

## DEVELOPMENT OF THE MOSS *POGONATUM URNIGERUM* (HEDW.) P. BEAUV. UNDER *IN VITRO* CULTURE CONDITIONS

TIJANA CVETIĆ<sup>1</sup>, ANETA SABOVLJEVIĆ<sup>1</sup>, M. SABOVLJEVIĆ<sup>1</sup>, and D. GRUBIŠIĆ<sup>1,2</sup>

<sup>1</sup>*Institute of Botany, Faculty of Biology, University of Belgrade, 11000 Belgrade, Serbia*

<sup>2</sup>*Siniša Stanković Institute for Biological Research, 11060 Belgrade, Serbia*

**Abstract** – *Pogonatum urnigerum* (Polytrichaceae) *in vitro* culture was established from spores collected in nature. Both protonema and gametophore stages of gametophyte development were obtained. Also, a stable callus culture was established using hormone-free nutrient medium. The best nutrient medium for development was half-strength Murashige-Skoog medium supplemented with 1.5% sucrose. Auxin treatment enabled some gametophores to develop, but prolonged treatment induced early senescence. Tissues grown on cytokinin did not produce any gametophytes and did not survive prolonged treatment.

**Key words:** Moss development, *Pogonatum urnigerum*, *in vitro* culture

UDC 582.325.1 : 57.08

### INTRODUCTION

The gametophyte phase of mosses, which is dominant in their life cycle, is a favorable model system for genetic, biochemical, metabolic, and developmental studies (C o v e et al., 2006). It consists of a filamentous stage, the protonema, part of which bears buds that develop into leafy gametophores. However, the establishment of axenic culture is essential for obtaining relevant results, since material from nature is hard to separate from other moss species, microorganisms, invertebrates, and dust and soil particles.

Although it is sometimes stated that bryophytes are easily cultured *in vitro* (C o v e et al., 2006), most of this work has been done on *Physcomitrella patens*, and relatively few species have in fact been successfully induced for form stable axenic cultures.

As in higher plants, development under axenic conditions is species-specific. There are many discrepancies in the response of different moss species to the same treatment under *in vitro* culture (B i j e l o v i ć et al., 2004; S a b o v l j e v i ć et al., 2005). Mosses also contain numerous biologically active compounds (M u e s, 2000; S a b o v l j e v i ć et al., 2001, 2006a), but only a small percent of species have yet been thoroughly stud-

ied. Introduction of new species into axenic conditions and maintenance of stable cell and tissue cultures is therefore essential as a start for in-depth investigation of the physiology and potential uses of bryophytes.

*Pogonatum urnigerum* (Polytrichaceae) is a relatively robust moss with characteristic glaucous green leaves that grows on well-drained acidic soil. The aim of the present study was to establish stable *in vitro* culture of this species and examine its development under axenic conditions.

### MATERIALS AND METHODS

Fully developed *Pogonatum urnigerum* plants were collected on Mt. Kopaonik (SE Serbia) in autumn of 2005. Fresh, unopened sporophytes were surface sterilized by dipping in 25% commercial bleach (8% active NaOCl) for 3 minutes, and thoroughly rinsed in sterile deionized water. The cap was then removed and spores released on nutrient medium.

Two nutrient media were tested: MS [containing mineral salts and vitamins according to Murashige and S k o o g (1962), 3% sucrose, and 0.7% agar] and MS/2 (containing half-strength Murashige-Skoog mineral salts and vitamins, 1.5% sucrose, and 0.7% agar). In some ex-

periments,  $10^{-6}$  M indole-3-acetic acid (IAA) or benzyladenine (BA) was added to the MS/2 medium. Prior to sterilization, pH was adjusted to 5.8. Cultures were grown at  $25\pm 2^\circ\text{C}$  under long-day conditions (16 h light/8 h dark). Light was provided by fluorescent tubes, and irradiance on the plant-growth shelf was  $45 \mu\text{mol}\cdot\text{s}^{-1}\cdot\text{m}^{-2}$ .

Fresh microscopic preparations were examined using a Leica DMLS microscope with a digital camera attached.

## RESULTS

Spore germination began soon after introducing spores to nutrient media. Independent of the nutrient medium used (hormone-free MS or MS/2 medium), spore germination was evident after 8 days of *in vitro* cultivation. (Fig. 1). Primary protonemata that emerged from germinated spores developed rapidly, and after three weeks both chloronemata and caulonemata were visible (Fig. 2). Even at this early stage caulonemata began to create bud-like structures.

At this stage further development was inhibited on MS medium, causing disintegration of chlorophyll and cessation of growth. Only a few “islets” of green callus-like tissue remained and continued growth as a callus tissue when transferred to MS/2 medium. Protonemata obtained from spores germinated on MS/2 medium continued elongation and formed many buds. In all further experiments, only MS/2 medium was used. After 90 days of subcultivation, a small percent of buds growing on MS/2 medium developed into plantlets (Fig. 3), while at the same time some explants transformed into calli (Fig. 4).

To test factors that influence further bud development, protonemata with buds were transferred to MS/2 media containing either  $1 \mu\text{M}$  IAA or  $1 \mu\text{M}$  BA. In the control group grown on hormone-free MS/2 medium, some protonemata developed plants in the course of 30 days. In IAA treated protonemata, only a few plantlets appeared, while protonemata began to lose chlorophyll without callus formation. After 30 days of BA treatment only non-green senescent calli and protonemata were visible (Fig. 5)

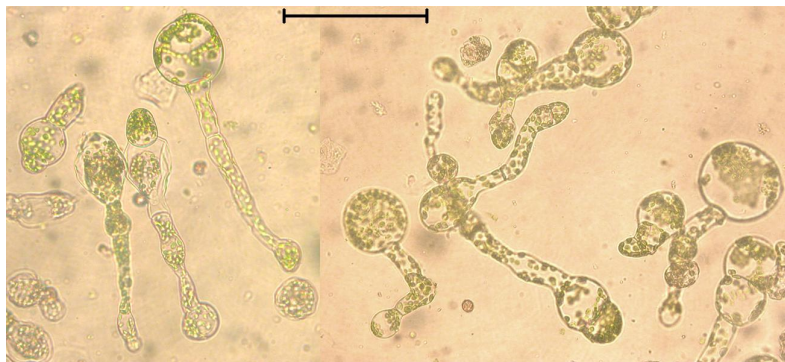


Fig. 1. *Pogonatum urnigerum* germinated spores 8 days after introduction on nutrient medium; the bar represents 100  $\mu\text{m}$ .

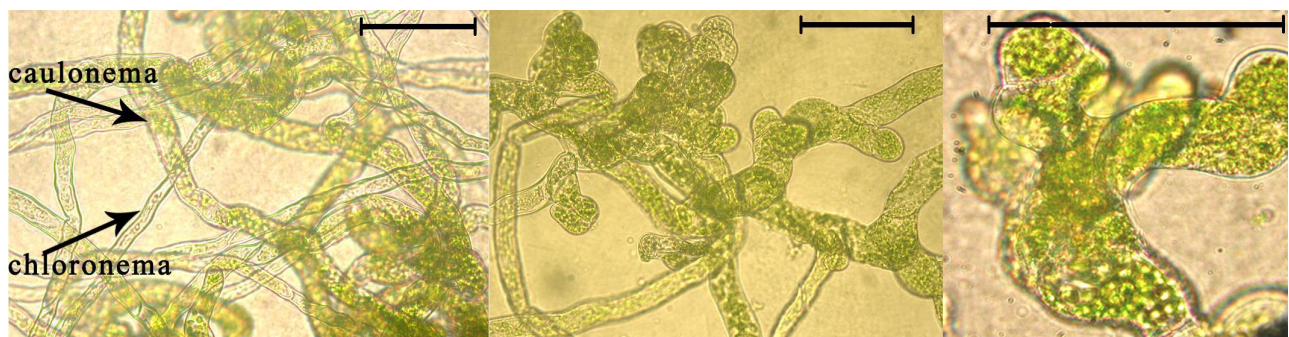


Fig. 2. Protonemata and early stages of bud formation 30 days after introduction of spores on MS/2 nutrient medium; the bar represents 100  $\mu\text{m}$ .



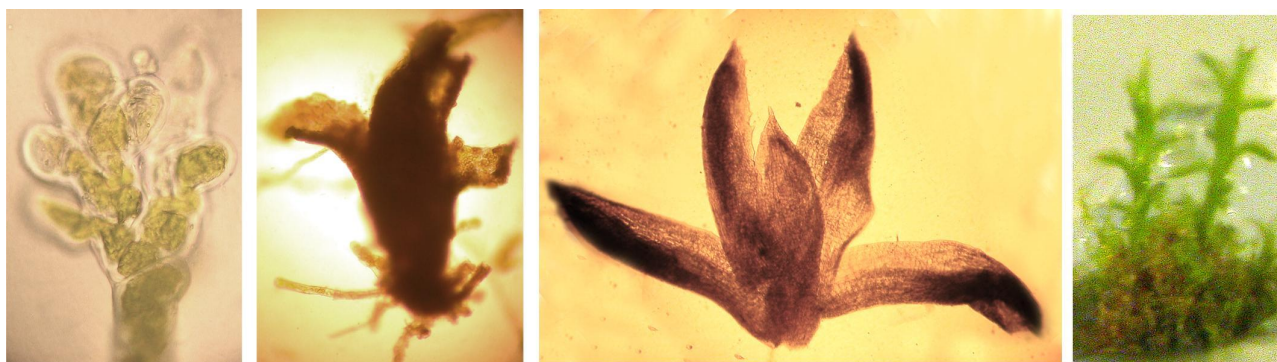


Fig. 3. Stages during development of a bud into a plantlet in axenic culture of *Pogonatum urnigerum*.

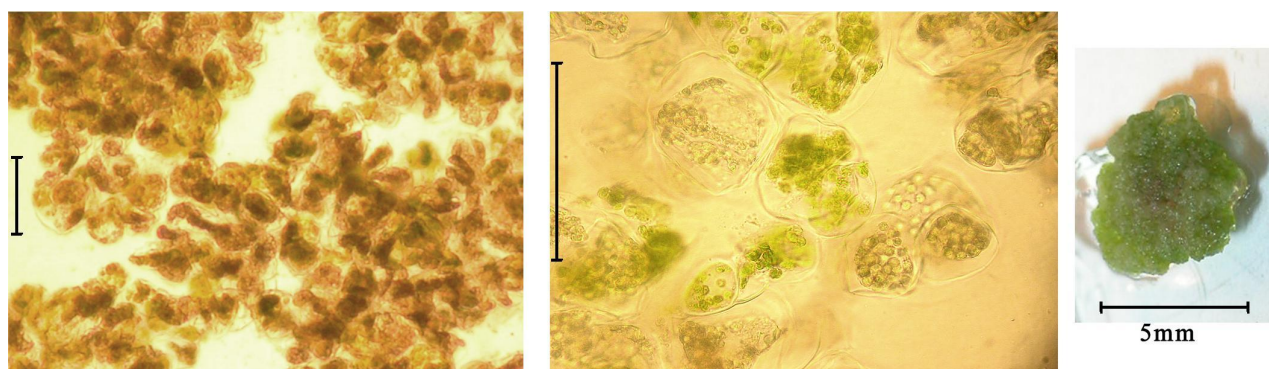


Fig. 4. Developed callus tissue after 120 days of subcultivation on MS/2 nutrient medium. The bar represents 50  $\mu\text{m}$ .



Fig. 5. Effect of hormone free MS/2 medium (A), MS/2 medium containing 1  $\mu\text{M}$  IAA (B), and MS/2 medium containing 1  $\mu\text{M}$  BA (C) on *in vitro* development of *Pogonatum urnigerum* protonemata.

## DISCUSSION

The formerly accepted protocol for bryophyte introduction into axenic culture developed in our laboratory (Sabovljević et al., 2002, 2003) was also successful in the case of *Pogonatum urnigerum*. Hormone-free

media with half-strength MS mineral salts and vitamins allowed full gametophyte development. Despite extensive bud formation, only a small fraction of buds developed into gametophores, indicating that other factors such as light regime, temperature, medium pH, and osmolarity should be varied in order to activate more exten-

sive gametophore development. Ours is the first report of callus formation in *Pogonatum urnigerum*, although this phenomenon has so far been reported for 18 other moss species (Felix, 1994).

Callus formation and relatively fast senescence of protonemata are probably due to the fact that the protonemata of this species are not persistent in nature. In keeping with the habitat of this species, growing in liquid medium resulted in somewhat faster growth and delayed senescence (data not presented).

Addition of 1.5% sucrose to the media positively affected gametophyte development in *P. urnigerum*, which was also observed in *Bryum argenteum* (Sabovljević et al., 2005) and another polytrichaceous moss, *Atrichum undulatum* (Sabovljević et al., 2006b). However, the level of sucrose influence on morphogenesis of different bryophyte species was not the same: in *B. argenteum* *in vitro* culture, a quite high index of multiplication was recorded with addition of 1.5% sucrose to the medium; in *A. undulatum*, on the other hand, normal gametophyte development was induced, as in the related polytrichaceous species *P. urnigerum*.

Of the five main groups of phytohormones, only auxins and cytokinins are documented as natural signal substances in mosses (Cove and Ashton, 1984; Bopp and Bhatla, 1985). Both of these hormone groups not only exist in mosses, but also have basic functions in the regulation of normal development. The known effects of auxins on moss development include inhibition of protonema growth, transformation of buds to filaments, torsion of young stems and complete suppression of leaves on gametophores (Bopp, 1953; Sokal et al., 1997). Cytokinins have been shown to induce bud formation in protonemata cultures of some moss species (Speiss, 1975, 1976; Takio, 1989; Christianson and Hornbuckle, 1999; Bijelović and Sabovljević, 2003). In our study, sterile spores germinated and formed protonemata that were able to produce buds without exogenous cytokinin supply. However, as in the case of *Physcomitrella* (Ashton et al., 1979), cytokinin treated buds did not develop into gametophores. Rapid senescence of tissue grown on 1 µM BA might indicate high endogenous levels of cytokinins, senescence being the result of supraoptimal concentrations caused by exogenous cytokinin supply. When auxin was applied, some gametophores but no rhizoids were formed, opposite to what was observed in *Physcomitrella* (Ashton et al., 1979). In studies of cytokinin

action on different moss species (Speiss, 1976), calli were obtained with most of the species, while the *Polytrichaceae* studied did not form calli. *Atrichum undulatum* formed calli when the growth medium contained 4% glucose and 0.2-2 mg/L benzyladenine (Gang et al., 2003), and Ono et al. (1988) reported that high sugar levels and high medium osmolarity were essential for callus formation. In our study, callus was obtained at low sugar and osmolarity values, again indicating that endogenous factors determine the development pattern, these factors being species-specific and probably of adaptive rather than evolutionary origin.

*Acknowledgement* – This work was supported financially by the Ministry of Science and Environment Protection of Serbia.

## REFERENCES

- Ashton, N.W., Grimsley, N.H., and Cove, D.J. (1979). Analysis of gametophytic development in the moss *Physcomitrella patens* using auxin and cytokinin resistant mutants. *Planta* **144**, 427-435.
- Bijelović, A., and Sabovljević, M. (2003). Callus induction and plant regeneration in the moss *Aloina aloides* (Schultz.) Kindb. (*Pottiaceae*, *Bryopsida*). *Archives of Biological Sciences*, Belgrade **55** (3-4), 77-80.
- Bijelović, A., Sabovljević, M., Grubišić, D., and Konjević, R. (2004). Phytohormone influence on the morphogenesis of two mosses [*Bryum argenteum* Hedw. and *Atrichum undulatum* (Hedw.) Beauv.]. *Israel J. Plant Sci.* **52**, 31-36.
- Bopp, M. (1953). Die Wirkung von Heteroauxin auf Protonemawachstum und Knospen-bildung von *Funaria hygrometrica*. *Z. Bot.* **41**, 1-16.
- Bopp, M., and Bhatla, S. C. (1985). Hormonal regulation of development in mosses. In: *Hormonal Regulation of Plant Growth and Development*, 2 (Ed. S. S. Purohit), p. 65. Agro-Botanical Publishers, Old Ginanni, Bikaner.
- Christianson, M. L., and Hornbuckle, J. S. (1999). Phenyl urea cytokinins assayed for induction of shoot buds in the moss *Funaria hygrometrica*. *Am. J. Bot.* **86**, 1645-1648.
- Cove, D. J., and Ashton, N. W. (1984). The hormonal regulation of gametophytic development in bryophytes. In: *The Experimental Biology of Bryophytes*, (Eds. Dyer, A. F., and Duckett, J. G.), p. 177. Academic Press, London.
- Cove, D. J., Bezanilla, M., Harries, P., and Quatrano, R. (2006). Mosses as model systems for the study of metabolism and development. *Annu. Rev. Plant Biol.* **57**, 497-520.
- Felix, H. (1994). Calli, cell and plantlet suspension cultures of bryophytes. *Candollea* **49**, 141-158.
- Gang, Y-Y, Du, G-S., Shi, D-J., Wang, M-Z., Li, X-D., and Hua Z-L. (2003). Establishment of *in vitro* regeneration systems of the *Atrichum* mosses. *Acta Botanica Sinica* **45**, 1475-1480.
- Mues, R. (2000). Chemical constituents and biochemistry. In: *Bryophyte*

- Biology*, (Eds. Shaw A.J. and Goffinet B.), 150-181. Cambridge University Press.
- Murashige, T., and Skoog, F. (1962). A revised medium for rapid growth and bioassays with tobacco tissue culture. *Physiologia Plantarum* **15**, 473-497.
- Ono, K., Murasaki, Y., and Takamiya, M. (1998). Induction and morphogenesis of cultured cells of bryophytes. *J. Hattori Bot. Lab.* **65**, 391-401.
- Sabovljević, M., Bijelović, A., and Grubišić, D. (2001). Bryophytes as a potential source of medicinal compounds. *Lekovite Sirovine* **21**, 17 – 29.
- Sabovljević, M., Bijelović, A., and Dragičević, I. (2002). Effective and easy way of establishing *in vitro* culture of the mosses *Bryum argenteum* Hedw. and *Bryum capillare* Hedw. (*Bryaceae*). *Arch. Biol. Sci. Belgrade* **54**, 7P-8P.
- Sabovljević, M., Bijelović, A., and Dragičević, I. (2003). *In vitro* culture of mosses: *Aloina aloides* (K.F. Schultz) Kindb., *Brachythecium velutinum* (Hedw.) B.S.&G., *Ceratodon purpureus* (Hedw.) Brid., *Eurhynchium praelongum* (Hedw.) B.S.&G. and *Grimmia pulvinata* (Hedw.) Sm. *Turk. J. Bot.* **27**, 441-446.
- Sabovljević, A., Sabovljević, M., Grubišić, D., and Konjević, R. (2005). The effect of sugars on development of two moss species (*Bryum argenteum* and *Atrichum undulatum*) during *in vitro* culture. *Belg. Journ. Bot.* **138**, 79-84.
- Sabovljević, A., Soković, M., Sabovljević, M., and Grubišić, D. (2006a). Antimicrobial activity of *Bryum argenteum*. *Fitoterapia* **77**, 144-145.
- Sabovljević, A., Cvetić, T., and Sabovljević, M. (2006b) Establishment and development of the Chaterine's moss *Atrichum undulatum* (Hedw.) P. Beauv. (*Polytrichaceae*), in *in vitro* conditions. *Archives of Biological Science Belgrade* **58** (2), 87-93.
- Sokal, I., Kuta, E. and Przywara, L. (1997). Callus induction and gametophyte regeneration in moss cultures. *Acta. Biol. Cracov. Ser. Bot.* **39**, 35-42.
- Speiss, L.D. (1975). Comparative activity of isomers of zeatin and ribosyl-zeatin on *Funaria hygrometrica*. *Plant Physiol* **55**, 583-585.
- Speiss, L.D. (1976). Developmental effects of zeatin, ribosyl-zeatin, and *Agrobacterium tumefaciens* B<sub>6</sub> on certain mosses. *Plant Physiol.* **58**, 107-109.
- Takio, S. (1989). Effect of cytokinin on morphological changes of suspension cultured cells of the moss *Barbula unguiculata*. *Plant Cell Reports* **7**, 603-606.

## РАЗВИЋЕ МАХОВИНЕ *POGONATUM URNIGERUM* (HEDW.) P. BEAUV. ГАЈЕНЕ У УСЛОВИМА КУЛТУРЕ *IN VITRO*

ТИЈАНА ЦВЕТИЋ<sup>1</sup>, АНЕТА САБОВЉЕВИЋ<sup>1</sup>, М. САБОВЉЕВИЋ<sup>1</sup> И Д. ГРУБИШИЋ<sup>1,2</sup>

<sup>1</sup> Институт за ботанику, Биолошки факултет Универзитета у Београду, 11000 Београд, Србија

<sup>2</sup> Институт за биолошка истраживања "Синиша Станковић", 11000 Београд, Србија

Стерилна култура маховине *Pogonatum urnigerum* (*Polytrichaceae*) успостављена је из спора сакупљених у природи. Гајењем на хранљивој подлози без биљних хормона из спора су се развиле протонеме и гаметофити. Успостављена је и стабилна култура калуса. Установљено је да је за гајење ове врсте најбоља хранљива подлога која садржи смањену концентрацију соли и витамина по Murashige-Skoog-у и

1.5% сахарозу. Третман ауксином није поспешило развиће гаметофита, док је продужени третман довео до постепеног одумирања протонеме. Ткива гајена на подлози која је садржала цитокидине уопште нису продукovala гаметофите и нису преживела продужени третман овим хормоном.

Овај рад представља прву студију развића врсте *Pogonatum urnigerum* у *in vitro* култури.