extracts were evaporated to the aqueous phase. After that, pH was adjusted to 2.0 with 2 N HCl, free phenolics transferred to ethyl acetate and evaporated to dryness. Dry residue was dissolved in 4 mL of 80% ethanol solution and maintained at -20 °C till the analysis. For the ex-

traction of bound phenolics (esterified forms) dry plant material remained after the extraction of free phenolics was boiled in 2 N HCl during 60 min and the extract was transferred into ethyl acetate. The mixture was evaporated and the dry residue dissolved in 4 mL of 80% ethanol was kept at -20 °C before the analysis (Mijđla *et al.* 1975).

Aliquots of the bog water (3.0 L) were evaporated to the volume of 50 mL. After 60 min of hydrolysis in 2N HCl total phenolics and phenolic acids were transferred into ethyl acetate and the samples were evaporated to dryness. Dry residue was dissolved in 4 mL of 80% ethanol solution and kept as above.

Samples of the sediment (2 kg) were collected at littoral, mid- and the deepest parts of the bog and dried. Free phenolics were extracted from 30 g of dry sediment with boiling ethyl acetate (30 x 50 mL) for 24 h. For the extraction of bound phenolics (esterified forms) the residues remaining upon the first extraction were resuspended in 15 mL of 2 N NaOH and boiled for 24 h. After that, pH of the aqueous phase was adjusted to 2.0 with concentrated HCl, the mixture was transferred to ethyl acetate and evaporated. Dry residues were dissolved in 4 mL of 80% ethanol and maintained at -20 °C until the analysis (Hennequin and Juste 1967; Katase 1981a,b).

Identification and quantification of phenolic compounds

Content of total phenolics (free and bound) was determined spectrophotometrically using Folin-Ciocalteu reagent (Feldman and Hanks 1968). Ferulic acid was employed as a reference. For both qualitative and quantitative analyses, ascending two-dimensional paper chromatography was applied using the mixture of isopropyl alcohol, ethyl acetate, NH₄OH and H₂O (30:50:1:19) and 2% aqueous (v/v) acetic acid as developers for the first and the second dimension, respectively. Dried chromatograms were sprayed with *p*-nitroaniline and 20% Na₂CO₃. The spots of phenolic acids were eluted with 45% ethanol solution and absorbancies were measured at the corresponding wavelengths at a Shimadzu UV 160 spectrophotometer (Mijđla *et al.* 1975). This method provides a satisfactory separation of five PA, two of which represent cinnamic acid derivatives (ferulic acid-4-hydroxy-3-methoxycinnamic and *p*-coumaric acid-*trans*-4-hydroxycinnamic acid) and three represent benzoic acid derivatives (*p*-hydroxybenzoic acid, vanillic acid-4-hydroxy-3-methoxybenzoic acid and syringic-3,5-dimethoxy-4-hydroxybenzoic acid). All analyses were done in triplicates in three independent experiments.

RESULTS AND DISCUSSION

Four of the five examined species are angiosperms and only one, *Salvinia natans*, belongs to pteridophytes. Content of total free phenolic compounds exceeded that of their bound forms with the exception of *Trapa natans* where an opposite situation was recorded. Based on the content of total free phenolics the above aquatic plant species can be arranged in the following order: *Nymphaea alba* > *Lemna minor* > *Salvinia natans* > *Trapa natans* > *Ceratophyllum demersum* (Table 1). Ratio of free and bound phenolic compound forms was the highest in *Lemna minor* (4.42) and the lowest in *Trapa natans* (0.66).

Table 1. Content of total phenolics and phenolic acids in several aquatic plants

		<i>Salvinia natans</i>	<i>Trapa natans</i>	<i>Lemna minor</i>	<i>Ceratophyllum demersum</i>	<i>Nymphaea alba</i>
Total Phenolics (mg/g)	free	12.50	9.77	15.18	3.80	16.86
	±	1.05	0.94	1.18	0.24	0.97
	bound	4.65	14.69	3.43	1.34	9.60
	±	0.19	0.27	0.07	0.10	0.95
	total	17.15	24.46	18.61	5.14	26.46
PAs (µg/g)						
<i>p</i> -coumaric	free	-	201.01	93.27	84.10	87.31
	±	-	19.49	9.78	7.45	8.91
	bound	-	45.32	-	-	-
	±	-	3.87	-	-	-
Ferulic	free	-	194.32	155.92	238.52	113.27
	±	-	20.91	17.06	21.44	10.97
	bound	-	-	-	-	-
	±	-	-	-	-	-
<i>p</i> -hydroxybenzoic	free	-	-	tr.	-	-
	±	-	-	-	-	-
	bound	-	-	24.71	-	-
	±	-	-	3.02	-	-
Vanillic	free	73.63	92.98	76.39	50.76	94.37
	±	6.42	9.07	8.65	4.62	8.97
	bound	15.44	23.70	24.37	15.16	24.02
	±	1.42	1.98	2.07	1.91	2.07
Syringic	free	-	154.76	98.33	253.69	-
	±	-	16.82	10.36	20.91	-
	bound	-	33.24	-	32.67	-
	±	-	2.74	-	2.98	-
Total	free + bound	89.07	745.33	472.99	674.90	318.97

A pteridophyte *Salvinia natans* contained only vanillic acid, predominantly as a free form. In the remaining species free forms of *p*-coumaric, ferulic, vanillic and syringic acid were dominant. Ferullic acid represents a dominant phenolic compound making 30-38% of total phenolic acids in all plant species examined, with the exception of *S. natans*. Only bound form of vanillic acid was common to all five plant species. In addition to the above phenolic acids, *Lemna minor* was found to contain *p*-hydroxybenzoic acid which was not detected in other plants examined.

According to the data reported so far, different phenolic compounds were identified in aquatic plants, e.g. tellimagrandin II in *Myriophyllum spicatum* (Gross *et al.* 1996), gallotannin and two ellagitannins in the root of *Nuphar variegatum* (Nishizawa *et al.* 1990), orientin, vitexin, apigenin and luteolin in the chloroplasts of *Spirodela intermedia* and *S. polyrhiza*, Lemnaceae (Saunders and McClure 1976). In the leaves of a marine phanerogam *Posidonia oceanica* *p*-coumaric, ferulic, *p*-hydroxybenzoic, cinnamic, sinapic and *p*-anisic acid were recorded together with some other phenolic compounds (Cuny *et al.* 1994, 1995), ferulic acid being the most abundant among phenolic acids. An intestine content of some cyprinid fishes composed of microalgae, contained 71.98-105.06 $\mu\text{g/g}$ of free and 15.88-32.26 $\mu\text{g/g}$ of bound phenolics. Phenolic acids found were: *p*-hydroxybenzoic and vanillic acid in free and bound form (Djurđević *et al.* 1997).

Littoral samples of the bog sediment contained several fold lower amounts of total free phenolics in comparison with those taken from the mid- and the deepest parts of the bog (Table 2). Content of bound forms of the phenolic compounds was several fold higher than that of free forms. Bound forms were unevenly distributed in the bog sediment, littoral and mid-part samples containing only vanillic acid while in those from the deepest parts of the bog *p*-coumaric, ferulic, vanillic and syringic acid were detected.

Phenolic acids were identified in different types of soil. Soil from different forest communities was found to contain several phenolic acids in a concentration range from 0.09-1,100 $\mu\text{g/g}$ (Lodhi 1975; Kuiters and Denneman 1987). Peat bog soil is characterized by significantly higher levels of phenolic acids, bound forms being predominant (Katase 1981a,b; Katase and Kondo 1984). Katase (1983) found in a pond sediment *p*-coumaric, ferulic, *p*-hydroxybenzoic and vanillic acid both as free and bound forms in concentrations similar to those observed in the present work. Levels of phenolic compounds in bog sediments are significant for emerged plants such as *Nymphaea alba* and *Trapa natans* with the roots develop-

Table 2. Content of total phenolics and phenolic acids in the bog sediment.

		littoral part	mid-part	deepest part
Total phenolics ($\mu\text{g/g}$)	free	3.92	17.55	10.74
	±	0.59	1.42	0.96
	bound	251.82	337.75	224.42
	±	20.98	30.04	18.67
	total	255.74	355.30	235.16
PAs ($\mu\text{g/g}$)				
<i>p</i> -coumaric	free	tr.	-	-
	±	-	-	-
	bound	-	-	7.94
ferulic	±	-	-	0.73
	free	-	-	tr.
	bound	-	-	10.89
<i>p</i> -hydroxybenzoic	±	-	-	0.12
	free	0.75	0.75	0.87
	bound	-	tr.	-
vanillic	±	-	-	-
	free	0.68	0.87	0.76
	bound	34.58	8.86	4.81
syringic	±	2.46	0.91	0.37
	free	-	-	tr.
	bound	-	-	11.06
total	±	-	-	0.92
	free + bound	36.01	10.48	36.33

ed in the sludge. Inhibitory effects of phenolics accumulated in soil on plants have been repeatedly demonstrated (Whittaker 1970; Chou and Muller 1972; Rice 1974; Lodhi 1976, 1978).

Samples of the bog water in which aquatic plants are either completely or partially submerged contained 785.28 $\mu\text{g/L}$ of total phenolics, while among phenolic acids only vanillic acid was identified in a concentration of 52.54 $\mu\text{g/L}$. Similar to terrestrial plants, aquatic plants release phenolic compounds into their environment. *Myriophyllum spicatum* L. propagated fast in the eastern United States and Canada after its introduction from

Europe during the end of the last century. It may displace the native vegetation and milfoil-dominated lakes usually have low phytoplankton densities. It was recently reported that *Myriophyllum spicatum* releases polyphenols acting as inhibitors of cyanobacteria, chlorophytes and diatoms. A part of the inhibitory activity is due to complexation and inactivation of algal extracellular enzymes by hydrolysable polyphenols from *M. spicatum* (Gross *et al.* 1996).

The presence of phenolic compounds in aquatic plants, as well as in the samples of both the bog sediment and water together with the dominance of one to two plant species suggest allelopathic influence of the released phenolics on other plant species within the aquatic ecosystem examined. Further studies along this line are in progress.

Acknowledgements - This work was supported by Ministry for Science and Technology of Serbia, contract #03E15.

REFERENCES

- Chou, C.H. and Muller, C.H. (1972). Allelopathic mechanisms of *Arctostaphylos glandulosa* var. *zacaensis*. *Amer. Midl. Natur.* **88**, 324-347.
- Colton, C.E. and Einhellig, F.A. (1980). Allelopathic mechanisms of velvetleaf (*Abutilon theophrasti* Medic., *Malvaceae*) on soybean. *Amer. J. Bot.* **67**, 1407-1413.
- Cuny, P., Serve, L., Jupin, H. and Boudouresque, C.F. (1994). Les composés phénoliques hydrosolubles de *Posidonia oceanica* (phanerogame marine) dans une zone colonisée par la chlorophyte introduite *Caulerpa taxifolia* (Alpes maritimes, France, Méditerranée). First International Workshop on *Caulerpa taxifolia*. Boudouresque C.F., Meinesz A. and Gravez V. edit., GIS Posidonie publ., Fr., 1994, 355-364.
- Cuny, P., Serve, L., Jupin, H., and Boudouresque, C.F. (1995). Water soluble phenolic compounds of the marine phanerogam *Posidonia oceanica* in a Mediterranean area colonised by the introduced chlorophyte *Caulerpa taxifolia*. *Aquat. Bot.* **52**, 237-242.
- Djurđević, L., Cakić, P. and Kataranovski, D. (1997). Phenolcarboxylic acids and total phenolics of plant origin in intestine content of three Cyprinid fishes from the Danube River. *Fisheries Sci.* **63**, 841-842.
- Feldman, A.W. and Hanks, R.W. (1968). Phenolic content in the roots and leaves of tolerant and susceptible citrus cultivars attacked by *Rodopholus similis*. *Phytochemistry*, **7**, 5-12.
- Gross, E.M., Meyer, H. and Schilling, G. (1996). Release and ecological impact of algicidal hydrolysable polyphenols in *Myriophyllum spicatum*. *Phytochemistry*, **41**, 133-138.
- Hennequin, J. R. and Juste, C. (1967). Présence d'acides phénoliques libres dans le sol: Etude de leur influence sur la germination et la croissance des végétaux. *Ann. agron.* **18**, 545-569.
- Katase, T. (1981a). The different forms in which *p*-coumaric acid exists in a peat soil. *Soil Sci.* **131**, 271-275.
- Katase, T. (1981b). The different forms in which *p*-hydroxybenzoic, vanillic, and ferulic acids exist in a peat soil. *Soil Sci.* **132**, 436-443.
- Katase, T. (1983). The significance to humification of different forms of *p*-coumaric and ferulic acids in a pond sediment. *Soil Sci.* **135**, 151-155.
- Katase, T. and Kondo, R. (1984). Distribution of some different forms of some phenolic acids in peat soils in Hokkaido, Japan: 1. Trans-4-hydroxycinnamic acid. *Soil Sci.*, **138**, 22Q-225.
- Kuiters, A.T. and Denneman, C.A.J. (1987). Water-soluble phenolic substances in soils under several coniferous and deciduous tree species. *Soil Biol. Biochem.* **19**, 765-769.
- Li, H.H., Urashima, M., Amano, M., Lajide, L., Nishimura, H., Hasegawa, K. and Mizutani, J. (1992). Allelopathy of Barnyardgrass (*Echinochloa crus-galli* L. Beauv. Var. *crus-galli*). *Weed Res.* (Japan), **37**, 146-152.
- Lodhi, M.A.K. (1975). Soil-plant phytotoxicity and its possible significance in patterning of herbaceous vegetation in a bottomland forest. *Am. J. Bot.* **62**, 618-622.
- Lodhi, M.A.K. (1976). Role of allelopathy as expressed by dominating trees in lowland forest in controlling the productivity and pattern of herbaceous growth. *Am. J. Bot.* **63**, 1-8.
- Lodhi, M.A.K. (1978). Allelopathic effects of decaying litter of dominant trees and their associated soil in a lowland forest community. *Am. J. Bot.* **65**, 340-344.
- Micevski, K. (1962). Typologiske Untersuchungen der Sumpfvvegetation Mazedoniens. *Ann. Fac. Sci. Univ. Skopje*, **14**, 79-130.
- Micevski, K. (1963). Die Wasser und Sumpfvvegetation des Dojran sees. *Acta Musei Macedon. Sci. Natur.* **8**, 175-195.
- Micevski, K. (1969). Die Wasservegetation der Seen von Ohrid und Prespa. *Acta Musei Macedon. Sci. Natur.* **9**, 61-80.
- Mijđla, N., Haldre, N. and Savisaar, S. (1975). Phenolcarboxylic acids in leaves of apple. *Tr. Fiziol. Biochem. Rast.* **362**, 3-13. Tartu.
- Nishizawa, K., Nakata, I., Kishida, A., Ayer, W.A. and Browne, L.M. (1990). Some biologically active tannins of *Nuphar variegatum*. *Phytochemistry* **29**, 2491-2494.
- Rauš, Đ., Šegulja, N., Topić, J. (1978). Investigations of the swamp and water vegetation in the lowlands of Slavonia. *Acta Bot. Croat.* **37**, 131-147.
- Rice, E.L. (1974). Allelopathy. Academic Press New York, San Francisco, London, PP. 353.
- Saunders, J.A. and McClure, J.W. (1976). The distribution of flavonoids in chloroplasts of twenty five species of vascular plants. *Phytochemistry* **15**, 809-810.
- Whittaker, R.H. (1970). The biochemical Ecology of higher plants. In: Chemical ecology, Eds. Sondheimer, E. and Simeone, J.B. Ac. Press, New York, London, pp. 43-70.

ФЕНОЛНЕ КИСЕЛИНЕ И УКУПНИ ФЕНОЛИ НЕКИХ ВОДЕНИХ БИЉАКА

Л. БУРЂЕВИЋ

Институт за биологију и истраживања "Синица Спанковић",
11060 Београд, Југославија

Рад се бави проучавањем фенолних једињења барских биљака, с обзиром на досадашња недовољна алелопатска проучавања водених екосистема. Према количини укупних слободних фенола којих има више од везаних, може се успоставити следећи низ водених биљака: *Nymphaea alba* > *Lemna minor* > *Salvinia natans* > *Trapa natans* > *Ceratophyllum demersum*. У ткивима проучаваних врста биљака преовлађују слободни облици *p*-кумаринске, ферулинске, ванилинске, и сиригинске

киселине. Муљ узет из приобалног, средњег и најдубљег дела баре садржи исте фенолне киселине, претежно у везаном облику. Растворена фенолна једињења детектована су и у барској води. Присуство фенолних једињења у воденим биљкама, барском муљу и води и доминација 1-2 биљне врсте, указују на алелопатски утицај њихових фенолних једињења на остале биљке у воденом екосистему, што треба даљим истраживањима потврдити.