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Influence of NG-nitro-L-arginine methyl ester on clinical and biochemical effects of methylene blue in pentylenetetrazole-evoked convulsions

Uticaj NG-nitro-L-arginin metil estra na kliničke i biohemijske efekte metilen plavog kod konvulzija izazvanih pentilentetrazolom

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Abstract

Background/Aim. Despite years of research in a number of experimental models the question whether nitric oxide (NO) and methylene blue (MB) have pro- or anticonvulsant effects remains to be fully resolved. Methods. In adult Wistar rats the influence of a nonselective inhibitor of nitric oxide synthase NG-nitro-L-arginine methyl ester (L-NAME, 10 µg) on clinical and biochemical effects of MB (10 µg) given before the intraperitoneally administered chemical convulsant pentylenetetrazole (PTZ, 80 mg/kg) was examined. MB and L-NAME were applied intracerebroventricularly. PTZ application was followed by a 4minute observation time, after which rats were sacrificed and elements of oxido-reductive balance were measured in a crude mitochondrial fraction of forebrain cortex, hippocampus and striatum. Results. Convulsive responses (forelimb dystonia - FLD, generalised clonic- and clonic-tonic convulsions - GCC and GCTC respectively) were observed in all rats received PTZ, together with significantly decreased lipid peroxidation in the forebrain cortex and striatum and increased superoxide dismutase activity in the hippocampus, in comparison to controls (saline treated). It was

Apstrakt

Uvod/Cilj. I pored višegodišnjeg istraživanja na različitim eksperimentalnim modelima, nije potpuno odgovoreno na pitanje da li azot-oksid (NO) i metilen plavo (MP) deluju konvulzivno ili antikonvulzivno. **Metode.** Na odraslim pacovima Vistar soja ispitivan je uticaj NG-nitro-L-arginin metil estra (L-NAME, 10 μ g), neselektivnog inhibitora azot oksid sintaze, na kliničke i biohemijske efekte metilen plavog (MP, 10 μ g) datog intracerebroventrikularno pre hemijskog konvulziva pentilentetrazola (PTZ, 80 mg/kg), primenjenog intraperitonealno. Pacovi su posmatrani četiri

registered anticonvulsant effects of L-NAME pretreatment. However, these effects were insignificant. In the hippocampus of these animals there was decreased lipid peroxidation (p < 0.01, p < 0.05 vs saline-treated and PTZ-treated rats, respectively) and reverted PTZ-induced increase of superoxide dismutase activity. But MB individually pretreatment significantly decreased the incidence of CTCs and GCCs (FLD: p = 0.0513), prolonged the convulsive latent time for FLD, GCTCs and GCCs, in all the examined brain regions increased lipid peroxidation and decreased the level of superoxide anion. Administration of L-NAME 10 minutes before MB reverted all MB-evoked clinical and biochemical effects. Conclusion. Methylene blue applied individually before PTZ has strong anticonvulsant effects that were eliminated by L-NAME pretreatment. These effects and changed biochemical parameters in the brains of animals treated by L-NAME before MB in comparison to MBtreated group suggest involvement of NO in MB's effects in the animal model of PTZ-evoked convulsions.

Key words: seizures; nitric oxide; methylene blue; pentylenetetrazole; rats; oxidoreductases.

minuta posle davanja PTZ-a, posle čega su žrtvovani i u neprečišćenoj mitohondrijskoj frakciji prednjeg mozga, hipokampusa i strijatuma određivani su parametri oksidoreduktivne ravnoteže. **Rezultati.** Posle primene PTZ-a, konvulzivni odgovor (distonija prednjih nogu – DPN, generalizovane klonične – GKK i generalizovane kloničnotonične konvulzije – GKTK) bio je ispoljen kod svih životinja, kao i statistički značajno sniženje lipidne peroksidacije u kori prednjeg mozga i strijatuma, i povećanje aktivnosti superoksid dizmutaze (SOD) u hipokampusu, u poređenju sa kontrolnom grupom (dobila fiziološki rastvor NaCl). Registrovani su antikonvulzivni efekti L-NAME

Correspondence to: Ankica Jelenković, Institute for Biological Research "Siniša Stanković", Bulevar Despota Stefana, 142, 11 000 Belgrade, Serbia. Phone/fax: +381 11 342 0606. E-mali: <u>jelaka@yahoo.com</u> koji nisu bili statistički značajni. U hipokampusu ovih životinja bila je snižena lipidna peroksidacija (p < 0,01 u poređenju sa kontrolnom grupom, p < 0,05 u poređenju sa životinjama koje su dobile PTZ), kao i aktivnost SOD u poređenju sa životinjama koje su dobile PTZ. Samo metilen plavo dovelo je do statistički značajnog smanjenja incidencije GKK I GKTK (DPN: p = 0,0513), produžilo je latentni period DPN, GKK i GKTK, a u svim ispitivanim strukturama mozga bila je povećana lipidna peroksidacija i smanjen nivo superoksidnog anjona. Svi klinički i biohemijski efekti izazvani primenom MP u potpunosti su odstranjeni primenom L-NAME 10 minuta pre davanja MP. **Zaključak.** Metilen plavo, dat samostalno pre PTZ, ispoljio je snažne antikonvulzivne efekte. Nestanak ovih efekata i izmenjeni biohemijski parametri u mozgovima pacova koji su pre MP dobili L-NAME, sugerišu da je NO uključen u efekte MP ispoljene na životinjskom modelu konvulzija izazvanih primenom PTZ-a.

Ključne reči:

konvulzije; azot, oksid; metilensko plavilo; pentilentetrazol; pacovi; oksidoredukcija.

Introduction

An accumulated body of evidence supports multiple physiological as well as pathological roles for nitric oxide (NO) in its free radical form. Acting both presynaptically and postsynaptically NO accomplishes its complex participation in a wide range of physiological and pathophysiological phenomena including regulation of vascular tone, inflammation and signalling in the central nervous system (CNS) *via* polysynaptic interacting circuits ^{1, 2}.

Nitric oxide is synthesised from L-arginine by nitric oxide synthases (NOS) in response to N-methyl-D-aspartate (NMDA) and non-NMDA receptor activation, and it is inhibited by a number of NOS inhibitors, among which is NGnitro-L-arginine methyl ester (L-NAME). Alternatively, it can be released from NO donors. Besides hemoglobin, NOS and other metaloenzymes which are targets for NO binding, the interaction between NO and heme moiety within the soluble (cytosolic) form of guanylate cyclase (sCG) has particular physiological significance. The consequence of NO binding to sGC is the activation of the latter resulting in an elevation of cyclic guanosine 3',5'-monophosphate (cGMP) and the initiation of a cascade of target cell-specific events³. Although much effort has been made to elucidate how NO interacts and stimulates sGC the precise mechanism is still unclear to date.

Despite years of research, the role of NO in the pathogenesis of epilepsy and convulsions still remains controversial. NO can provoke convulsions but also it can exhibit anticonvulsant effects ^{4–6}. In addition, some convulsive patterns are NO-independent ^{7, 8}. Such unbalanced reports maybe reflected by the use of different experimental conditions, involving not only different animal species and strains, but also other factors including age, sex, convulsive models, the form, the route and doses of the applied substances.

During convulsions the concentration of cGMP increases in specific brain regions. Within the first four minutes of pentylenetetrazole (PTZ) application, which evokes generalised clonic convulsions (GCCs) and generalised clonic-tonic convulsions (GCTcs) in almost all mice, increases in cGMP (3- to 5-fold) were found primarily in the hippocampus and cerebral cortex but also (to a lesser extent) in the cerebellum ⁹. Within the first minute of PTZ application a rapid and very high level of cGMP was found in ventilated and non-ventilated guinea pigs in all the above-mentioned brain structures and also in the striatum, suggesting that increased cGMP is a pathogenic component of PTZ-evoked convulsions. The above mentioned brain structures are most important in convulsions initiation and propagation.

Methylene blue (MB), a highly active redox compound that readily cycles between oxidised (methylene blue) and reduced (leukomethylene blue) states, is a blue-coloured organic dye. Besides in experimental conditions, MB has importance in human clinical practice too, although data about its effects in humans are not uniform ^{10, 11}.

The effects of MB on cGMP and on NO have already been documented in several studies. Since MB is a nonselective and a weak incomplete inhibitor of sGC, this effect of MB on sGC has been widely studied ^{12, 13}. MB can attenuate cGMP accumulation by inhibiting NO-stimulated cGC without an effect on the basal cGMP level ¹⁴. Masaki and Kondo ¹⁵ intracerebrovetricularly administered MB to rats 30 minutes prior to sacrificing them. In whole brain homogenate the level of cGMP decreased in a MB dose-dependent manner. MB administration can also decrease the NOS activity and NO content in certain brain regions and also prevents experimental seizures ^{16, 17}.

The NO-sGC-cGMP signalling pathway is present in virtually all cells but sGC is expressed at particularly high levels within neurons. This pathway is largely influenced by glutamatergic neurotransmission that is modulated in response to PTZ treatment ¹⁸.

According to a number of data, as well as to our own results, it is obvious that elements of oxidative/antioxidative balance are changed in the brains of rats with convulsive response to PTZ and upon L-NAME treatment in such kind of experimental convulsions^{19, 20}. Because of these facts but also due to MB's ability as well as L-NAME and PTZ to interfere with synthesis/actions of NO, and due to their convulsive/anticonvulsive properties, we sought to determine their influence on both clinical and brain biochemical changes (parameters of oxidative defence and stress) in a PTZ seizure model in rats^{5–7, 12, 16, 17}.

Methods

Animals and surgery

Experiments were performed on 13-week old male Wistar rats housed in a temperature-controlled room $(23 \pm 2^{\circ}C)$ with 11-hour light/13-hour dark cycles with free access to food and water. Experiments were conducted within a period of two weeks always between the hours of 9:00 am and 2:00 pm. Animals were randomly assigned to different drug treatment regimes. All protocols for handling the rats were approved by the local ethical committee for the use of experimental animals (Military Medical Academy, Belgrade, Serbia).

For intracerebroventricular (*icv*) application of drugs a polyethylene cannula was stereotaxically implanted into the left lateral ventricle (coordinates: 1.3 mm behind the bregma, 1.8 mm left from the midline suture, 3.7 mm ventral from the durra ²¹) under sodium pentobarbital intraperitoneally (*ip*) anaesthesia (45 mg/kg body weight, Vetanarcol[®], Werfft-Chemie, Vienna). The cannula was fixed to the skull with dental cement and two jeweller screws. The postoperative recovery period lasted six days before experiments were continued.

Treatments

Rats were divided into five groups (Figure 1; in each group there were 7–8 rats). PTZ (Sigma Chemical Co., St

observation following PTZ injection was limited to 4-min. During that time convulsive responses were monitored and were graded according to the following scale: fore limb dystonia (FLD), generalised clonic seizures (clonus of the whole body with a loss of righting reflex, GCCs) and generalised clonic-tonic convulsions with tonic hind limb extension (GCTCs). The appearance (incidence) and the time to onset of every convulsive pattern were registered. If the rats did not exhibit convulsive responses, a latent time of 240 sec was assigned.

Biochemical parameters

All the rats were sacrificed by decapitation 4 min after PTZ administration. Their heads were immediately frozen in liquid nitrogen and stored at -70°C.

For biochemical analysis the forebrain cortex, striatum and hippocampus from the contralateral hemisphere relative to cannula insertion were dissected on ice, homogenized in icecold buffer, centrifuged and sonicated. Biochemical parameters were determined spectrophotometrically in the crude mitochondrial fraction according to the method of Gurd et al.²².



Fig. 1 – Protocol of treatments (n = 7–8 Wistar rats in each group)

NaCl – 0.9% saline; PTZ – pentylenetetrazole; L-NAME – NG-nitro-L-arginine methyl ester; MB – methylene blue; *ip* – intraperitoneally; *icv* – intracerebroventricularly

Louis) was used as a chemoconvulsant. It was applied *ip* in a single dose of 80 mg/kg body weight. The control group received sterile isotonic saline *icv* twice: 30 minutes and 20 minutes before *ip* application of the same solution. Another group of rats also received sterile isotonic saline *icv* twice: 30 and 20 minutes before *ip* application of PTZ (PTZ-treated group). The third group received 10 μ g NG-nitro-L-arginine methyl ester (L-NAME, Sigma Chemical Co.) *icv* 30 minutes before PTZ. Ten minutes after the L-NAME injection the rats also received sterile isotonic saline *icv*.

MB (10 μ g) was administered *icv* to another two groups of rats 10 minutes after sterile isotonic saline (*icv*) or L-NAME (10 μ g, *icv*)¹⁵. Twenty minutes after MB (30 minutes after sterile isotonic saline/L-NAME treatment), both groups of rats received PTZ. All drugs were dissolved in sterile isotonic saline. Volume of *ip* and *icv* applied solutions were 1 mL/kg body weight and 10 μ L, respectively.

Behavioural evaluation

Immediately after PTZ administration the rats were placed individually in transparent perspex cages. The time of

The assay for the superoxide radical is based on the reduction of nitrobluetetrazolium (Merck) in an alkaline, nitrogen-saturated buffer with absorbance monitoring at 515 nm^{23} .

Total superoxide dismutase (SOD) activity was measured as the inhibition of epinephrine auto-oxidation (Sigma Chemical Co.) in sodium carbonate buffer (Serva) containing 0.1 mM EDTA (Sigma Chemical Co.) at 480 nm 24 .

Lipid peroxidation was measured as a function of malondialdehyde (MDA) production at 533 nm²⁵.

The incidence of convulsive patterns was expressed as the percentage of convulsing rats of the total number of rats in a group and was analysed using Kolmogorov-Smirnov test. The latent time of convulsions was calculated in seconds and expressed as the mean \pm standard deviation (SD) and was analysed using the Kruskal-Wallis test. Biochemical parameters were expressed as the mean \pm SD and were analysed using the ANOVA, followed by Tuckey test. Differences between experimental groups were considered significant when p < 0.05.

Results

Behavioural effects

In the control group of rats (sterile isotonic saline treatment, *icv* and *ip*) no convulsions were observed. In contrast, convulsive responses were observed in all PTZ-treated rats (Figure 2).







Pretreatment with L-NAME led to some anticonvulsant, however insignificant effects on PTZ-induced convulsions.

MB pretreatment was very effective against PTZevoked convulsions. GCTCs were completely prevented (p < 0.01, compared to the PTZ-treated group). GCCs appeared only in 25% of animals (p < 0.01). FLD was apparent in 50% of the rats (p = 0.0513).

When L-NAME was administered before MB, the anticonvulsant effects of MB were lost, in particular GCTCs and GCCs; their incidence increased from 0 to 83%; (p < 0.01), and from 25 to 83% (p < 0.05), respectively. In addition the incidence of FLD was doubled, from 50 to 100% (p = 0.0513).

The latent time of PTZ-evoked convulsions was not influenced by L-NAME pretreatment (Table 1). MB very strongly influenced the latent time of all three convulsive patterns (p < 0.01 for FLD and GCTC, p < 0.05 for GCC, compared with the PTZ-treated group and groups pretreated with L-NAME). But MB's ability to extend the latent time of PTZ-induced convolutions was completely blocked by L-NAME.

Biochemical effects

In the PTZ-treated group of rats decreased lipid peroxidation in the forebrain cortex and striatum compared with the control group was found (p < 0.05 and p < 0.01, respectively) (Figure 3). In addition, increased SOD activity in the hippocampus compared with the control group was noted (p < 0.05) (Figure 4).







NAME 10 μg) and methylene blue (MB, 10 μg) on the superoxide dismutase (SOD) activity in the brain of Wistar rats (n = 7–8) treated with pentylenetetrazole (PTZ (80 mg/kg)

p < 0.05, 0.01 is the level of significance when compared with the corresponding values (mean ± SD) of 0.9% NaCl (*, **) and PTZ-treated rats (♦, ♦♦; ANOVA). PTZ was applied intraperitoneally, NaCl intraperitoneally and intracerebroventricularly, L-NAME and MB intracerebroventricularly

Table 1

Time to onset of n	entvlenetetrazole (PTZ	80 mg/kg) – evoked	convulsions in Wista	• rats (n = 7_8)
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Treatment		Latent time (seconds)	
L-NAME and MB (µg)	FLD	GCC	GCTC
NaCl + PTZ	49.5 ± 11.3♦♦	118.6 ± 25.8♦	131.7 ± 35.6♦♦
L-NAME $(10) + PTZ$	57.7 ± 11.9♦♦	125 ± 68.3♦	$128 \pm 67 \bigstar$
MB(10) + PTZ	$159.2 \pm 88.7 **$	$213.3 \pm 65.3*$	$240 \pm 0^{**}$
L-NAME (10) + MB (10) + PTZ	53.3 ± 8.8♦♦	123.5 ± 70.4♦	126.7 ± 70♦♦

p < 0.05, 0.01 is the level of significance when compared with the corresponding PTZ (*, **) and MB+PTZ treatments (\bullet , $\bullet \bullet$; Kruskal-Wallis test). Values are expressed as means ± SD. In the case without convulsive response, the latent time was assigned as 240 seconds. PTZ was applied intraperitoneally, 0.9% NaCl, L-NAME and MB intracerebroventricularly

FLD – forelimb dystonia; GCC – generalised clonic convulsions; GCTC – generalised clonic-tonic convulsions; L-NAME – NG-nitro-Larginine methyl ester; MB – methylen blue

Treatment with L-NAME prior to PTZ administration decreased lipid peroxidation in the hippocampus (p < 0.01, compared with the control group; p < 0.05, compared with the PTZ-treated group) (Figure 3). SOD activity was decreased in the forebrain cortex (p < 0.05, compared with the control group) (Figure 4). Superoxide anion content of the hippocampus was also decreased (p < 0.05, compared with the control group) (Figure 5).



Fig. 5 – Influence of NG-nitro-L-arginine methyl ester (L-NAME 10 μg) and methylene blue (MB, 10 μg) on the superoxide anion content of the brain of Wistar rats (n = 7–8) treated with pentylenetetrazole (PTZ, 80 mg/kg). p < 0.05, 0.01 is the level of significance when compared with the corresponding values (mean ± SD) of 0.9% NaCl (*, ***) and PTZ-treated rats (•, ••; ANOVA). PTZ was applied intraperitoneally, NaCl intraperitoneally and intracerebroventricularly. L-NAME and MB intracerebroventricularly</p>

MB administered before PTZ resulted in increased lipid peroxidation compared with the control group (p < 0.05 in the forebrain cortex, p < 0.01 both in the striatum and hippocampus) and compared with the PTZ-treated group of rats (p < 0.01 for all brain regions) (Figure 3).

The level of superoxide anion was decreased in the MB+PTZ treated group of rats compared with both the control and PTZ-treated groups (p < 0.01 in all brain regions) (Figure 5).

Pretreatment with L-NAME reverted the MB+PTZinduced increase in lipid peroxidation (Figure 3) and decrease in superoxide anion (Figure 5) in all brain regions back to the control values, except in the forebrain cortex where the superoxide content was increased, while SOD activity was decreased (p < 0.05 in comparison with the control and PTZ-treated groups of rats, and the control group, respectively).

Discussion

We found that both L-NAME and MB affected PTZevoked convulsions (insignificant and almost completely/completely prevented, respectively). However, coadministration of L-NAME and MB did not result in an additive/synergistic effects. Instead the effects (clinical and biochemical) of MB were severely abrogated by L-NAME.

The convulsant effects of PTZ are quite complex and are not yet completely understood. Reduced γ -aminobutyric (GABA) activity (GABA suppresses NOS), and enhanced excitatory amino acids release/transmission is believed to be

the underlying mechanism ^{26, 27}. Thus, PTZ's effects on both the GABA-ergic and glutamatergic system lead to overproduction of NO and potentiation of sGC activity.

NO preferentially reacts with other radicals despite being a free radical itself. Its reactivity with biological molecules is low but it readily reacts with the hydroxyl radical ²⁸. In this way it can act as an antioxidant by scavenging free radicals thereby inhibiting lipid peroxidation. Chiueh 29 proposed at least four mechanisms by which NO acts as an antioxidant, including inhibition of lipid peroxidation. Thus, within the first few minutes of PTZ administration when cGMP production is rapidly increased 9, which is an indirect measure of NO production, a defence reaction against PTZ's toxic effects (via the antioxidative activity of NO) may take place. Under very strong production of NO and at later times this antioxidative defence is overcome and lipid peroxidation ensues, as described by a number of studies. For example, Patsoukis et al.^{30, 31} found that the intensity of lipid peroxidation was increased in the mouse hippocampus and the striatum 15 min after PTZ was applied ip at a dose of 60 mg/kg. Furthermore, increased lipid peroxidation was within control values 30 min and 24 h post PTZ administration. However, lipid peroxidation was not increased in the mouse cerebral cortex in each monitoring time ³². A lower dose of PTZ (40 mg/kg) did not in any way influence lipid peroxidation, meaning that lipid peroxidatrion is PTZ dose- and time- dependent.

In the research of Bashkatova et al. ³³ PTZ (120 mg/kg administered subcutaneously) caused convulsions in all animals within 2–3 minutes. Moreover, one hour later the level of NO was increased 5-fold and lipid peroxidation 2-fold in the frontal cortex. Despite the fact that PTZ was used as a chemoconvulsant in both our and the above-mentioned studies, to some degree different results concerning lipid peroxidation are most likely explainable due to different experimental procedures.

The anticonvulsant effects of MB described in our studies are in accordance with those of Furian et al. ¹⁷. In addition to MB's ability to increase the latency of methylmalonate-induced convulsions in adult male Wistar rats, abrogation of the methylmalonate-induced striatal NO level elevation was also observed. Furthermore, in the case of catalepsy induction Echeverry et al. ³⁴ demonstrated similar effects of L-NAME and MB, both applied *icv*. Also, the level of NO products was decreased in the striatum.

However, discrepancies between our results and the results obtained by Deutsch et al. ³⁵, who did not record the anticonvulsant effects of MB in electrically precipitated tonic hind limb extension in mice, could be explained by the use of different experimental models of convulsions, routes of drug administration, time points and dose of MB application, since pharmacokinetics and organ distribution of MB depends on the way of its application³⁶.

Apart from inhibition of sGC, MB has several other nonspecific effects on NO including inhibition of NOS. In cultured endothelial cells Shimizu et al. ³⁷ found that MB, which is known to inactivate iron-containing enzymes, reacts with the heme moiety of NOS. Vallo et al. ¹⁶ found similar effects on hippocampal brain NOS activity *in vivo*. Further-

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more, MB-mediated inhibition of hippocampal NOS was dose-dependent and the degree of inhibition was similar to that obtained with an unselective NOS inhibitor N-G-nitro-L-arginine (L-NNA). In comparison with sGC inhibition, MB-mediated NOS inhibition appeared more potent ¹². The susceptibility of different NOS isoforms to MB remains unclear. Because of NOS inhibition by MB, in our research it was expected that L-NAME, a competitive non-selective NOS inhibitor, enhances MB's anticonvulsant effects. However, the opposite was apparent: MB's anti-convulsant effects were reverted back to control values. In other words, combined pretreatment with L-NAME and MB exerted deleterious effects on MB-preventable PTZ-evoked convulsions. It is difficult to propose the reason for the obtained events. In addition to nitrergic system a number of other effects of both substances have to be taken into consideration before any conclusion, such as their influence on cholinergic, dopaminergic, serotoninergic and other transmissions that are important in the pathogenesis of convulsions ³⁸.

Despite much more evidence pointing towards its antioxidant effects, MB can also be prooxidative causing singlet oxygen and superoxide formation, weak peroxide generation and direct glutathione oxidation ³⁹. Inactivation of NO by superoxide is ranked as one of the most rapid nonenzymatic reactions in biology. In other words one of MB effect could be related to the removal of NO already present. The chemical reaction between NO and superoxide, which could be generated in the presence of MB, may lead to the formation of peroxynitrite anion. The latter is a source of the hydroxyl radical which can intensify lipid peroxidation ⁴⁰. In our previous study it was found increased NO leveles in the hippocampus, forebrain cortex, striatum and some other brains structures within 30 minutes of PTZ application in the PTZevoked convulsions in rats 41. Therefore, abovementioned effects of PTZ are in agreement with our observation regarding increased lipid peroxidation and a decreased level of superoxide anion in all the examined brain structures isolated from rats pre-treated with MB. Reestablishing level of lipid peroxidation in all brain regions and superoxide anion (except in the forebrain cortex) without increased SOD activity when L-NAME was coadministered with MB coincide with the suggested explanation of increased lipid peroxidation upon MB treatment ie it could indicate decreased interaction between NO and superoxide anion. However, increased lipid peroxidation that was associated with anticonvulsant effects of MB is undesired for any antiepileptic drug and substance and needs scientific evidence reliable of benefit/risk ratio with the aim to study whether benefits of such treatments (anticonvulsant effects) overwhelm risks of their potential harmful effects (increased lipid peroxidation).

A cascade of prooxidative events associated with MB pretreatment was not at all disrupt by coadministration of L-NAME. That was demonstrated in increased level of superoxide anion in the forebrain cortex. It could be the result of L-NAME effects on SOD activity, which was decreased in this structure when L-NAME was individually applied before PTZ. Thus, L-NAME did not only prevent anticonvulsant and lipid peroxidation effects of MB, but also it made conditions for forebrain cortex oxidative damage. In other words, all three structures in which most of the complex events of epileptic seizures are generated were very sensitive and prone to PTZ, MB and L-NAME influence.

In our PTZ-mediated convulsing rat model it was registered very early and rapid events (within the first 4 minutes after PTZ application): MB, when applied individually, prevented convulsive responses, especially GCTCs and GCCs; modulation of the NO system by MB in the settings of PTZinduced convulsions may elicit anticonvulsant effects, but only if MB was applied on its own, not in combination with L-NAME, indicating that NO plays important role into anticonvulsant effects of MB under applied conditions; convulsive responses to PTZ was associated with a decreased and/or unchanged level of lipid peroxidation in the forebrain cortex, striatum and hippocampus. In contrast, anticonvulsant effects of MB coincide with increased lipid peroxidation. These clinical and biochemical effects of MB were abandon by L-NAME treatment.

Conclusion

MB's strong anticonvulsant effects in PTZ-evoked convulsions, especially against generalised clonic and clonictonic convulsions, and changes of the examined biochemical parameters in brain structures, as well as prevention of all these effects by L-NAME applied before MB could be partly the result of the nitrergic system modulation. Our results have far reaching therapeutic implications, *ie* translation into clinical practice benefits, particularly the anticonvulsant effects of MB. Future investigations will hopefully shed some light on the intricacies of MB's effects.

REFERENCES

- Vallance P, Moncada S. Nitric oxide--from mediator to medicines. J R Coll Physicians Lond 1994; 28(3): 209–19.
- Steinert JR, Chernova T, Forsythe ID. Nitric oxide signaling in brain function, dysfunction, and dementia. Neuroscientist 2010; 16(4): 435–52.
- Bredt DS, Snyder SH. Nitric oxide mediates glutamate-linked enhancement of cGMP levels in the cerebellum. Proc Natl Acad Sci U S A 1989; 86(22): 9030–3.
- Mollace V, Bagetta G, Nisticò G. Evidence that L-arginine possesses proconvulsant effects mediated through nitric oxide. Neuroreport 1991; 2(5): 269–72.
- Jelenković A, Jovanović M, Ninković M, Maksimović M, Bokonjić D, Bosković B. Nitric oxide (NO) and convulsions induced by pentylenetetrazol. Ann N Y Acad Sci 2002; 962: 296-305.
- Buisson A, Lakhmeche N, Verrecchia C, Plotkine M, Boulu RG. Nitric oxide: an endogenous anticonvulsant substance. Neuroreport 1993; 4(4): 444–6.
- Hara S, Kuriiwa F, Iwata N, Mukai T, Kano S, Endo T. Distinct effects of N omega-nitro-L-arginine on seizures induced by several drugs in mice. Pharmacol Biochem Behav 1996; 53(3): 673–7.

- Kaneko K, Itoh K, Berliner LJ, Miyasaka K, Fujii H. Consequences of nitric oxide generation in epileptic-seizure rodent models as studied by in vivo EPR. Magn Reson Med 2002; 48(6): 1051–6.
- Ferrendelli JA, Blank AC, Gross RA. Relationships between seizure activity and cyclic nucleotide levels in brain. Brain Res 1980; 200(1): 93–103.
- Pacinllo CA, McMahon Horner D, Hatton KW, Flynn JD. Methylene blue for the treatment of septic shock. Pharmacotherapy 2010; 30(7): 702–15.
- Stawicki SP, Sims C, Sarani B, Grossman MD, Gracias VH. Methylene blue and vasoplegia: who, when, and how? Mini Rev Med Chem 2008; 8(5): 472–90.
- Mayer B, Brunner F, Schmidt K. Inhibition of nitric oxide synthesis by methylene blue. Biochem Pharmacol 1993; 45(2): 367–74.
- Jing M, Ling GS, Bina S, Hart JL, Muldoon SM. Halothane attenuates nitric oxide relaxation of rat aortas by competition for the nitric oxide receptor site on soluble guanylyl cyclase. Eur J Pharmacol 1998; 342(2–3): 217–24.
- Hwang TL, Wu CC, Teng CM. Comparison of two soluble guanylyl cyclase inhibitors, methylene blue and ODQ, on sodium nitroprusside-induced relaxation in guinea-pig trachea. Br J Pharmacol 1998; 125(6): 1158–63.
- Masaki E, Kondo I. Methylene blue, a soluble guanylyl cyclase inhibitor, reduces the sevoflurane minimum alveolar anesthetic concentration and decreases the brain cyclic guanosine monophosphate content in rats. Anesth Analg 1999; 89(2): 484–9.
- Volke V, Wegener G, Vasar E, Rosenberg R. Methylene blue inhibits hippocampal nitric oxide synthase activity in vivo. Brain Res 1999; 826(2): 303-5.
- Furian AF, Fighera MR, Oliveira MS, Ferreira AP, Fiorenza NG, de Carvalho Myskinv J, et al. Methylene blue prevents methylmalonate-induced seizures and oxidative damage in rat striatum. Neurochem Int 2007; 50(1): 164–71.
- Fedele E, Marchi M, Raiteri M. In vivo NO/cGMP signalling in the hippocampus. Neurochem Res 2001; 26(8–9): 1069–78.
- Obay BD, Taşdemir E, Tümer C, Bilgin HM, Atmaca M. Dose dependent effects of ghrelin on pentylenetetrazole-induced oxidative stress in a rat seizure model. Peptides 2008; 29(3): 448-55.
- Jelenković A, Jovanović M, Ninković M, Maksimović M, Bošković B. Nitric oxide (NO) and an NMDA receptor antagonist in pentylenetetrazole-induced convulsions. Acta Veterinaria (Beograd) 2003; 53(2–3): 103–12.
- Paxinos G, Watson C. The rat brain in stereotaxic coordinates. 3rd ed. San Diego: Academic Press; 1982.
- Gurd JW, Jones LR, Mahler HR, Moore WJ. Isolation and partial characterization of rat brain synaptic plasma membranes. J Neurochem 1974; 22(2): 281–90.
- Sun M, Zigman S. An improved spectrophotometric assay for superoxide dismutase based on epinephrine autoxidation. Anal Biochem 1978; 90(1): 81–9.
- Auclair C, Voisin E. Nitroblue tetrazolium reduction. In: Greenwald RA, editor. Handbook of methods for oxygen radical research. Boca Raton, FL: CRC Press; 1985. p. 123-32.
- Villacara A, Kumami K, Yamamoto T, Mrsulja BB, Spatz M. Ischemic modification of cerebrocortical membranes: 5hydroxytryptamine receptors, fluidity, and inducible in vitro lipid peroxidation. J Neurochem 1989; 53(2): 595–601.

- Schröder H, Becker A, Lössner B. Glutamate binding to brain membranes is increased in pentylenetetrazole-kindled rats. J Neurochem 1993; 60(3): 1007–11.
- Walsh LA, Li M, Zhao TJ, Chiu TH, Rosenberg HC. Acute pentylenetetrazol injection reduces rat GABAA receptor mRNA levels and GABA stimulation of benzodiazepine binding with No effect on benzodiazepine binding site density. J Pharmacol Exp Ther 1999; 289(3): 1626–33.
- Halliwell B, Gutteridge JMC. Free radicals in biology and medicine. New York: Oxford University Press; 2007.
- Chiueh CC. Neuroprotective properties of nitric oxide. Ann N Y Acad Sci 1999; 890: 301–11.
- Patsoukis N, Zervoudakis G, Panagopoulos NT, Georgiou CD, Angelatou F, Matsokis NA. Thiol redox state (TRS) and oxidative stress in the mouse hippocampus after pentylenetetrazolinduced epileptic seizure. Neurosci Lett 2004; 357(2): 83–6.
- Patsoukis N, Zervoudakis G, Georgiou CD, Angelatou F, Matsokis NA, Panagopoulos NT. Thiol redox state and lipid and protein oxidation in the mouse striatum after pentylenetetrazolinduced epileptic seizure. Epilepsia 2005; 46(8): 1205–11.
- Patsoukis N, Zervoudakis G, Georgiou CD, Angelatou F, Matsokis NA, Panagopoulos NT. Effect of pentyleneterrazol-induced epileptic seizure on thiol redox state in the mouse cerebral cortex. Epilepsy Res 2004; 62(1): 65–74.
- Bashkatova V, Narkevich V, Vitskova G, Vanin A. The influence of anticonvulsant and antioxidant drugs on nitric oxide level and lipid peroxidation in the rat brain during penthylenetetrazole-induced epileptiform model seizures. Prog Neuropsychopharmacol Biol Psychiatry 2003; 27(3): 487–92.
- Echeverry MB, Salgado ML, Ferreira FR, da-Silva CA, Del Bel EA. Intracerebroventricular administration of nitric oxide-sensitive guanylyl cyclase inhibitors induces catalepsy in mice. Psychopharmacology (Berl) 2007; 194(2): 271–8.
- Deutsch SI, Rosse RB, Morn C, Koetzner L, Mastropaolo J. Interference with nitric oxide production and action potentiates the antiseizure efficacy of flurazepam. Pharmacol Biochem Behav 1995; 51(1): 133-7.
- Shimizu S, Yamamoto T, Momose K. Inhibition by methylene blue of the L-arginine metabolism to L-citrulline coupled with nitric oxide synthesis in cultured endothelial cells. Res Commun Chem Pathol Pharmacol 1993; 82(1): 35–48.
- Wolin MS, Cherry PD, Rodenburg JM, Messina EJ, Kaley G. Methylene blue inhibits vasodilation of skeletal muscle arterioles to acetylcholine and nitric oxide via the extracellular generation of superoxide anion. J Pharmacol Exp Ther 1990; 254(3): 872–6.
- O_ζ M, Lorke DE, Petroianu GA. Methylene blue and Alzheimer's disease. Biochem Pharmacol 2009; 78(8): 927–32.
- Hayashi Y, Ueda Y, Nakajima A, Yokoyama H, Mitsuyama Y, Ohya-Nishiguchi H, et al. Nitric oxide and hydroxyl radicals initiate lipid peroxidation by NMDA receptor activation. Brain Res 2002; 941(1-2): 107-12.
- Stojanović I, Jelenković A, Stevanović I, Pavlović D, Bjelaković G, Jevtović-Stoimenov T. Spermidine influence on the nitric oxide synthase and arginase activity relationship during experimentally induced seizures. J Basic Clin Physiol Pharmacol 2010; 21(2): 169–85.

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