



УНИВЕРЗИТЕТ У КРАГУЈЕВЦУ
ПРИРОДНО-МАТЕМАТИЧКИ ФАКУЛТЕТ

БЕНТОСНЕ СИЛИКАТНЕ АЛГЕ (BACILLARIOPHYTA)
У ПРОЦЕНИ ЕКОЛОШКОГ СТАТУСА
РЕКА ВЕЛИКЕ МОРАВЕ И САВЕ

• ,

, 2017.

<i>I</i>	
:	
: 23.12.1980.	
: ”,	
<i>II</i>	
: (Bacillari phyta)	
: 170	
: 31 , 36	
: 258	
: , -	
(): , (574). . Bacillariophyta. Diatome (582.26)	
: .	
<i>III</i>	
: 13.04.2016.	
:	
<p>1. , ; - ;</p> <p>2. ; :A , -</p> <p>3. , ; ; :</p> <p>4. ”; ; , ;</p>	
:	
<hr/> <p style="text-align: center;">:A , -</p> <hr/> <p style="text-align: center;">: ”</p> <hr/> <p style="text-align: center;">: , ;</p>	
:	

„ O 173025 „
37009,
FP7
„GLOBAQUA” No. 603629-ENV-2013-6.2.

(Bacillari phyta)

РЕЗИМЕ

OMNIDIA

(2010. 2011.)

(2011., 2012., 2014. 2015.)

pH,

(),

(),

2014.

Sh nnon-

Mayamaea cahabaensis

(),

Didymosphenia geminata *Diadlesmis confervacea*,

T

IPS

- CEE,

CEE

2

Benthic diatoms (Bacillariophyta) in assessment of the ecological status of the Velika Morava and the Sava rivers

ABSTRACT

Diatoms are dominant group in a phyto-benthos community of freshwater ecosystems, widely distributed, present throughout the year and they inhabit diverse habitats. Considering to be good bioindicators, many countries developed assessment of the ecological status of waters and monitoring of changes in aquatic ecosystems based on diatom indices. In accordance with the guidelines of the Water Framework Directive, the legislation of the Republic of Serbia formalized the assessment of ecological status/potential on the bases of biological water quality elements, which include benthic diatoms.

Until now algological investigations of the Velika Morava River and the Sava River primarily concerned the phytoplankton community, while the studies of the benthic diatoms communities recently began. Studies that include the assessment of the ecological status of large rivers based on benthic diatoms have not been done on the territory of Serbia so far.

The goals of our research were: qualitative and quantitative analysis of the benthic diatoms community composition, to determine their spatial and seasonal dynamics, to determine the values of physical and chemical parameters at sampling sites and the most important environmental parameters that affect the investigated communities, assessment of the water quality based on the diatom indices values using OMNIDIA software, assessment of the ecological potential of the Velika Morava River and the Sava River (part of the flow through Serbia), in accordance with the legislation of the Republic of Serbia, consideration of the efficiency of benthic diatoms as indicators in the water monitoring and the effectiveness of selected ecological indices, evaluation of environmental potential and identification of the specific indicator taxa and forms.

Phyto-benthos samples from the Velika Morava River were collected annually (from 2010 to 2011) at five sampling sites, while samples from the Sava River were collected each September during a period of four years (2011, 2012, 2014 and 2015) at a total of 33 sites along the entire watercourse. After processing the material, the analysis of the benthic diatoms community was carried out according to the research objectives.

In the Velika Morava River presence of 162 taxa was detected, while in the Sava River presence of 184 taxa was registered. The dominant and frequent taxa of the Sava and the Velika Morava rivers are considered as eutrophic and hypereutrophic taxa.

The greatest influence on seasonal dynamic of benthic diatoms of the Velika Morava have environmental parameters pH, temperature and arsenic. The most important environmental parameters affecting the community of benthic diatoms of the Sava are arsenic and silicon, with the greatest influence on sites in the lower course of the river.

Along the Sava River, the composition of the benthic diatoms communities changes from the dominance of the forms closely attached to the substrate (upstream) to the dominance of the motile forms (middle and lower flow), which is in accordance with the general changes in the Sava River, from the sub-alpine river to the lowland river, with the dominance of the smaller fractions of the substrate.

The high water levels recorded in September 2014 on the Sava River, didn't result in decrease of Shannon's diversity index values, which confirms the resistance of benthic diatoms to this type of pressure. Our research suggests that large rivers are an important habitat for benthic diatoms.

The species *Mayamaea cahabaensis*, first time identified in the Sava and the Velika Morava rivers (and therefore in diatom flora of the Serbia), was recorded with a high abundance. Two potentially invasive taxa *Didymosphenia geminata* and *Diadesmis confervacea* are present in the Sava River with a low abundance. Teratological forms of diatoms have been recorded at all sites in the Velika Morava River and at several sites on the Sava River. It has been confirmed that the share of teratological forms in diatom community has a bioindicator potential.

In the case of the Velika Morava River, our research indicate that assessment according to national legislation is more reliable using IPS index in comparison to CEE. It is necessary to consider changing class boundaries for the CEE index for type 2 watercourses.

Diatom indices are sensitive to increased concentrations of arsenic and iron, although indices were primarily designed as indicators of organic pollution and nutrient load. Having this in mind, benthic diatoms can be considered as a reliable indicator of the presence of multiple pressures in the case of large lowland rivers, and they can be used as a parameter of general degradation.

The reliability of the standard methodology for benthic diatoms sampling in routine monitoring, in the case of the Velika Morava and Sava rivers, is confirmed.

1.	1
1.1.	(Bacillariophyta)	2
1.2.	8
1.3.	11
1.4.	19
1.5.	20
2.	21
3.	23
3.1.	24
3.1.1.	25
3.1.2.	26
3.2.	27
3.3.	33
3.4.	,	35
3.5.	36
4.	39
4.1.	40
4.1.1.	40
4.1.1.1.	<i>Mayamaea cahabaensis</i> Morales & Manoylov	46
4.1.2.	48
4.1.3.	Sh nnon-	55
4.1.4.	56

4.2.	58
4.3.	64
4.4.	67
4.4.1.	81
4.5.	82
4.6.	84
4.6.1.	84
4.6.2.	2011., 2012., 2014. 2015.	92
4.6.3.	Sh nnon-	98
4.6.4.	99
4.7.	102
4.8.	108
4.9.	111
4.9.1.	118
4.10.	120
5.	121
6.	141
7.	146

1.	-	12
2.	(Prygiel Coste, 2000 , 2001 Coste , 2009) Predojevi , 2017)	(Kelly 14
3.	IPS CEE, (. , 74/2011)	17
4.		18
5.		19
6.		29
7.		31
8.	(+ ; +* o ;	40
9.	(% (2010.)	48
10.	(% (, 2010.)	50
11.	(% (, 2010.)	51
12.	(% (2010. , , 2011.)	54

13.	61
14.	62
15.	62
16.		
(p 0,05)	64
17.	(2010. 2011.) (1 ,2 ,3 , 4 5), :	
	(), (), (), () () (Prygiel Coste, 2000 Coste ., 2009); TDI (TDI_PT 20 %), (TDI_PT < 20 %)	
	80
18.	* (Spearman- ; p<0,05) (2010. 2011.)	81
19.	, 2010. 2011.	83
20.	2011., 2012., 2014. 2015. () , 7) (+ ; +* o ;)	85
21.	(%) 2011.	93
22.	(%) 2012.	95

23.	(%)	2014.	96
24.	(%)	2015.	97
25.		2014.	106
26.		2015.	106
27.		(p 0,05)	108
28.	(2011., 2012., 2014. 2015.) (7), : (), (), (), () (Prygiel Coste, 2000 Coste ., 2009); TDI (TDI_PT 20 %), (TDI_PT <20 %)		117
29.	- (p<0,05)	2014. ()	2015. () 118
30.	* (Pearson- ; p<0,05)	2014. ()	2015. () 119
31.		2011., 2012., 2014. 2015.	120

1.	(
	https://commons.wikimedia.org/wiki/File:Diatom_biology.jpg);	
A.	; 3
2.	Surirellales:	.	
	,	5 µm;	.
	,	2 µm (
	Ruck	Theriot, 2011). 5
3.	<i>Navicula tripunctata</i> (O.F.Müller)		
Bory,	1 µm;	(),
	<i>Navicula</i> (
	http://westerndiatoms.colorado.edu/taxa/species/Navicula_tripunctata		
)	 6
4.	(
	http://www.seagrant.umn.edu/newsletter/2013/01/what_good_is_a_diatom.html		
)	 7
5.	–	 24
6.	: 1 –	, 2 –	
	, 3 –	, 4 –	5 –
		 28
7.	: 1 –	, 2 –	
	, 3 –	, 4 –	, 5 –
	, 6 –	, 7 –	, 8 –
	, 9 –	, 10 –	, 11 –
	, 12 –		, 13 –
	, 14 –	, 15 –	, 16 –
	, 17 –		, 18 –
	, 19 –	, 20 –	, 21 –
	, 22 –		
	, 23 –	, 24 –	25 –
		 30
8.			
		 44
9.	,		
	10 µm: <i>Ulnaria ulna</i> (), <i>Navicula capitatoradiata</i> (), <i>Fragilaria recapitellata</i> (
	<i>F. vaucherie</i> (), <i>Diatoma moniliformis</i> (,) <i>D. vulgaris</i> (
		 45

10. <i>Mayamaea cahabaensis</i> ; 10 μm (, , ,) ;)	47
11. Sh nnon- (H)	55
12. DAPC (PA1 31,2 % PA2 18,8 %); . ; . (5)	57
13. CCA (; Monte Carlo ; CCA1 29 % CCA2 23,2%); . ; . (8)	66
14. (%)	67
15. 2010. () TDI_PT (%) ()	68
16. 2010. () TDI_PT (%) ()	69
17. 2010. () TDI_PT (%) ()	70
18. 2010. () TDI_PT (%) ()	71
19. 2010. () TDI_PT (%) ()	72
20. 2010. () TDI_PT (%) ()	73

21.	2010.		
	() TDI_PT (%) ()	74
22.	2010.		
	() TDI_PT (%) ()	75
23.	2010.		
	() TDI_PT (%) ()	76
24.	2011.		
	() TDI_PT (%) ()	77
25.	2011.		
	() TDI_PT (%) ()	78
26.	2011.		
	() TDI_PT (%) ()	79
27.		91
28.	, 10 µm:		
	<i>Diatoma ehrenbergii</i> () <i>D. vulgaris</i> (,)	92
29. Sh nnon-	(H)		
	(1 33, 7)	99
30. CA	(CA1 – 11,38		
% , CA2 – 9,24 %); .		
	(2011., 2012., 2014. 2015.		
);	20.....	101
31. CCA	(
	, Monte Carlo ; 24,2% (CCA1) 21,1%		
(CCA2)); .		
	(4); .		
	(18)	110
32.	(%)		
		111

33.		2011.	
	()	TDI_PT (%) () 112
34.		2012.	
	()	TDI_PT (%) () 113
35.		2014.	
	()	TDI_PT (%) () 115
36.		2015.	
	()	TDI_PT (%) () 116

1. Увод

1.1. Опште карактеристике силикатних алги (Bacillariophyta)

2014).

(Sims ., 2006; Medlin, (Sims ., 2006).

(Pascher, 1921 Round ., 1990).

(Harwood Gersonde, 1990; Sims ., 2006).

72.500, 20.000 (Bacillariophyta) (Guiry, 2012). 200.000 (Mann Doop, 1996).

.

.

(Falkowski Raven, 1997 Bellinger Sigee, 2010).

(Cox, 2011).

.

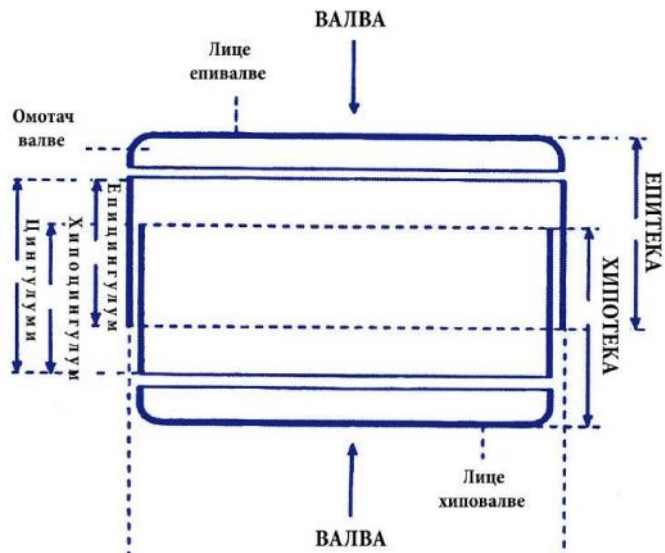
Navicula Bory de Saint-Vincent *Nitzschia* Hassall (Tuchman, 1996),

.

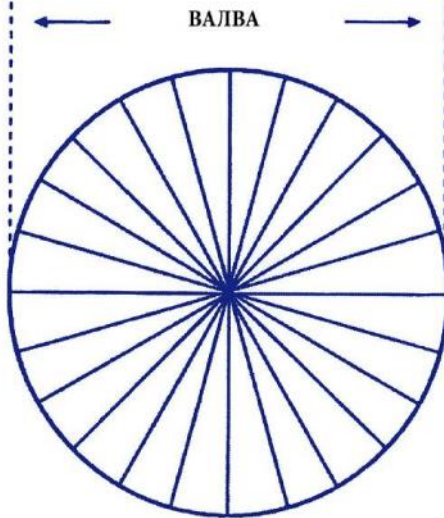
.

() () (Gaul ., 1993),

А



Б



1.

(
https://commons.wikimedia.org/wiki/File:Diatom_biology.jpg);

А.

; .

(1), „ ” (),

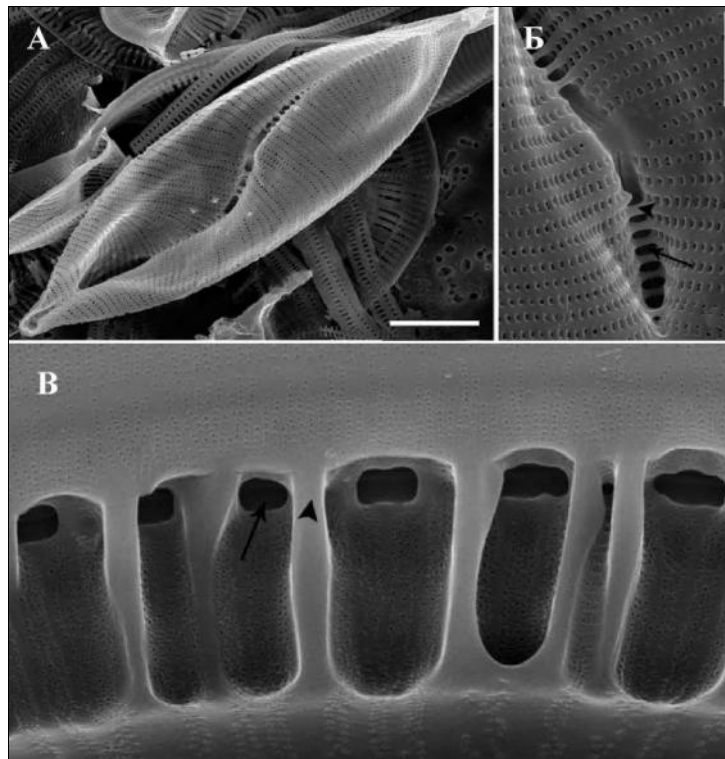
– (Blaženi, 2000).

(. μ).
 , ()
 . -
 - (Round ., 1990).
 , .
 ,
 (1), - ,
 (1), .
 ,
 : , (Blažen i , 2000).
 , ,
 , . (Krizmani , 2009).
 :
 , .
 , , .
 ,
 : ,
 , .
 ,
 (Bellinger Sigeo, 2015).
 ,
 (Cox, 2011).
 Round- (1990)
 : Coscinodiscophyceae (),
 Fragilariophyceae () Bacillariophyceae ().
 ,
 (Medlin Kaczmarska, 2004; Medlin, 2016).

()
 , (2).
 ,
 . ()
 ,
 ()
 ,
 . 27

Bacillariales, Rhopalodiales Surirellales

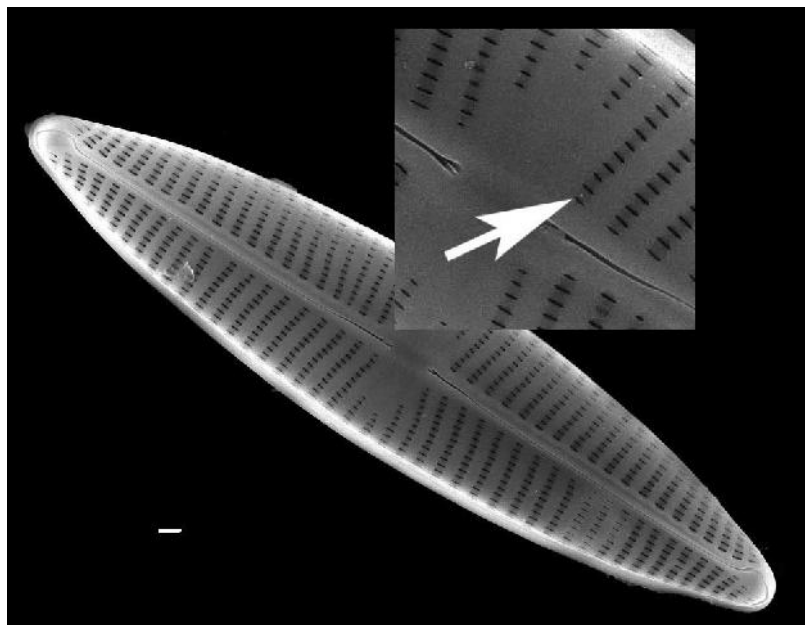
(Ruck Theriot, 2011).



2. Surirellales: .
 , 5 μm; . , 2 μm (Ruck
 Theriot, 2011)

(Blaženi, 2000)

(3), () ()
,).
,
() / (Cox,
2011).
, - .
,
10 μm (Barber
Haworth, 1981).



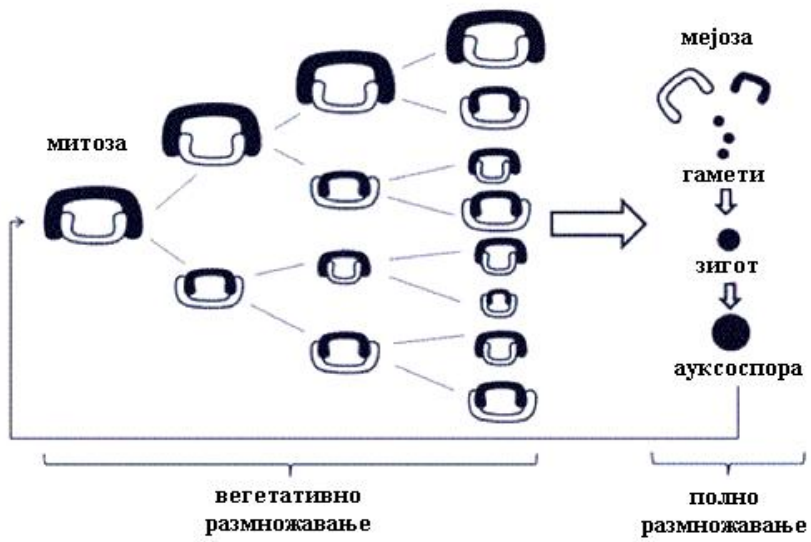
3. *Navicula tri-punctata* (O.F.Müller) Bory,
1 μm; (),
Navicula (http://westerndiatoms.colorado.edu/taxa/species/Navicula_tripunctata
)

(Krizmani , 2009).

2011).

(Cox,

(4).



4.

(

http://www.seagrant.umn.edu/newsletter/2013/01/what_good_is_a_diatom.html

)

(Round , 1990).

30 % 40 %
(Geitler, 1932 Edlund Stoermer, 1997).

(Edlund Stoermer, 1997).

1.2. Екологија бентосних силикатних алги

(Lamberti, 1996; Steinman, 1996).
(Allan
Castillo, 2007).

(Steinman , 1987; Tuchman Stevenson, 1991)

(Barnese Lowe, 1992).

(Bacillari phyceae),
 (Chlorophyta), - (Cyanobacteria), - (Xanthophyta)
 (Rhodophyta) (Simi Simi , 2012).

,

,

,

(), (),
 () ()
).

,

(Manoylov, 2009),
Achnantheidium Kütz., *Planothidium* Round & Bukht.,
Achnanthes Bory, *Gomphonema*, *Rhoicosphenia* Grunow .

(. *Achnanthes delicatula* (Kütz.) Grunow, *Amphora ovalis* Kütz.,
Planothidium dubium (Grun.) Round & Bukht.) ,
 (*Staurosira construens* var. *venter* (Ehr.) Hamilton,
Achnanthes exigua Grun., *Staurosirella pinnata* (Ehr.) Williams & Round) (Lowe, 2011).

(Krejci Lowe, 1986).

,

(*Nitzschia* Hassall, *Surirella* Turpin, *Cymatopleura* W. Smith)
 (*Gyrosigma* Hassall, *Pleurosigma* W. Smith).
 (Moss, 1977)

,

,

(Stal, 2010).

(. *Cocconeis pediculus* Ehr.),
 (. *Ulnaria ulna* (itzsch.)
 Compère, *Rhoicosphenia abbreviat* (C. Agardh) Lange-Bertalot).

(Burkholder, 1996),

(Burkholder Wetzzel,
 1990).

(Gross, 2003),
 (Dudley, 1992; Kupferberg, 1997).

Bacillariophyta, e e e
 Round Lee (1989)

Amphora copulata var. *epiphytica* Round & Lee
Nitzschia sigmoidea (Nitzsch) W. Smith., Tiffany Lange (2002) *Cocconeis*
pediculus Ehr., *Rhoicosphenia* sp. *Planothidium* cf. *delicatum* (Kutz.) Round &
 Bukhtiyarova e *Pleurosira laevis* (Ehr.) Compère.
 (Round, 1990),
 (Tiffany Lange, 2002).

a.
 (Gaiser Bachmann, 1993).

(McCormic Cairns, 1994).

(Stevenson, 1984; Hall Smol, 1992; Bennion , 1996; Descy, 1979; Kelly Whitton, 1995; Rott , 1997, 1999), (Ziemann, 1999; Cumming Smol 1993; Potapova Charles, 2003), pH (Andrén Jarlman, 2008; Keithan , 1988; Round, 1990; Van Dam , 1994) (Medley Clements,1998; De Jonge , 2008; Sabater, 2000; Morin , 2012).

1.3. Бентосне силикатне алге у оцени еколошког статуса

(1)

OMNIDIA (Lecoite ., 1993)

: IPS, IBD, EPID, DICH, IDP, IDAP, LOBO, DESCY, IDSE, CEE IDG.

: TDI, TID SHE,

: SID, SLA WAT.

(Bellinger et al., 2006; Ndiritu et al., 2006), (Maznah Mansor, 2002) (Newall Walsh, 2005).

1.

	*		
IPS	Specific Pollution Index	Cemagref, 1982	
IBD	Biological Diatom Index	Lenoir Coste, 1996	
EPID	Diatom-based Eutrophication/Pollution Index	Dell'Uomo, 2004	
SHE	Schiefele and Schreiner's Index	Steinberg Schiefele, 1988	
TDI	Trophic Diatom Index	Kelly Whitton, 1995; Kelly et al., 2001	
TID	Rott's Trophic Index	Rott et al., 1999	
SID	Rott's Saprobic Index	Rott et al., 1997	
SLA	Sládeček's Index	Sládeček, 1986	
DICH	Swiss Diatom Index	Hürlimann Niederhauser, 2002	
PDI	Pampean Diatom Index	Gómez Licursi, 2001	
IDAP	Artois-Picardie Diatom Index	Prygiel et al., 1996	
LOBO	Lobo's Index	Lobo et al., 2002	
WAT	Watanabe's Index	Watanabe et al., 1988	
DESCY	Descy's Index	Descy, 1979	
IDSE	Leclereq and Manquet's Index	Leclereq Manquet, 1987	
CEE	European Economic Community Index	Descy Coste 1991	
IDG	Generic Diatom Index	Rumeau Coste, 1988	

*

Zelinka Marvan (1961):

$$X_i = \frac{\sum_{i=1}^n h_i \cdot G_i \cdot x_i}{\sum_{i=1}^n h_i \cdot G_i}$$

X_i –

h –

G –

x_i –

IPS.
(Prygiel ., 2002),

OMNIDIA . IPS
(Kelly ., 1995), j (Prygiel Coste,
1993), (Vilbaste, 2004), Ács ., 2004, Van Dam ., 2005),
(Kwandrans ., 1998), (Hlúbiková ., 2007) (Gomá
. , 2004). IPS IDG, IBD,
EPID CEE. IDG,

. IBD
(Coste , 2009)
209
(Lenoir Coste, 1996), . Coste
(2009) 838 ,
,
. OMNIDIA (a 6.04),
(IBD
2014) , EPID CEE,
,
(), TDI.
,
OMNIDIA (6.04)
1 20.
IBD (Prygiel Coste, 2000 Coste ., 2009)
(2).
TDI
(Kelly ., 2001 Predojevi ,
2017) (2). ,

(TDI_PT, OMNIDIA 6.04). TDI_PT

TDI : TDI_PT 20 %
, 20 %
(Kelly Whitton, 1995; Kelly ., 2001).

2.

(Prygiel Coste,
2000 Coste ., 2009) (Kelly ., 2001

Predojevi , 2017)

	(TDI)		
20 17			
17 13			
13 9			
9 5			
5			

” ”,

, , ,

.

– () (WFD, 2000),

,

.

,

,

(WFD, 2000),

,

/ .

,

,

,

(McGarrigle, 2010).

”

”(2.10

) (WFD CIS Guidance Document No 2.,

2003).

(Ilies, 1978),

Paunovi (2007).

(Wallin ., 2003).

(WFD, 2000).

(. *ecological quality ratio* EQR). EQR

) () (WFD, 2000; ., 2015).

(
), .
.
.
,
(, 2014).
- ,
” ” (..
2015), ,
.
, -
, ,
, 2004. (ikanovi ., 2010).
(. 30/2010)
/
, -
.”
” (. , 74/2011).
.
, ”
”
”
: ,, 1”
, ,, 2”
, ,, 3”
500 m , ,, 4”

500 m

, „ 5”

1 „ 6”

3 4,

” (. 96/2010).

(,),
(Kelly, 2013).

: IPS

CEE (3).

3.

IPS

CEE, (. , 74/2011)

		I	II	III	IV	V
1	IPS	14	14 10	10 8	8 6	6
	CEE	12	12 9	9 7	7 5	5
2	IPS	16	16 14	14 12	12 9	9
	CEE	12	12 9	9 7	7 5	5
3	IPS	16	16 14	14 12	12 9	9
	CEE	12	12 9	9 7	7 5	5
4	IPS	16	16 14	14 12	12 9	9
	CEE	12	12 9	9 7	7 5	5
5	IPS	14	14 10	10 8	8 6	6
	CEE	12	12 9	9 7	7 5	5
6	IPS	14	14 10	10 8	8 6	6

(WFD CIS Guidance Document No 13., 2005).
 (WFD CIS Guidance Document No 13., 2005).
 : IPS, CEE
 EPID,
 (Kelly, 2013).

(„ , ”; . *one out, all out*).
 / (I),
 (II), (III), (IV) (V),
 (. , 74/2011).
 ” ” ” ”
 ” ”. (. , 74/2011),
 (4) (5), /

4.

5.

	-		-	
	-		-	
	-		-	
	-		-	

1.4. Употреба бентосних силикатних алги у оцени еколошког статуса великих река

...

(Schöll ..., 2012).

/

(Makovinska Hlubikova, 2015).

(*Joint Danube Survey*,

<https://www.icpdr.org/main/activities-projects/joint-danube-survey>)

(2001., 2007. 2013.)

(...),

(Kireta , 2012),
(De Jonge , 2008; Szabó , 2005; Fore
Grafe, 2002).

1.5. Преглед истраживања бентосних силикатних алг Велике Мораве и Саве

(Obuškovi , 1979; Obuškovi , 1985; Martinovi -
Vitanovi , 1996; Martinovi -Vitanovi , 2004; Obuškovi Markovi , 1987; Lauševi
, 1998; а о , 2008) (Simi , 2014; Simi Pantovi ,
2010).

(Simi , 2015; Vasiljevi , 2017).

(Obuškovi Kalafati , 1979;
, 2010).

(Andreji , 2012),
(Krizmani , 2013), (Vidakovi , 2013),
(Vasiljevi , 2014), (Jakovljevi
, 2016), (Jakovljevi , 2016), - - (Jakovljevi
, 2014), - (Predojevi , 2017).

2. Циљеви

:

•

;

•

;

•

;

•

OMNIDIA ;

• O

(

)

;

•

;

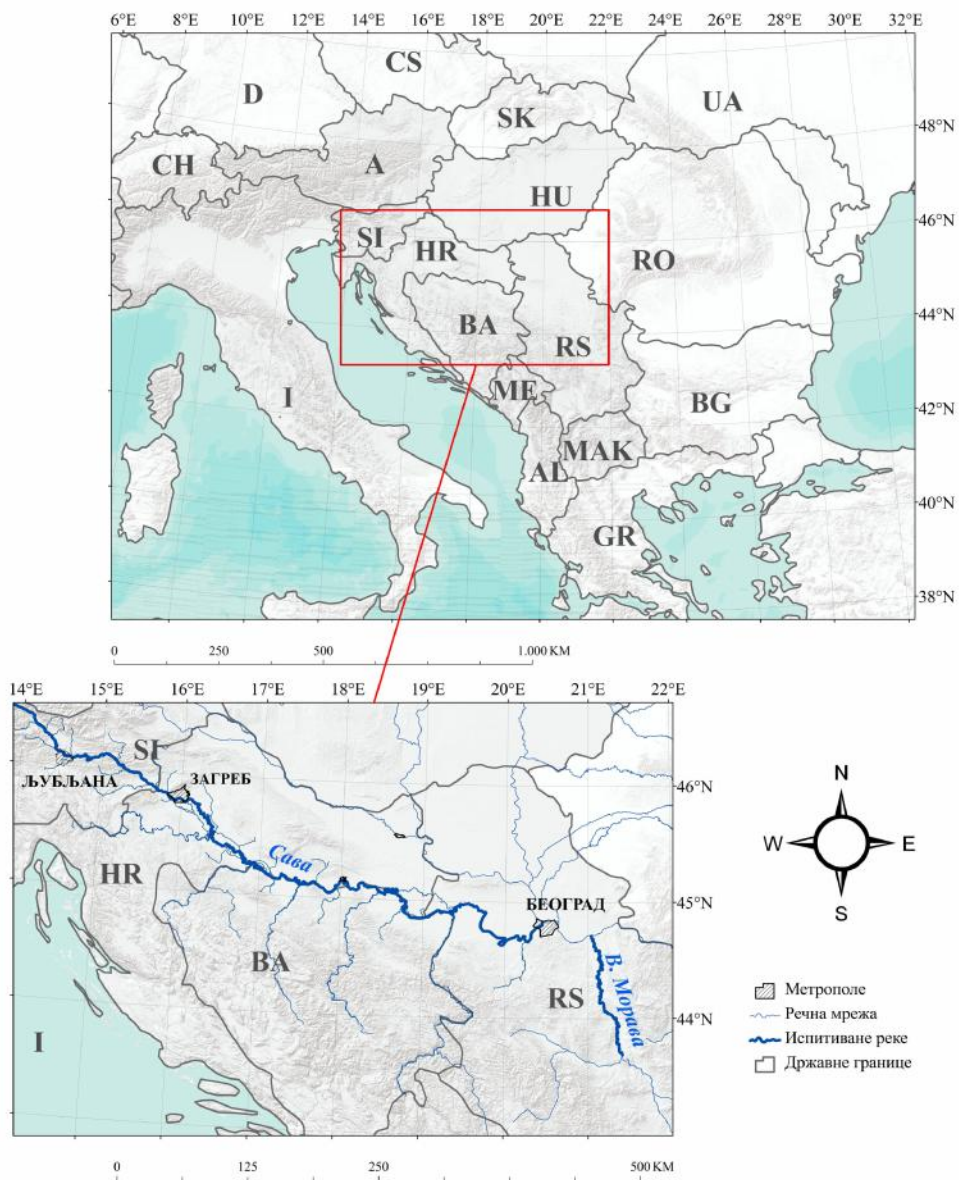
•

.

3. Материјал и методе

3.1. Подручје истраживања

— (5). ,



5.

3.1.1. Река Велика Морава

(5),
185 km
129 m
67 m (,
, 2014).
37.561 km² 42,4%
(, 2014).
45 km
17 km
(, 2014).
,
(, 2014).
, 1966.
,
245,5 km.
60 km.
,
,
(Kolarevi ., 2012),
,
(Markovi ., 2011).

3.1.2. Река Сава

je 97.713 km², (Sava River Basin Analysis Report, 2009). (12%, 26%, 39,2%, 15,5%, 7,1% 0,2%) 9 (Komatina Grošelj, 2015).

(Sava River Basin Management Plan, 2014).

(5) 945 km.

1.722 m³/s,

(265 km), (129 km) (597 km) (Sava River Basin Management Plan, 2013).

„ ” (Schwarz, 2016).

(Sava River Basin Management Plan, 2013)

(Vidmar ., 2016)

(Vidmar ., 2016), (Sava River Basin Management Plan, 2013).

3.2. Прикупљање узорака и пратећих података о локалитетима

„

“

“

37009 „

” 173025 „

” (

),

” (2011. 2012.

) „GLOBAQUA” (Navarro-Ortega ., 2015) (

2014. 2015.).

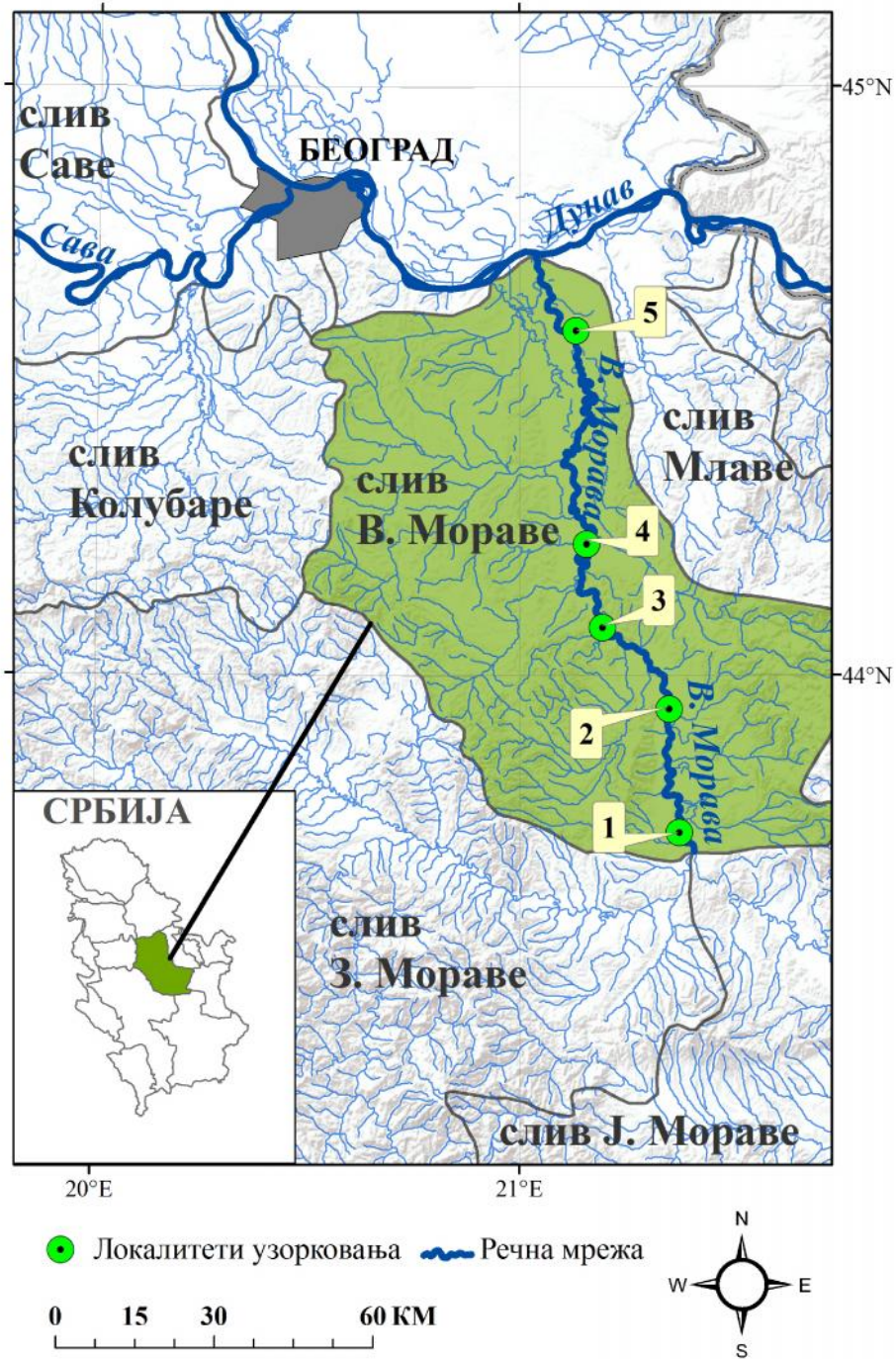
, 2010. 2011.

5 (6),

. 59 .

,

6.



6. : 1 – , 2 – ,
 3 – , 4 – 5 –

6.

			(m)		
	(°)	(°)			
	43,717033	21,384217	130	90% 10%	, , ,
	43,948100	21,364467	115	70% 30%	,
	44,084983	21,189133	102	50% 45% 5%	, ,
	44,224750	21,152900	95	65% 20% 15%	, *
	44,603200	21,086317	72	80% 20%	,

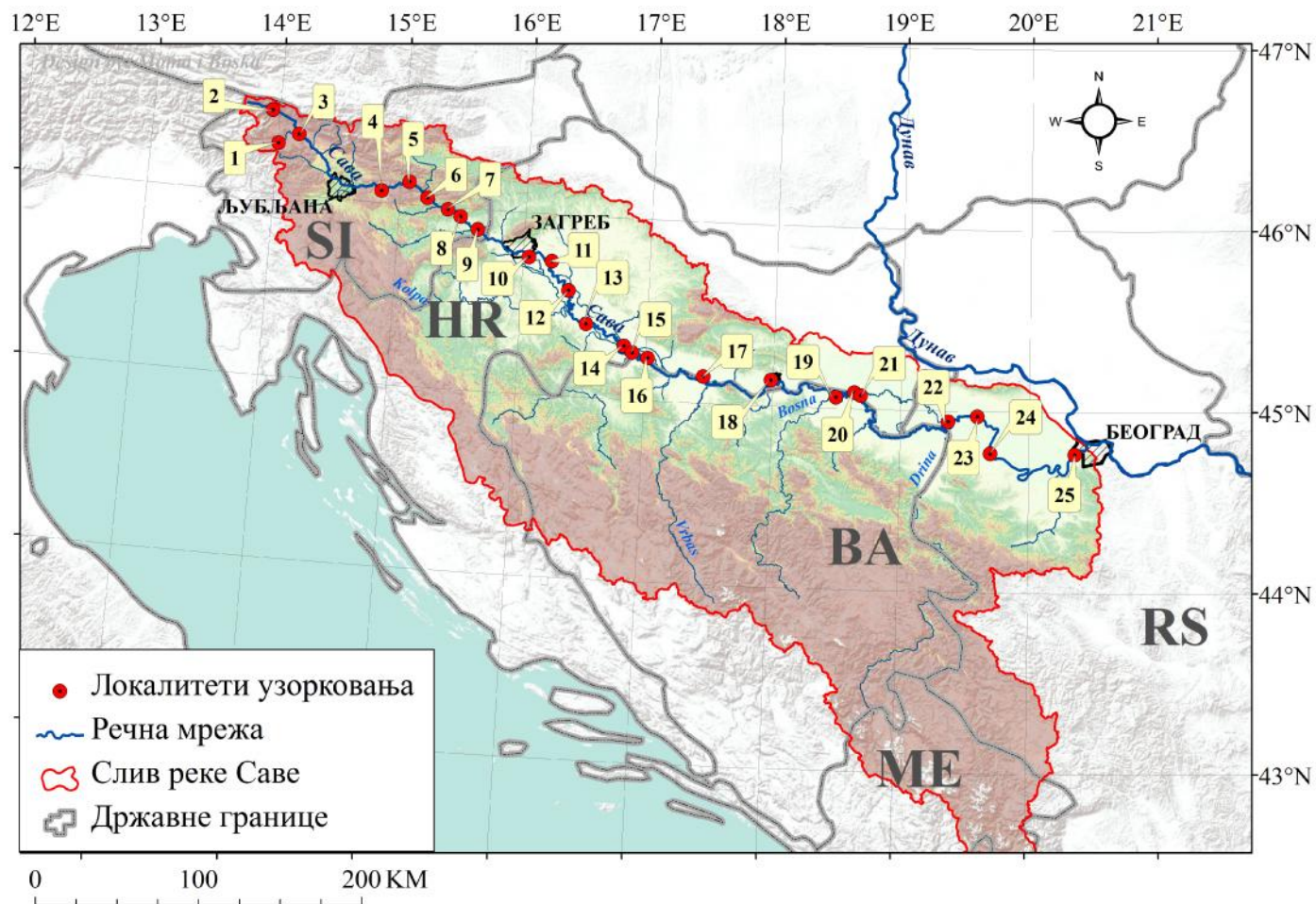
* e , (Jovanovi Rosi , 2010)

2011., 2012., 2014. 2015.

33 .
(7).

48 . , ,

7.



7. : 1 – , 2 – , 3 – , 4 – , 5 – ,
 6 – , 7 – , 8 – , 9 – , 10 – , 11 – , 12 – , 13 – , 14 – ,
 15 – , 16 – , 17 – , 18 – , 19 – , 20 – , 21 – , 22 – ,
 23 – , 24 – 25 –

7.

								(m)	(rkm)	*	
		2011	2012	2014	2015	(°)	(°)				
1				+		46,280317	14,005291	507	937	1	
2					+	46,459967	13,940096	661	930	1	
3	()			+	+	46,339529	14,163860	409	908	1	
4	()			+		46,292997	14,260589	406	900	1	
5	()				+	46,055600	14,823100	230	819	1	
6	()			+	+	46,066067	14,850483	225	810	1	
7			+			46,124800	15,059900	209	790	1	
8	()		+		+	46,042900	15,226300	194	776	1	
9	()		+			45,990000	15,383500	158	760	1	
10	()		+			45,955800	15,487300	154	750	1	
11	()		+			45,897700	15,593100	142	739	1	
12	()			+	+	45,890362	15,630107	137	736	1	
13	()			+		45,854966	15,694481	134	729	1	
14	()				+	45,785695	15,981591	112	664	2	
15			+			45,741400	16,226100	98	657	2	
16		+				45,586800	16,376970	95	622	2	
17			+			45,407129	16,523449	93	578	2	
18		+				45,293320	16,830050	92	533	2	
19	()				+	45,263670	16,894265	90	489	3	
20			+			45,238006	17,015899	90	492	3	
21		+				45,161669	17,444322	87	441	3	
22	()		+	+	+	45,144906	17,984106	82	360	3	
23		+				45,065470	18,478120	80	314	3	
24			+			45,096240	18,633730	81	284	3	
25	()			+	+	45,075484	18,686883	77	262	3	
26	()			+		45,015038	18,739811	76	249	3	
27		+	+			44,941889	19,369525	75	163	3	
28	()	+	+	+		44,973012	19,596115	73	139	3	
29	()		+	+	+	44,913575	19,752491	72	118	3	
30	()	+			+	44,769900	19,699400	71	106	3	
31		+				44,704040	20,313800	70	16	3	
32	()		+	+	+	44,768511	20,355560	69	14	3	
33	()			+		44,806247	20,443660	69	2	3	

* : 1 – , 2 – , 3 –

, 2011, 2012).

(http://www.hidmet.gov.rs/ciril/hidrologija/karakteristicne_v.php).

2011, 2012. 2015.

(, 2012, 2013, 2016; Meteorološki i hidrološki bilten br. 9, 2011, 2012, 2015; Pregled hidroloških razmer površinskih voda v Sloveniji, 2011, 2012, 2015). 2014.

(, 2015; Meteorološki i hidrološki bilten br. 9, 2014; Pregled hidroloških razmer površinskih voda v Sloveniji, 2014).

EN 13946 (2003).

10 m

64 mm – 256 mm,

10 cm²,

(HCHO)

1 % 4 %,

(6) AQEM (AQEM, 2002),
Simonovi . (2017),

3.3. Обрада материјала

(Krammer
Lange-Bertalot, 1986)

Naphrax.
1000 x Carl Zeiss Axio Lab1, Axiocam ERc 5s
ZEN

1000 x 1600 x
Carl Zeiss Axio Imager M1, Axiocam MRc5 Axio Vision 4.8

Tescan VEGA TS 5130MM,

EN
14407 (2004).
(Hofmann ., 2011; Krammer

Lange-Bertalot, 1986, 1988, 2004, 2011; Lange-Bertalot, 1993, 2001; Levkov , 2013; Krammer, 1997 a, 1997 , 2000, 2002, 2003),

„AlgaeBase” (Guiry Guiry, 2017).

(%) 300 500

(Falasco , 2009).

OMNIDIA

Shannon-

(H) (Shannon 1948),

(Krebs, 2001).

() ,

0,

. Shannon-

5 (Washington, 1984 Krebs, 2014).

2010. 2011. ,

(- , 2010),

(, 2011).

a (T; °C), pH, (EC; µS/cm), (DO; mg/l), (O₂%), (mg/l), (TH; mg CaCO₃/l), (Ca²⁺; mg/l), (Mg²⁺; mg/l), (NH₄⁺; mg/l), (NO₃⁻; mg/l), (NO₂⁻; mg/l), (SO₄²⁻; mg/l), (PO₄³⁻; mg/l), (Fe; µg/l), (Mn; µg/l), (Zn; µg/l), (Cd; µg/l), (Pb; µg/l), (Cu; µg/l), (Hg; µg/l), (Ni; µg/l) (As; µg/l).

2014. 2015. ,

” ” ,

, : (T; °C), (DO; mg/l), (O₂%), (EC; µS/cm), pH - (ORP; mV), (Alk; meq/kg),

(DOC), (Na⁺; mg/l), (K⁺; mg/l), (Ca²⁺; mg/l),
(Mg²⁺; mg/l), (Si; mg/l), (NO₃; mg/l),
(SO₄²⁻; mg/l), (Cl; mg/l), (Br; mg/l) (F; mg/l), o (Ag;
μg/l), (As; μg/l), (Ba; μg/l), (Cd; μg/l), (Co; μg/l),
(Cr; μg/l), a (Cu; μg/l), (Fe; μg/l), (Mn; μg/l), (Mo; μg/l),
(Ni; μg/l), (P; μg/l), (Pb; μg/l), (Rb; μg/l), (Se; μg/l),
(Sr; μg/l), (V; μg/l) (Zn; μg/l).

3.4. Дијатомни индекси, одређивање класе вода и индикативног еколошког потенцијала

17 OMNIDIA
6.04 (Lecointe .., 1993): IBD (Lenoir Coste, 1996), IPS (Cemagref, 1982),
IDG (Rumeau Coste, 1988), DESCY (Descy, 1979), SLA (Sládec k, 1986), IDSE (Leclerq
Manquet, 1987), IDAP (Prygiel .., 1996), EPID (Dell'Uomo, 2004), LOBO (Lobo
.., 2002), DI-CH (Hürlimann Niederhauser, 2002), RTI (Rott .., 1999), RSI (Rott
.., 1997), CEE (Descy Coste 1991), WAT (Watanabe .., 1990), TDI (Kelly
Whitton, 1995; Kelly .., 2001), PDI (Gómez Licursi, 2001) SHE (Steinberg
Schiefele, 1988).

96/2010)

(. , 74/2011).

VMOR_2 (VMOR_3
2.
SA_1 (), (), ()

SA_2 (), () ()
 1 .
 , IPS CEE
 (3),
 , 74/2011.
 (VMOR_3 SA_1 SA_2),
 .
 (WFD
 CIS Guidance Document No 13., 2005),
 „ , ”;
 .
 ,
 , 74/2011.

3.5. Статистичка обрада података

(.
Discriminant Principal Components Analysis – DPCA) (Yendle MacFie, 1989)
 „FLORA” (Karadži Marinkovi , 2009; Karadži , 2013),
 ().
 -
 , 2010.
 2011. .
 :
 , 2010.

2011.

34

„FLORA” (Karadži Marinkovi , 2009; Karadži , 2013).

Mantel- (Mantel, 1967) ()

(T, pH, EC, DO, O₂%,
, NH₄⁺, NO₃⁻, NO₂⁻, SO₄²⁻, PO₄³⁻, Ca²⁺, Mg²⁺ TH),

(Zn, Cd, Pb, Cu, Fe, Mn, Hg, Ni
As).

(. *Forward Selection* – FS) Pearson-
(p<0,05) Monte Carlo test (999 ; p<0,05) (Karadži ,
2013). (. *Canonical Correspondence Analysis* –
CCA) (Ter Braak, 1987)

FS STATISTICA 6 (StatSoft Inc., 2001). Shapiro Wilk-
(Shapiro , 1968) (p<0,05)
FS
(Spearman-).

(. *Correspondence Analysis* – CA) (Greenacre, 1984) „FLORA” (Karadži
Marinkovi , 2009; Karadži , 2013).

2014. () 2015. ()
)
25 (12 2014. 13 2015.
, 7).

„FLORA” (Karadži Marinkovi , 2009; Karadži , 2013).

Mantel- (Mantel, 1967) ()

c (T, DO, O₂%, Alk, EC, DOC, ORP, pH, Ca²⁺, Mg²⁺, Na⁺, NO₃⁻, SO₄²⁻, Cl⁻, P³⁻, Si), (Ag, As, Ba, Br, Cd, Co, Cr, Cu, F⁻, Fe, K⁺, Mn, Mo, Ni, Pb, Rb, Se, Sr, V⁵⁺, Zn).

(. *Forward Selection* – FS) Pearson- (p<0,05) Monte Carlo test (999 ; p<0,05) (Karadži , 2013). (. *Canonical Correspondence Analysis* – CCA) (Ter Braak, 1987)

FS STATISTICA 6 (StatSoft Inc., 2001). Shapiro Wilk- (Shapiro .., 1968)

p<0,05), (2014.) (2015.) (p<0,05) (Pearson-).

4. Резултати

4.1. Бентосна заједница силикатних

Мораве

4.1.1. Флористички састав

162 (101), 50 (99), (126), (115), (109),
Navicula (19), *Gomphonema* (16) *Nitzschia* (16).
 8.

8.

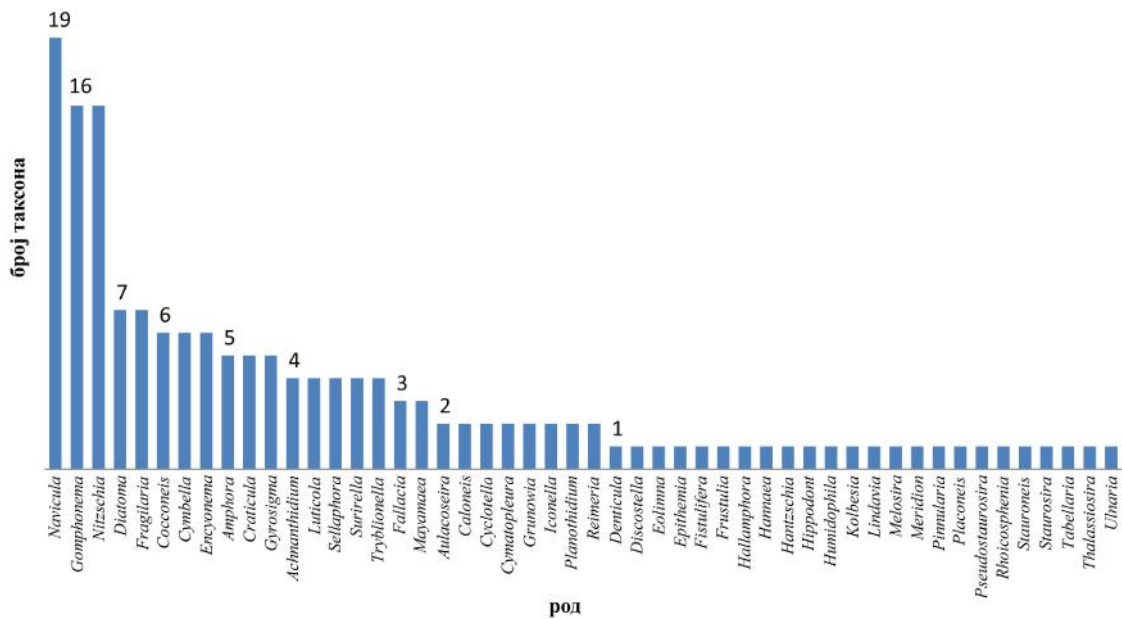
(+ ; +*)
 o ;)

Bacillariophyta						
<i>Achnantheidium affine</i> (Grun.) Czarnecki	ACAF	+				
<i>A. eutrophilum</i> (Lan.-Bert.) Lange-Bertalot	ADEU	+	+	+	+	+
<i>A. minutissimum</i> (Kütz.) Czarnecki	ADMI	+	+	+		
<i>A. pyrenaicum</i> (Hust.) Kobayasi	ADPY	+	+	+	+	+
<i>Amhora copulata</i> (Kütz.) Schoeman & Archibald	ACOP	+	+	+	+	+
<i>A. inariensis</i> Krammer	AINA	+	+	+	+	+
<i>A. minutissima</i> W.Smith	AMNU			+		
<i>A. ovalis</i> (Kütz.) Kützing	AOVA	+	+	+	+	
<i>A. pediculus</i> (Kütz.) Grunow	APED	+	+	+	+	+
<i>Aulacoseira granulata</i> (Ehr.) Simonsen	AUGR					+
<i>A. italica</i> (Ehr.) Simonsen	AUIT					+
<i>Caloneis lancetulla</i> (Sch.) Lange-Bertalot & Witkowski	CLCT	+	+	+	+	+
<i>C. permagna</i> (Bail.) Cleve	CPRM		+			
<i>Cocconeis disculus</i> (Sch.) Cleve	CDIS		+	+		+
<i>C. pediculus</i> Ehrenberg	CPED	+	+	+	+	+
<i>C. placentula</i> Ehrenberg var. <i>placentula</i>	CPLA			+		
<i>C. placentula</i> var. <i>euglypta</i> (Ehr.) Grunow	CPLE	+	+	+	+	+
<i>C. placentula</i> var. <i>lineata</i> (Ehr.) Van Heurck	CPLI	+	+	+	+	+
<i>C. pseudolineata</i> (Geit.) Lange-Bertalot	COPL		+	+		+
<i>Craticula accomoda</i> (Hust.) D.G.Mann	CRAC	+		+		+

<i>C. ambigua</i> (Ehr.) D.G.Mann	CAMB				+	+
<i>C. cuspidata</i> (Kütz.) D.G.Mann	CRCU	+	+	+	+	
<i>C. minusculoides</i> (Hust.) Lange-Bertalot	CMNO				+	
<i>C. subminuscula</i> (Mang.) C.E.Wetzel & L.Ector	CSNU	+	+	+	+	+
<i>Cyclotella atomus</i> Hustedt	CATO					+
<i>C. meneghiniana</i> Kützing	CMEN	+	+	+	+	+
<i>Cymatopleura solea</i> (Bréb.) W. Smith	CSOL	+	+	+	+	+
<i>C. solea</i> var. <i>apiculata</i> (W.Sm.) Ralfs	CSAP		+		+	
<i>Cymbella compacta</i> Østrup	CCMP	+	+	+	+	+
<i>C. excisa</i> Kützing	CAEX	+	+	+		+
<i>C. lanceolata</i> (Agar.) Agardh	CLAN	+	+	+	+	+
<i>C. neocistula</i> Krammer	CNCI	+		+		+
<i>C. subcistula</i> Krammer	CSCI		+			+
<i>C. tumida</i> (Bréb.) Van Heurck	CTUM	+	+	+	+	+
<i>Denticula tenuis</i> Kützing	DTEN	+				+
<i>Diatoma ehrenbergii</i> f. <i>capitulata</i> (Grun.) Lange-Bertalot	DVCA	+	+	+	+	+
<i>D. ehrenbergii</i> Kützing	DEHR	+	+			
<i>D. mesodon</i> (Ehr.) Kützing	DMES		+		+	
<i>D. moniliformis</i> Kützing	DMON	+*	+	+*	+	+
<i>D. moniliformis</i> ssp. <i>ovalis</i> (Fric.) Lange-Bertalot, Rumrich & G.Hofmann	DMOV				+	
<i>D. problematica</i> Lange-Bertalot	DPRO	+	+	+	+	+
<i>D. vulgaris</i> Bory	DVUL	+*	+*	+	+	+*
<i>Discostella pseudostelligera</i> (Hust.) Houk & Klee	DPST					+
<i>Encyonema auerswaldii</i> Rabenhorst	EAUE	+			+	
<i>E. lange-bertalotii</i> Krammer	ENLB			+	+	+
<i>E. leibleinii</i> (C.Agar.) W.J.Silva, R.Jahn, T.A.Veiga Ludwig & M.Menezes	ELEI		+	+	+	+
<i>E. minutum</i> (Hil.) D.G. Mann	ENMI	+		+		+
<i>E. silesiacum</i> (Blei.) D.G. Mann	ESLE	+	+	+	+	+
<i>E. ventricosum</i> (Agar.) Grunow	ENVE	+	+	+	+	+
<i>Eolimna minima</i> (Grun.) Lange-Bertalot	EOMI	+	+	+	+	+
<i>Epithemia sorex</i> Kützing	ESOR					+
<i>Fallacia insociabilis</i> (Kras.) D.G.Mann	FINS					+
<i>F. pygmaea</i> (Kütz.) A.J. Stickle & D.G. Mann ssp. <i>pygmaea</i>	FPYG		+			+
<i>F. subhamulata</i> (Grun.) D.G.Mann	FSBH	+	+	+	+	+
<i>Fistulifera saprophila</i> (Lan.-Bert.& Bon.) Lange-Bertalot	FSAP	+	+	+	+	
<i>Fragilaria acus</i> (Kütz.) Lange-Bertalot	FRAC		+		+	+
<i>F. capucina</i> Desmazières var. <i>capucina</i>	FCAP	+				
<i>F. recapitellata</i> H. Lange-Bertalot & Metzeltin	FRCP	+	+	+	+*	+
<i>F. rumpens</i> (Kütz.) G.W.F.Carlson	FRUM		+		+	
<i>F. vaucheriae</i> (Kütz.) J.B.Petersen	FVAU	+	+	+	+*	+
<i>Frustulia vulgaris</i> (Thwa.) De Toni	FVUL					+
<i>Gomphonema affine</i> Kützing	GAFF		+	+		
<i>G. augur</i> Ehrenberg	GAUG			+		
<i>G. clavatum</i> Reichardt	GCVT				+	
<i>G. elegantissimum</i> Reichardt & Lange-Bertalot	GELG			+	+	+
<i>G. gracile</i> Ehrenberg	GGRA	+	+	+	+	
<i>G. italicum</i> Kützing	GITA			+		
<i>G. micropus</i> Kützing	GMIC	+		+	+	+

<i>G. minutum</i> (Ag.) Agardh	GMIN	+	+	+	+	
<i>G. olivaceum</i> (Horne.) Brébisson	GOLI	+	+	+	+	+
<i>G. parvulum</i> (Kütz.) Kützing	GPAR	+	+	+	+	+
<i>G. pumilum</i> var. <i>rigidum</i> Reichardt & Lange-Bertalot	GPRI	+	+	+	+	+
<i>G. saprophilum</i> (Lan.-Bert. & Reich.) Abraca, R.Jahn, J.Zimmermann & Enke	GSPP	+			+	+
<i>G. sarcophagus</i> Gregory	GSAR					+
<i>G. subclavatum</i> (Grun.) Grunow	GSLC					+
<i>G. supertergestinum</i> Reichardt	GSUT	+			+	
<i>G. tergestinum</i> (Grun.) Fricke	GTER	+	+	+	+	+
<i>Grunowia solgensis</i> (A.Clev.) Aboal	GRSO		+			
<i>G. tabellaria</i> (Grun.) Rabenhorst	NSIT		+		+	
<i>Gyrosigma acuminatum</i> (Kütz.) Rabenhorst	GYAC	+		+	+	
<i>G. attenuatum</i> (Kütz.) Rabenhorst	GYAT		+		+	+
<i>G. kuetzingii</i> (Grun.) Cleve	GYKU	+	+	+	+	+
<i>G. obtusatum</i> (Sull. & Worm.) Boyer	GYOB		+		+	+
<i>G. sciotense</i> (Sull.) Cleve	GSCI	+	+	+	+	+
<i>Hallamphora montana</i> (Kras.) Levkov	HLMO	+	+	+	+	+
<i>Hannaea arcus</i> (Ehr.) R.M.Patrick	HARC	+	+	+	+	+
<i>Hantzschia amphioxys</i> (Ehr.) Grunow	HAMP	+	+	+		+
<i>Hippodonta capitata</i> (Ehr.) Lange-Bertalot, Metzeltin & Witkowski	HCAP		+	+		+
<i>Humidophila contenta</i> (Grun.) Lowe, Kociolek, J.R.Johansen, Van de Vijver, Lange-Bertalot & Kopalová	HCAP		+			+
<i>Iconella linearis</i> (W.Sm.) Ruck & Nakov	ILIN					+
<i>I. tenera</i> (W.Greg.) Ruck & Nakov	ITEN	+	+	+	+	+
<i>Kolbesia gessneri</i> (Hust.) Aboal	KGES	+		+		+
<i>Lindavia comta</i> (Kütz.) Nakov, Gullory, Julius, Theriot & Alverson	LCMT		+			+
<i>Luticola acidoclinata</i> Lange-Bertalot	LACD					+
<i>L. goeppertiana</i> (Blei.) D.G.Mann	LGOE	+	+	+	+	+
<i>L. mutica</i> (Kütz.) D.G.Mann	LMUT		+			
<i>L. nivalis</i> (Ehr.) Mann	LNIV		+			+
<i>Mayamaea atomus</i> (Kütz.) Lange-Bert.	MAAT	+	+	+	+	+
<i>M. cahabaensis</i> Morales & Manoylov	MCAH	+	+	+	+	+
<i>M. permissis</i> (Hust.) K.Bruder & Medlin	MPMI	+	+	+	+	+
<i>Melosira varians</i> C. Agardh	MVAR		+	+	+	+
<i>Meridion circulare</i> (Grev.) C. Agardh var. <i>circulare</i>	MCIR	+	+	+		+
<i>Navicula amphiceropsis</i> Lange-Bertalot & U.Rumrich	NAAM		+			
<i>N. antonii</i> Lange-Bertalot & Rumrich	NANT	+	+	+	+	+
<i>N. capitatoradiata</i> Germain	NCPR	+	+	+	+	+
<i>N. cryptocephala</i> Kützing	NCRY	+	+	+		+
<i>N. cryptotenella</i> Lange-Bertalot	NCTE	+	+	+	+	+
<i>N. erifuga</i> Lange-Bertalot	NERI		+	+	+	+
<i>N. germainii</i> Wallace	NGER	+	+	+	+	+
<i>N. gregaria</i> Donkin	NGRE	+	+	+	+	+
<i>N. lanceolata</i> (Ag.) Ehrenberg	NLAN	+	+	+	+	+
<i>N. radiosa</i> Kützing	NRAD	+		+		
<i>N. reichardtiana</i> Lange-Bertalot	NRCH	+	+	+	+	+
<i>N. rostellata</i> Kützing	NROS	+	+	+	+	+
<i>N. simulata</i> Manguin	NSIA		+			+

<i>N. slesvicensis</i> Grunow	NSLE		+			
<i>N. tripunctata</i> (O.F. Muell.) Bory	NTPT	+	+	+	+	+
<i>N. trivialis</i> Lange-Bertalot	NTRV	+	+	+	+	+
<i>N. vandamii</i> Schoeman & Archibald var. <i>vandamii</i>	NVDA	+				
<i>N. veneta</i> Kützing	NVEN		+	+		+
<i>N. viridula</i> (Kütz.) Ehrenberg	NVIR	+	+	+	+	+
<i>Nitzschia abbreviata</i> Hustedt	NZAB	+	+	+	+	+
<i>N. acicularis</i> (Kütz.) W.Smith	NACI		+			+
<i>N. amphibia</i> Grunow	NAMP	+	+	+	+	+
<i>N. capitellata</i> Hustedt	NCPL	+	+	+	+	+
<i>N. dissipata</i> (Kütz.) Grunow	NDIS	+	+	+	+	+
<i>N. fonticola</i> (Grun.) Grunow	NFON	+	+	+	+	+
<i>N. frustulum</i> var. <i>inconspicua</i> (Grun.) Grunow	NINC	+	+	+	+	+
<i>N. gracilis</i> Hantzsch	NIGR					+
<i>N. heufleriana</i> Grunow	NHEU	+	+	+	+	+
<i>N. intermedia</i> Hantzsch	NINT	+	+		+	+
<i>N. linearis</i> (C. Agar.) W. Smith	NLIN	+	+	+	+	+
<i>N. palea</i> (Kütz.) W. Smith	NPAL	+	+	+	+	+
<i>N. salinarum</i> Grunow	NZSA		+	+	+	+
<i>N. sigmoidea</i> (Nitz.) W.Smith	NSIO	+	+	+	+	+
<i>N. sociabilis</i> Hustedt	NSOC					+
<i>N. tubicola</i> Grunow	NTUB				+	
<i>Pinnularia borealis</i> Ehrenberg	PBOR			+		
<i>Placoneis paraelginensis</i> Lange-Bertalot	PPAE			+		
<i>Planothidium frequentissimum</i> (Lan.-Bert.) Lange-Bertalot	PLFR	+	+	+		+
<i>P. lanceolatum</i> (Bréb. ex Kütz.) Bukhtiyarova	PTLA	+	+	+	+	+
<i>Pseudostaurosira parasitica</i> (Grun.) Morales	PPRS	+	+	+		+
<i>Reimeria sinuata</i> (Greg.) Kociolek & Stoermer	RSIN	+	+	+	+	+
<i>R. uniseriata</i> Sala, Guerrero & Ferrario	RUNI	+	+	+	+	+
<i>Rhoicosphenia abbreviata</i> (C. Ag.) Lange-Bertalot	RABB	+	+	+	+	+
<i>Sellaphora bacillum</i> (Ehr.) D.G.Mann	SEBA	+		+		+
<i>S. capitata</i> D.G.Mann & S.M.McDonald	SECA	+	+			+
<i>S. pupula</i> (Kütz.) Mereschkovsky	SPUP	+	+	+	+	+
<i>S. seminulum</i> (Grun.) Mann	SSEM	+	+	+	+	+
<i>Stauroneis smithii</i> Grunow	SSMI		+			
<i>Staurosira venter</i> (Ehr.) Cleve & J.D.Möller	SSVE					+
<i>Surirella angusta</i> Kützing	SANG		+	+		+
<i>S. brebissonii</i> var. <i>kuetzingii</i> Krammer & Lange-Bertalot	SBKU	+	+	+	+	+
<i>S. minuta</i> Brébisson ex Kützing	SUMI	+	+	+	+	+
<i>S. splendida</i> (Ehr.) Kützing	SSPL			+		+
<i>Tabellaria flocculosa</i> (Roth) Kützing	TFLO		+	+	+	
<i>Thalassiosiras</i> sp. Cleve						+
<i>Tryblionella angustata</i> W. Smith	TANG					+
<i>T. apiculata</i> W.Gregory	TAPI	+	+	+	+	+
<i>T. hungarica</i> (Grun.) Frenguelli	THUN				+	+
<i>T. levidensis</i> W. Smith	TLEV				+	+
<i>Ulnaria ulna</i> (Nitz.) Compère	UULN	+	+	+	+	+



8.

75 %

: *A. pediculus*, *C. placentula* var. *euglypta*, *C. subminuscula*, *D. vulgaris*, *G. parvulum*, *G. pumilum* var. *rigidum*, *N. cryptotenella*, *N. lanceolata*, *N. tripunctata*, *N. abbreviata*, *N. amphibia*, *N. dissipata*, *N. frustulum* var. *inconspicua*, *R. sinuata*, *R. uniseriata*, *R. abbreviata*
U. ulna. *N. dissipata* *A. pediculus*

98%

: *A. pediculus*, *C. subminuscula*, *Cyclotella atomus*, *Diatoma problematica*, *Gomphonema olivaceum*, *N. lanceolata*, *N. abbreviata* *N. dissipata*.

(9). *Diatoma moniliformis*, *D. vulgaris*, *Fragilaria recapitellata*, *F. vaucheriae*, *Navicula capitatoradiata* *Ulnaria ulna*

(1), *D. vulgaris*

(2). *D. vulgaris*

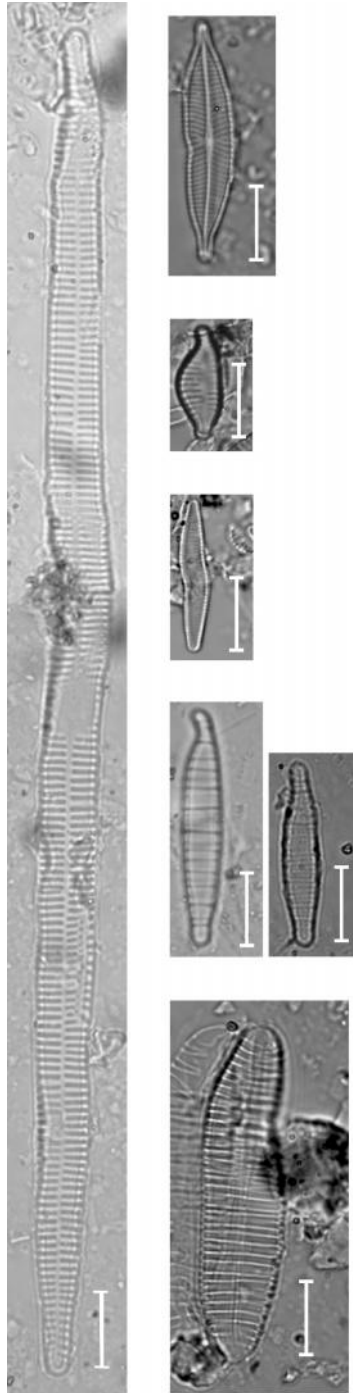
(0,41 %

(0,59 %),

(0,58 %) *U. ulna* (1,98 % *D. vulgaris* 1,78 %

U. ulna 0,20 %). *D. moniliformis* 0,21 %

. *F. recapitellata* *F. vaucheriae* 0,18 %
0,37 %



9.

, 10 μm:

Ulnaria ulna (), *Navicula capitatoradiata* (), *Fragilaria recapitellata* (), *F. vaucheriae* (),
Diatoma moniliformis (,) *D. vulgare* ()

4.1.1.1. Нови налаз за флору алги Србије – *Mayamaea cahabaensis* Morales & Manoylov

Mayamaea

cahabaensis Morales & Manoylov (10),

: Naviculales

: Naviculales incertae sedis

: Mayamaea

: *Mayamaea cahabaensis*

: Morales Manoylov (2009)

: , 5 14 μm 1,5 3,7 μm .

- :

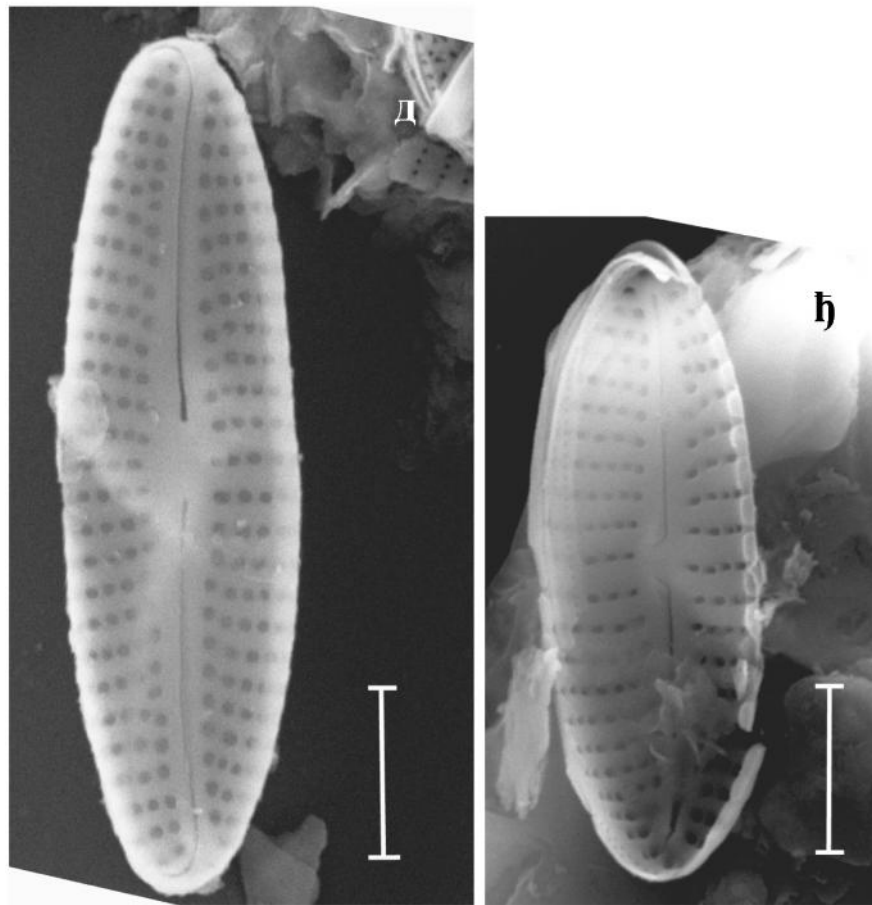
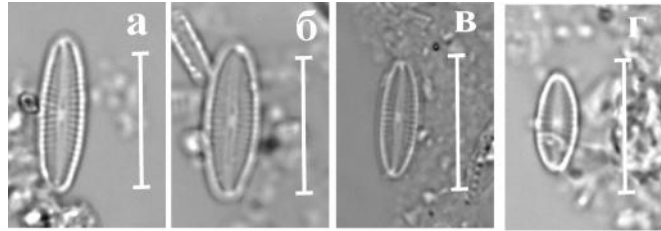
: *Eolimna comperei* Ector, Coste & Iserentant,

Morales Manoylov (2009);

(Falasco Bona, 2013).

: , , ,
(, ,).

: (, , , , ,)
(, , , , ,).



10. *Mayamaea cahabaensis*; , 10 μm
 (, , ,) , 2 μm (;
)

4.1.2. Сезонска динамика

(9),
(45) (42),
(32)
(35).

9. (%)
(2010.)

	32	35	42	45	39	33	35	35	37	
	(%)*									
<i>Amphora inariensis</i>	5,8		7,9							
<i>A. pediculus</i>	24,6	11,3	53,3	21,2	41,0	16,3	32,3	6,6	6,7	
<i>Craticula subminuscula</i>	5,4	8,2		11,0			5,0			
<i>Diatoma moniliformis</i>						8,0				
<i>Fistulifera saprophila</i>		24,2			7,2	5,6				
<i>Gomphonema olivaceum</i>	8,9	6,1		15,0		17,1		6,7		
<i>Mayamaea atomus</i>		5,0								
<i>M. permitis</i>		8,6			10,0		9,2			
<i>Melosira varians</i>								18,7		
<i>Navicula cryptotenella</i>							5,9			
<i>N. gregaria</i>										
<i>N. lanceolata</i>	19,4	7,6	5,2	9,6		21,3		34,9	32,8	
<i>N. reichardtiana</i>										
<i>Nitzschia abbreviata</i>							7,2			
<i>N. dissipata</i>		9,9		5,0			14,2		8,9	
<i>Surirella brebissonii</i> var. <i>kuetzingii</i>	12,8			10,0		6,6			13,4	
(%)	76,9	80,9	66,4	71,7	58,1	74,8	73,9	66,9	61,8	

* 5%

, 16
5 %.

: *A. pediculus*, *G. olivaceum*, *N. lanceolata* *M. varians*,
: *A. pediculus*, *F. saprophila* *N. lanceolata*.

5 %
(80,9 %).
(*Fistulifera saprophila*, 24,2 %),

5 %
(58,1 %).

(*A. pediculus*, 40 %),
5 % (9), *N. cryptotenella*, 2,5 %
() 4,5 % () *N. reichardtiana*, 0,4 ()
4,2% (). *N. frustulum* var. *inconspicua*
0,4 % () 3,9 % ().

(10),
(37 43)
(41).
(30), (31) (34).

2010. , 18
5 %.
A. pediculus, *C. subminuscula* *N. abbreviata*, *A. pediculus*, *C. placentula* var. *euglypta*, *C. subminuscula* *N. abbreviat* ,
A. pediculus, *C. meneghiniana*, *C. atomus*, *N. abbreviat*
N. frustulum var. *inconspicua*.

5 %
(82,6 %).
, *A. pediculus* (35,3 %).
(53,9 %), , *A. pediculus* (15,5%) *C. subminuscula* (14,7 %),

10.

(%)

(, 2010.)

	34	33	34	36	31	37	34	34	41	30	37	37	37	43	39	
	(%)*															
<i>Amphora inariensis</i>					8,7		7,1		9,9						7,2	
<i>A. pediculus</i>	17,7	18,7		45,9	35,3	7,4	21,6	22,1	21,4	23,2	17,9		47,7	15,5		
<i>Cocconeis placentula</i> var. <i>euglypta</i>					5,0		6,4	21,9	7,8							
<i>C. placentula</i> var. <i>lineata</i>								11,2								
<i>Craticula subminuscula</i>	9,9	8,8	11,5	7,4			14,1	8,9		17,5	15,5	7,4	5,6	14,7		
<i>Cyclotella atomus</i>															23,0	
<i>C. meneghiniana</i>						12,9									38,4	
<i>Eolimna minima</i>					6,5	12,9		5,6	7,1							
<i>Fistulifera saprophila</i>	9,0															
<i>Mayamaea cahabaensis</i>						5,1										
<i>M. permitis</i>	5,0															
<i>Navicula cryptotenella</i>										8,0						
<i>Nitzschia abbreviata</i>	17,5	16,2	6,9	15,7	11,8	20,3	13,0		8,4	7,0	22,0	17,3				
<i>N. dissipata</i>													10,8	7,2		
<i>N. fonticola</i>			13,1													
<i>N. frustulum</i> var. <i>inconspicua</i>	14,3	20,1	24,2		9,2	11,0				8,9	11,8	40,1				
<i>N. palea</i>	7,2		15,1							7,2				9,3		
<i>Reimeria uniseriata</i>					6,1											
(%)	80,6	63,7	70,8	69,0	82,6	69,7	62,2	69,7	54,6	71,9	67,2	64,8	64,1	53,9	61,4	

*

5%

5 % (

10),

D. moniliformis

1,4 % () 2,1 %

() *R. sinuata* 0,6 % () 2,5 % ().

G. pumilum var. *rigidum* 0,2 % () 2,8 %

(), *G. tergestinum* 1,8 % () 3,9 % (), *N.*

cryptotenella 0,9 % () 3,6 % (), *N. amphibia* 0,7 %

() 2,3 % () *R. sinuata* 0,4 % () 2,6 %

(). *N. cryptotenella*

0,4 % () 4,4 % () *S. seminulum* 0,2 % () 4,2 % ().

(11),
, (53, 50 57,)
(57).

(40), (43)

(44).

11. (%)

(,

2010.)

	52	50	57	43	49	44	40	46	46	41	43	49	42	47	57	
	(%)*															
<i>Amphora inariensis</i>								13,7								
<i>A. pediculus</i>	6,0			11,8	9,8		35,2	26,2	19,9	6,4				5,4		
<i>Cocconeis placentula</i> var. <i>euglypta</i>	6,4	9,1			6,9		7,6	8,8	7,6							
<i>Cyclotella atomus</i>													40,4			
<i>C. meneghiniana</i>													21,3	8,7		
<i>Diatoma vulgare</i>		6,4	5,5			5,9				5,5	5,0	6,8				
<i>Eolimna minima</i>				8,9		13,6	11,3	6,0								
<i>Mayamaea cahabaensis</i>	9,0				5,7	5,5				9,5	5,1					
<i>Navicula capitatoradiata</i>	9,2	12,6														
<i>N. cryptotenella</i>						8,7				7,9	5,9	6,2		9,3		
<i>N. tripunctata</i>		8,9									6,9	5,5			6,0	
<i>Nitzschia abbreviata</i>			6,2	19,1	14,3	16,2	7,4		6,8	9,5	7,8					
<i>N. amphibia</i>		6,8														
<i>N. dissipata</i>	5,1		35,1	7,9	9,0	13,8			13,3	19,8	21,5	48,9		32,8	34,3	
<i>N. fonticola</i>		11,7	5,0													
<i>N. frustulum</i> var. <i>inconspicua</i>	11,1			10,1	6,9	6,9		8,5	10,3	5,3	25,3					
<i>Ulnaria ulna</i>										6,6						
(%)	46,9	55,4	51,8	57,9	52,5	70,8	61,3	63,2	57,9	70,6	77,5	67,3	61,7	56,2	40,4	

*

5%

2010. , 17
5 %.
: *A. pediculus*, *C. atomus*, *N. abbreviat* , *N. dissipata* *N. frustulum* var. *inconspicua*. *A. pediculus*, *N. capitatoradiat* , *N. abbreviat* , *N. dissipata* *N. frustulum* var. *inconspicua*
, *A. pediculus*, *N. abbreviat*
N. dissipata.
5 %
, (77,5 %). *N. frustulum* var. *inconspicua*
(25,3 %) *N. dissipata* (21,5 %), ,
. (40,4 %),
N. dissipata (34,3 %) *N. tripunctata* (6 %).
, *H. montana* (4,9 %), *D. vulgaris* (4,7 %), *N. cryptotenella* (3,3 %) *G. sciotense* (3,1 %),
. 5 % (
11), *A. eutrophilum* 4,1 %
() 3,7 % (), *M. permitis*, 0,8 %
() 3,5 % (), *N. tripunctata* 0,5 % () 2,8 % (
), *N. palea* 0,7 % () 2,6 % () *S. seminulum* 0,2 %
() 4,4 % (). *R. abbreviata*
0,2 % () 3,5 % () *U. ulna*
0,4 % () 3,3 % ().
N. capitatoradiata 1 % () 4,1 % () *N. amphibia* 0,2 %
() 4,8 % ().
(12),
2010. (51),
2011. (38 37) 2011. (47).
(28),
(25), (27) (24).

2010. 2011. , 26
5 %.

: *A. pediculus*, *D. vulgaris*, *G. olivaceum*
N. dissipata. : *A. pediculus*, *G. olivaceum* *N.*
dissipata, : *A. pediculus*, *D. problematica*, *G. olivaceum*, *N. lanceolata*, *N.*
dissipata *R. abbreviata*, : *A.*
pediculus, *D. problematica*, *G. olivaceum* *N. lanceolata*.

5 %
(86,5 %),
N. lanceolata (37,3 %) *D. problematica*
(36,7 %).

(38, 1 %),

.

A. pyrenaicum *N. capitatoradiata*
3,6 %, *N. tripunctata* 4,4 %, *N. cryptotenella* 4,6 %, *N. frustulum* var. *inconspicua* 4,6 % *N.*
fonticola 4,8 %.

5 % (
12), *A. pyrenaicum*, 0,6 %
() 3,6 % (), *G. pumilum* var. *rigidum*, 0,6 % (
) 4,2 % (), *N. lanceolata* 0,6% () 3,6 %
() *R. abbreviata* 1,2 % () 4,6 % ().
N. capitatoradiata 0,6 % () 3,2 %
(), *N. reichardtiana* 0,8 % () 4,3 % ().
E. silesiacum 0,6 % () 4 % () *R.*
sinuata 0,2 % () 3,9 % (). *F. recapitellata*,
0,6 % () 3,8 % () *N.*
tripunctata 0,7 % () 4,8 % ().

12.

(%)

(2010. , , 2011.)

	28	25	31	30	43	39	27	47	37	38	37	27	28	27	34	36	51	34	32	24
	(%)*																			
<i>Amphora inariensis</i>	13,3												6,5							
<i>A. pediculus</i>	28,6	19,4				33,9	7,0		15,9	7,7	23,3	28,0	29,2				6,9		6,1	
<i>Cocconeis placentula</i> var. <i>euglypta</i>							10,6		5,8	5,0	6,8						7,9			
<i>Cyclotella meneghiniana</i>																	5,2			
<i>Diatoma moniliformis</i> ssp. <i>valis</i>															5,7					
<i>D. problematica</i>		7,2	7,8	36,7		5,1									28,7	30,3				
<i>D. vulgaris</i>							9,4		5,8								9,3			
<i>Gomphonema olivaceum</i>		29,4	19,1	7,2			12,9	5,2				19,7	24,2	23,9		7,5	15,4			9,0
<i>G. tergestinum</i>													5,3							
<i>Hallamphora montana</i>									9,4											
<i>Melosira varians</i>										5,4										
<i>Navicula cryptotenella</i>														11,9	5,7			5,4	8,8	
<i>N. cryptotenella</i>																				
<i>N. lanceolata</i>			16,9	37,3		5,5		41,1			7,2	11,0			5,5	35,7				47,2
<i>N. tripunctata</i>			10,6		14,2	7,1			5,1	12,1	7,6			17,6	9,2			7,7	9,2	
<i>N. abbreviata</i>	6,6								6,5											
<i>Nitzschia amphibia</i>					6,0															
<i>N. dissipata</i>		19,6	7,2		26,9	16,1		5,0		33,1				48,1	18,9		8,7	59,3	40,7	
<i>N. frustulum</i> var. <i>inconspicua</i>	14,3				12,3															
<i>Reimeria sinuata</i>												5,0	11,4							
<i>R. uniseriata</i>													7,2							
<i>Rhoicosphenia abbreviata</i>							17,6				10,0	6,7								
<i>Surirella brebissonii</i> var. <i>kuetzingii</i>			6,4	5,2		7,3		9,8		6,7										11,8
<i>S. minuta</i>			11,9																	
<i>Ulnaria ulna</i>							10,2													
(%)	62,8	75,5	79,9	86,5	59,3	75,0	67,6	61,0	48,6	70,0	74,5	74,8	83,4	77,6	81,3	81,3	38,1	72,4	64,8	68,0

*

5%

4.1.3. Вредности Shannon-овог индекса диверзитета

Shannon-ovog indeksa dиверзитета

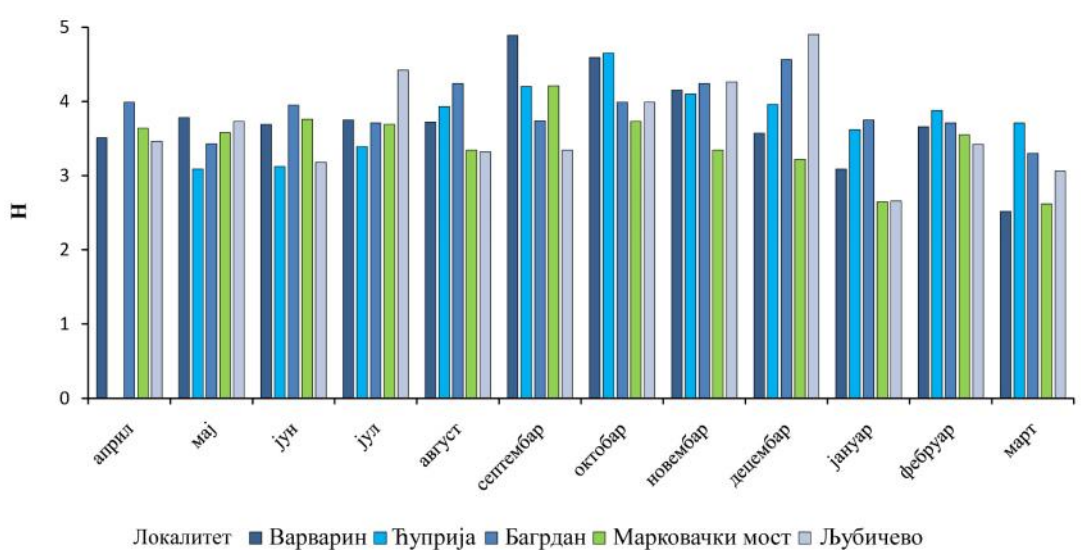
12 (11)

(4,89) (4,21)

(4,65) (4,56)

(4,9), Shannon-

(2,62) (2,66) (2,52) (3,3)



11. Shannon-ovog indeksa dиверзитета (H)

Shannon-ovog indeksa dиверзитета

2010.

2011.

3,18 (,) 3,78 (,)

3,34 (,)

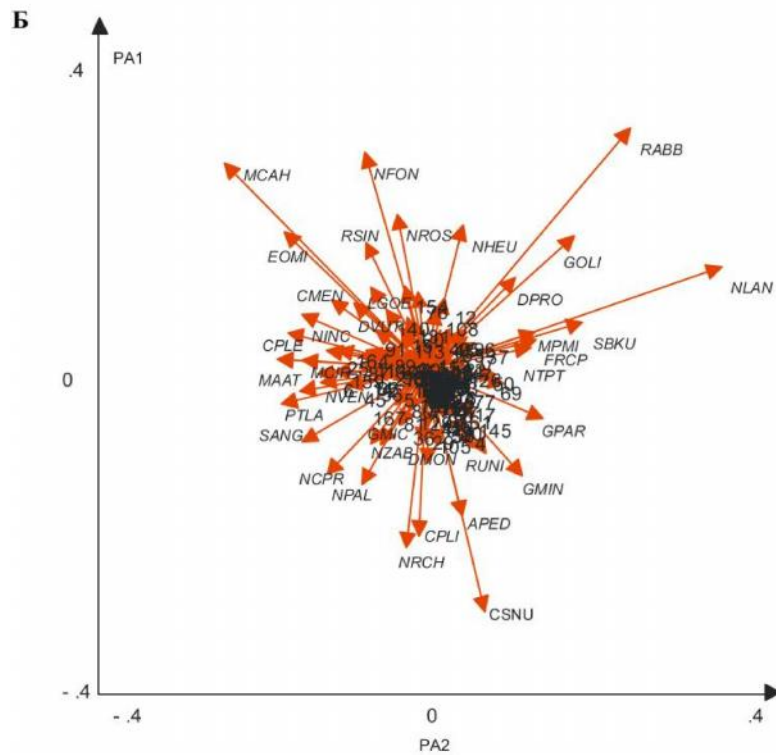
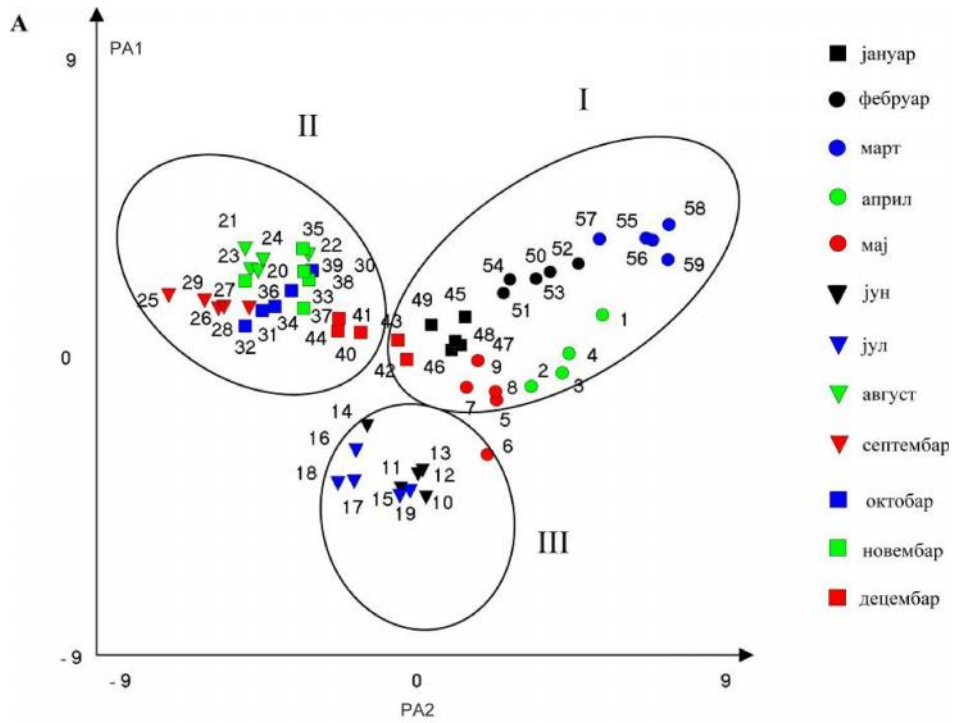
) 4,89 (,), 4.

Sh non-
(4,42) (4,24)
(3,99),
(4,9)

2011. .

4.1.4. Статистичка анализа бентосне заједнице силикатних алг Велике Мораве

(DAPC) 12
„FLORA”, (2010. 2011.).
DAPC , (12),
(12).
(PA1) 31,2% , (PA2)
18,8%. :
(I),
(II). ,
(III), .
, , : *D. problematica*, *G. olivaceum*, *N. lanceolata*, *N. tripunctata*, *R. abbreviata* *S. brebissonii* var. *kuetzingii*.
, : *C. placentula* var. *euglypta*, *C. meneghiniana*, *E. minima*, *M. cahabaensis*, *N. fonticola* *N. frustulum* var. *inconspicua*,
, *R. sinuata*
, *A. pediculus*, *C. subminuscula*, *N. abbreviata* *N. palea* .



12. DAPC

(PA1 31,2 % PA2 18,8 %

); .

; .

(

5)

4.2. Физичке и хемијске карактеристике воде Велике Мораве

Година	Температура (°C)	pH	Концентрација (µS/cm)
2010.	(24,8 °C)	(8,5)	500 µS/cm
2011.	(23 °C)	(8,4)	377 µS/cm
2011.	(25,2 °C)	(7,6 - 8)	326 µS/cm
2011.	(3,4 °C - 4,2 °C)		315 µS/cm
2011.	(2 °C)		569 µS/cm
2010.			512 µS/cm
2010.			2010.

(9,6 mg/l, 9,3 mg/l 9,1 mg/l),
(12,6 mg/l),
(11,2 mg/l 13,3 mg/l).
(89 %) (80 %
96 %), (123 %, 165 % 152 %).

(1102 mg/l, 119 mg/l 543 mg/l),
(7 mg/l), (3
mg/l) (3 mg/l).

0,01 mg/l 0,59 mg/l , 0,01 mg/l ,
0,56 mg/l 0,01 mg/l 0,9 mg/l .
0,2 mg/l
1,1 mg/ ,
0,1 mg/l 2,1 mg/l 0,3
2,1 mg/l .
0,007 mg /l 0,08 mg/l , 0,007 mg/l
0,3 mg/l
0,02 mg/l 0,11 mg/l .

0,0025
mg/l 0,144 mg/l , 0,039 mg/l 0,197 mg/l
0,029 mg/l 0,185 mg/l .

29
mg/l 47 mg/l .
(22 mg/l 19 mg/l), (96 mg/l
84 mg/l).

175 mg CaCO ₃ /l	276 mg CaCO ₃ /l	2010.,
175 mg CaCO ₃ /l	2011. 292 mg CaCO ₃ /l	2010.
177 mg CaCO ₃ /l	314 mg CaCO ₃ /l	2010.

.

.

.

0,28 µg/l (), 0,01 µg/l () 0,05 µg/l ()

() 0,1 µg/l (,) 0,025 µg/l

7,4 µg/l () 46 µg/l (),

11,5 µg/l () 34,2 µg/l () 3,9 µg/l

() 37,9 µg/l ().

0,05 µg/l () 0,25 µg/l (, ,

,) 2 µg/l (,),

0,025 µg/l () 0,7 µg/l ().

90 µg/l () 490 µg/l (),

60 µg/l () 210 () 50 µg/l ()

210 µg/l (). 5 µg/l

() 40 µg/l (), 20 µg/l (,

) 110 µg/l () 5 µg/l (, , ,

) 130 µg/l ().

0,05 µg/l () 0,1 µg/l ()

0,05 µg/l (, , ,) 0,2 µg/l

() 0,05 µg/l ().

13.

		2010.									2011.	
(T)	°C	12,0	15,5	20,5	22,5	23,5	18,0	10,0	9,0	6,0	2,0	4,0
pH		7,9	8,0	8,3	8,4	8,4	8,4	8,0	8,0	8,0	8,0	8,1
(EC)	µS/cm	377	379	420	480	500	467	447	407	395	434	442
(DO)	mg/l	10,5	9,7	9,6	10,1	10,4	9,9	10,8	10,5	11,2	12,6	12,4
(O ₂ %)	%	98	97	107	118	123	104	95	90	89	90	94
	mg/l	161	70	1102	21	26	11	7	20	50	28	17
(NH ₄ ⁺)	mg/l	0,37	0,01	0,59	0,06	0,02	0,04	0,02	0,14	0,06	0,27	0,27
(NO ₃ ⁻)	mg N/l	0,50	0,50	1,10	0,40	0,40	0,20	0,50	0,20	0,90	0,20	0,20
(NO ₂ ⁻)	mg N/l	0,009	0,030	0,063	0,040	0,007	0,080	0,053	0,050	0,035	0,031	0,018
(SO ₄ ²⁻)	mg/l	31	29	47	37	35	40	38	40	35	42	45
(PO ₄ ³⁻)	mgP/l	0,091	0,079	0,125	0,074	0,015	0,144	0,117	0,094	/	0,064	0,0025
(Ca ²⁺)	mg/l	56	54	54	58	36	66	71	56	58	44	57
(Mg ²⁺)	mg/l	17,0	15,1	17,9	23,0	21,0	27,0	16,0	24,0	19,4	24,0	28,0
(TH)	mg CaCO ₃ /l	210	196	208	237	175	276	245	240	226	209	256
(Zn)	µg/l	18,0	19,9	31,0	31,7	11,5	12,0	24,8	17,6	30,6	34,20	31,8
(Cd)	µg/l	0,10	0,07	0,17	0,25	0,08	0,28	0,05	0,06	0,10	0,12	0,20
(Pb)	µg/l	2,00	0,60	0,25	1,80	0,50	0,05	0,05	0,90	0,90	0,80	0,25
(Cu)	µg/l	30,0	10,9	8,7	11,0	13,8	13,5	9,5	24,7	16,6	5,5	8,0
(Fe)	µg/l	110	130	160	160	100	170	490	90	130	140	140
(Mn)	µg/l	20	5	5	5	5	5	5	5	20	40	40
(Hg)	µg/l	0,05	0,05	0,05	0,05	0,10	0,05	0,05	0,05	0,05	0,05	0,05
(Ni)	µg/l	3,0	4,8	2,9	6,0	3,2	4,2	4,8	3,2	5,4	5,1	5,0
(As)	µg/l	4,0	2,8	2,3	4,5	4,5	4,3	3,2	2,1	2,4	2,8	2,5

14.

		2010.								2011.		
(T)	°C	11,1	17,0	18,2	24,8	24,2	20,0	10,2	4,2	5,0	3,4	6,6
pH		8,0	8,3	8,3	8,4	8,4	8,5	8,0	8,0	8,0	8,0	8,1
(EC)	µS /cm	375	406	441	444	435	512	451	370	431	376	326
(DO)	mg/l	9,9	10,0	9,3	7,9	13,7	9,7	9,8	10,8	10,2	11,2	11,4
(O ₂ %)	%	90	104	99	96	165	107	87	83	80	84	93
	mg/l	31	37	119	68	18	6	3	6	7	14	55
(NH ₄ ⁺)	mg/l	0,01	0,13	0,01	0,01	0,44	0,42	0,09	0,11	0,56	0,14	0,08
(NO ₃)	mg N/l	1,50	0,10	1,20	0,50	0,80	0,50	0,25	2,10	2,00	1,90	1,40
(NO ₂)	mg N/l	0,016	0,075	0,087	0,033	0,140	0,310	0,268	0,038	0,098	0,047	0,007
(SO ₄ ²⁻)	mg/l	31	22	53	28	96	46	42	43	38	38	26
(PO ₄)	mgP/l	0,075	0,081	0,140	0,039	0,077	0,161	0,197	0,108	0,106	0,121	0,095
(Ca ²⁺)	mg/l	56	59	73	59	63	69	61	64	55	50	50
(Mg ²⁺)	mg/l	21,2	12,5	16,7	22,0	18,0	29,0	25,0	12,0	24,0	23,0	12,0
(TH)	mg CaCO ₃ /l	227	198	250	239	232	292	255	210	234	220	175
(Zn)	µg/l	24,0	17,0	46,0	8,2	18,1	44,7	10,8	12,8	20,2	19,8	7,4
(Cd)	µg/l	0,10	0,04	0,09	0,01	0,01	0,05	0,04	0,07	0,07	0,04	0,04
(Pb)	µg/l	0,50	0,25	0,25	0,25	0,25	0,70	0,50	0,60	0,70	0,25	0,25
(Cu)	µg/l	21,0	9,9	9,7	11,5	7,6	24,2	7,3	3,7	13,2	4,1	3,6
(Fe)	µg/l	140	140	150	150	70	210	60	70	110	160	80
(Mn)	µg/l	70	110	30	20	20	10	10	20	10	10	10
(Hg)	µg/l	0,05	0,10	0,05	0,05	0,10	0,20	0,20	0,20	0,05	0,05	0,05
(Ni)	µg/l	2,0	3,7	4,1	3,6	3,1	6,5	3,6	3,1	3,4	4,6	2,0
(As)	µg/l	3,0	2,7	3,8	3,2	3,5	4,2	2,3	1,7	2,1	2,0	1,9

15.

		2010.									2011.		
(T)	°C	11,2	12,6	19,7	25,8	25,2	20,0	12,3	12,2	4,3	6,7	4,2	9,4
pH		7,6	7,9	7,8	8,4	8,4	8,4	8,0	7,9	7,7	7,9	7,8	7,9
(EC)	µS/cm	315	359	430	458	371	551	569	499	452	420	452	341
(DO)	mg/l	11,1	10,1	9,1	9,8	12,4	10,4	9,4	11,6	12,8	11,7	13,3	11,1
(O ₂ %)	%	101	96	98	121	152	115	98	108	98	96	102	97
	mg/l	395	192	543	36	12	10	5	11	30	3	5	49
(NH ₄ ⁺)	mg/l	0,05	0,17	0,25	0,71	0,95	0,09	0,02	0,13	0,21	0,31	0,13	0,01
(NO ₃ ⁻)	mg N/l	1,7	2,1	0,5	0,5	1,0	2,2	0,5	0,3	1,7	0,5	0,5	1,6
(NO ₂ ⁻)	mg N/l	0,108	0,020	0,078	0,040	0,062	0,118	0,062	0,079	0,045	0,097	0,037	0,052
(SO ₄ ²⁻)	mg/l	29	19	47	37	84	44	60	52	35	44	44	33
(PO ₄ ³⁻)	mgP/l	0,091	0,094	0,122	0,029	0,045	0,176	0,159	0,149	0,104	0,185	0,0970	0,115
(Ca ²⁺)	mg/l	45	55	65	59	45	70	79	70	69	63	66	46
(Mg ²⁺)	mg/l	13,9	13,1	14,0	26,0	24,0	26,0	28,0	23,0	27,0	21,0	20,0	18,0
(TH)	mg CaCO ₃ /l	177	191	221	256	223	283	314	271	285	244	247	188
(Zn)	µg/l	13,0	37,9	8,3	9,6	14,8	3,9	7,1	6,6	11,5	11,3	15,4	4,9
(Cd)	µg/l	0,100	0,090	0,125	0,125	0,125	0,125	0,025	0,125	0,040	0,040	0,060	0,030
(Pb)	µg/l	2,00	0,60	0,50	0,25	0,60	0,25	0,25	0,25	0,60	0,60	0,50	0,60
(Cu)	µg/l	33,0	12,0	14,7	10,2	14,2	7,0	6,2	10,5	7,8	11,2	9,2	3,4
(Fe)	µg/l	170	160	100	130	90	210	80	50	70	110	130	120
(Mn)	µg/l	50	130	10	5	20	5	10	5	10	5	5	5
(Hg)	µg/l	0,05	0,10	0,05	0,05	0,20	0,10	0,20	0,10	0,40	0,05	0,05	0,05
(Ni)	µg/l	3,0	4,5	2,8	2,9	4,1	4,9	3,4	3,0	3,8	3,8	4,7	3,4
(As)	µg/l	3,0	2,2	3,1	3,0	2,8	4,3	3,2	2,2	2,3	2,3	2,1	2,2

4.3. Статистичка анализа утицаја физичких и хемијских параметара на састав заједнице бентосних силикатних алги реке Велике Мораве

, . 34 ,
 (2010. 2011.), -
 Mantel- ()
 (r=0,121; p=0,175)
 (FS) (p 0,05) ()
 -) ()
), :
 (, TH, NO₃ , Mg²⁺, pH EC) (As Pb) (16).

16. (p 0,05)

		F	
(T)	0,293	3,206	0,001
(TH)	0,2399	2,58	0,001
(NO ₃)	0,1752	1,845	0,006
(Mg ²⁺)	0,1578	1,653	0,003
pH	0,1287	1,336	0,037
(EC)	0,1189	1,23	0,011
(As)	0,1728	1,818	0,03
(Pb)	0,1638	1,719	0,037

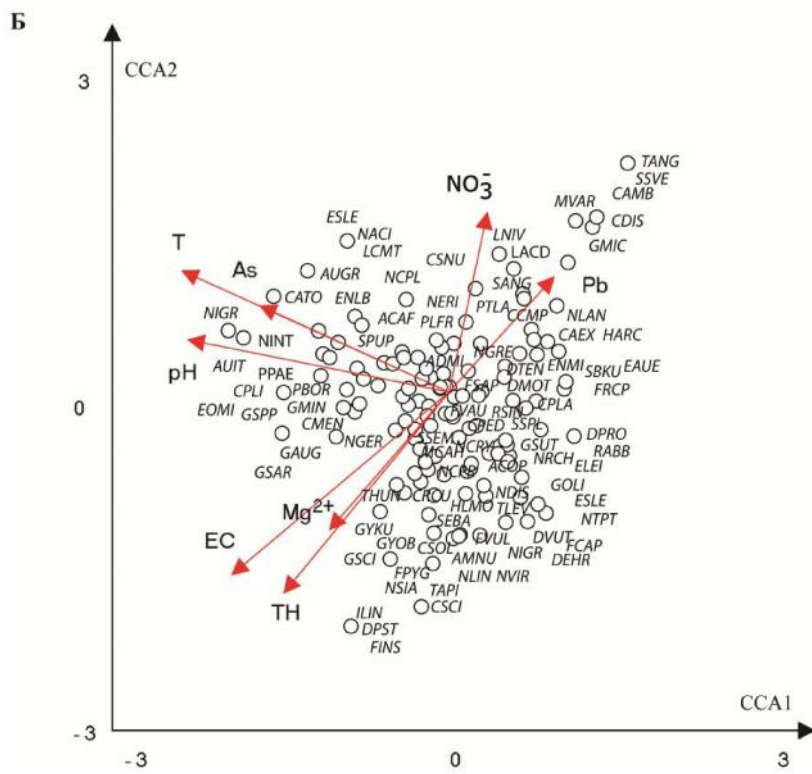
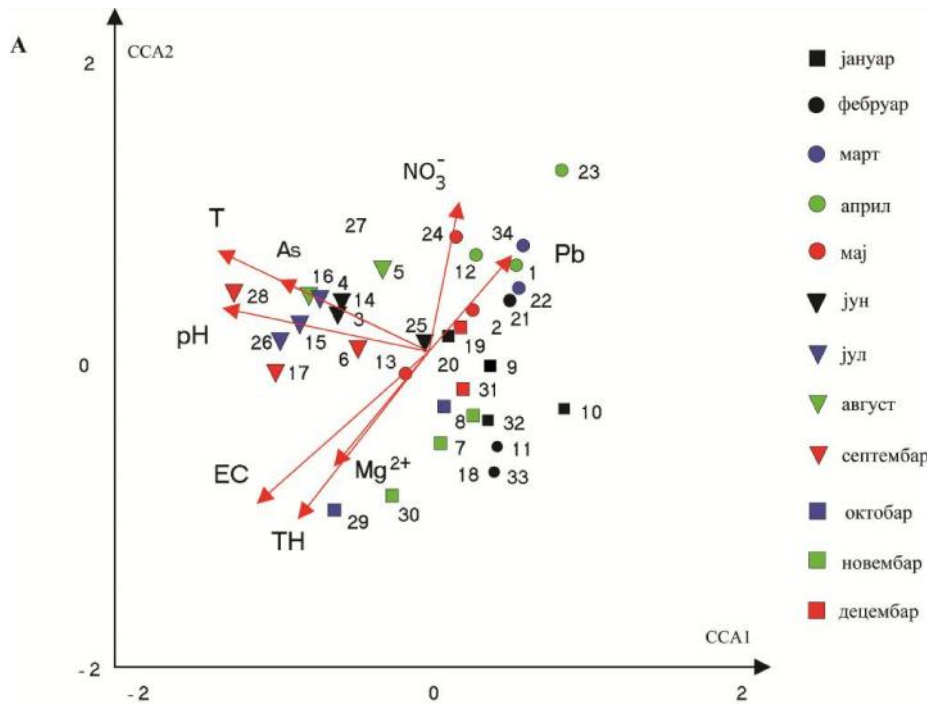
(CCA)

(13 13).

CCA , 29 % ,
 pH, .
 , .
 ()
),
 ().

2010. , 2010.
 2010. .
C. placentula
var. lineata, N. capitellata, E. minima, A. granulata, L. comta
C. atomus C. meneghiniana. pH, As T,
 2010. 2011. 2010.
 2011. .
 , CCA *D. problematica, E. leibleinii, E. silesiacum*
G. olivaceum, N. reichardtiana N. tripunctata.

CCA , 23,2 % ,
 . .
 NO₃ Pb, 2011.
 2010. .
 ,
F. recapitellata, H. arcus, M. circulare, N. lanceolata S.
brebissonii var. kuetzingii. CCA , Pb NO₃
 , ,
 .
Gyrosigma sciotense G. kuetzingii.



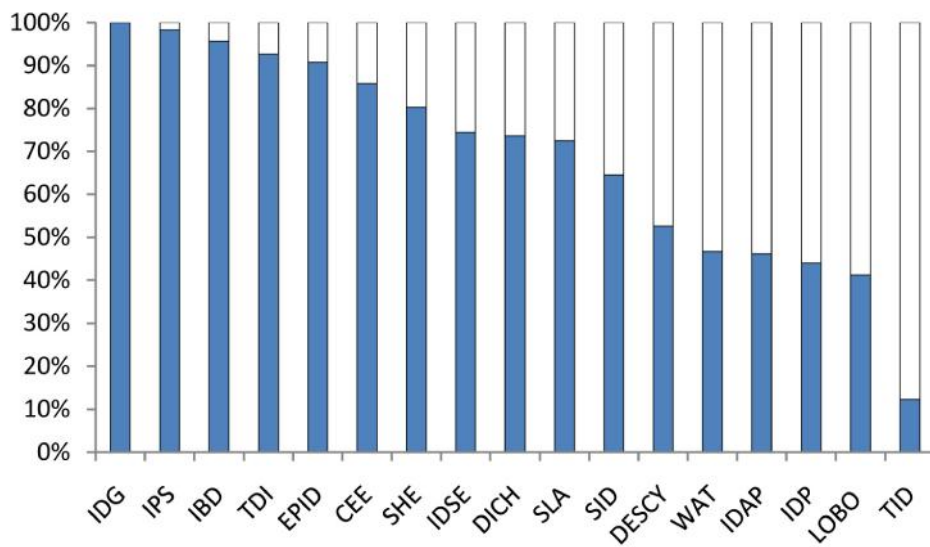
13. CCA

(; Monte Carlo ; CCA1 29% CCA2 23,2%); . ; . (8)

**4.4. Квалитет вод
индекса**

елике Мораве на основу дијатомних

,
12
,
(14).
80%
(15 26,
80%).



14. (%)

2010.

(15). IBD, IPS DESCY

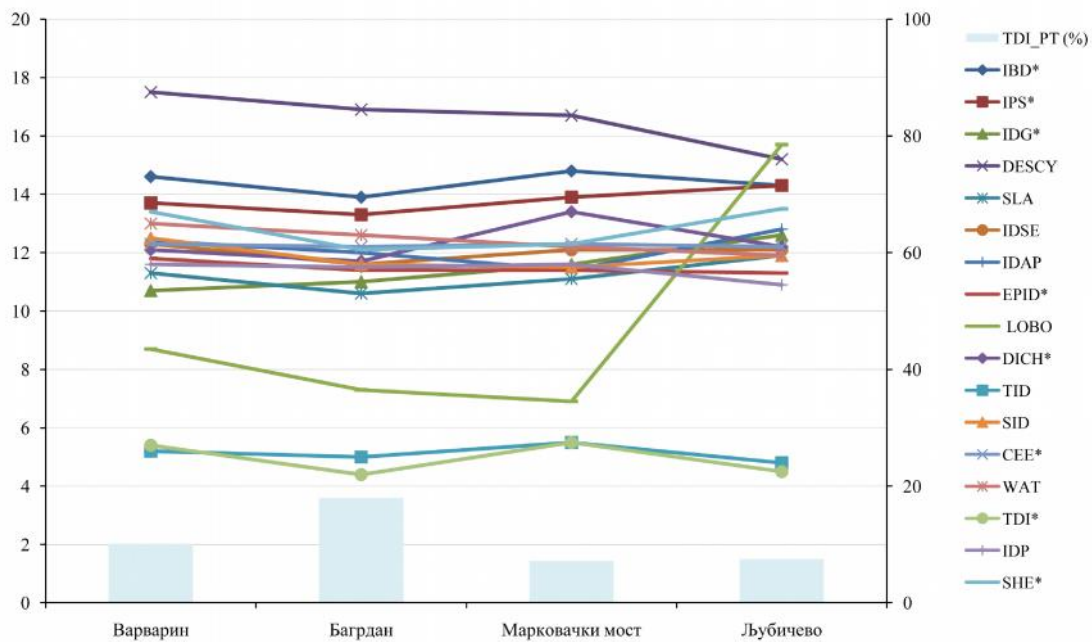
(IDG, SLA, IDSE, IDAP, EPID, DICH, SID,

CEE, WAT IDP)

(17). LOBO TID

(TDI_PT) 20 % ()
), TDI

(17).



15.

2010.

() TDI_PT (%) ()

2010.

(16).

DESCY

EPID, LOBO, DICH, SID, TID SHE (17).

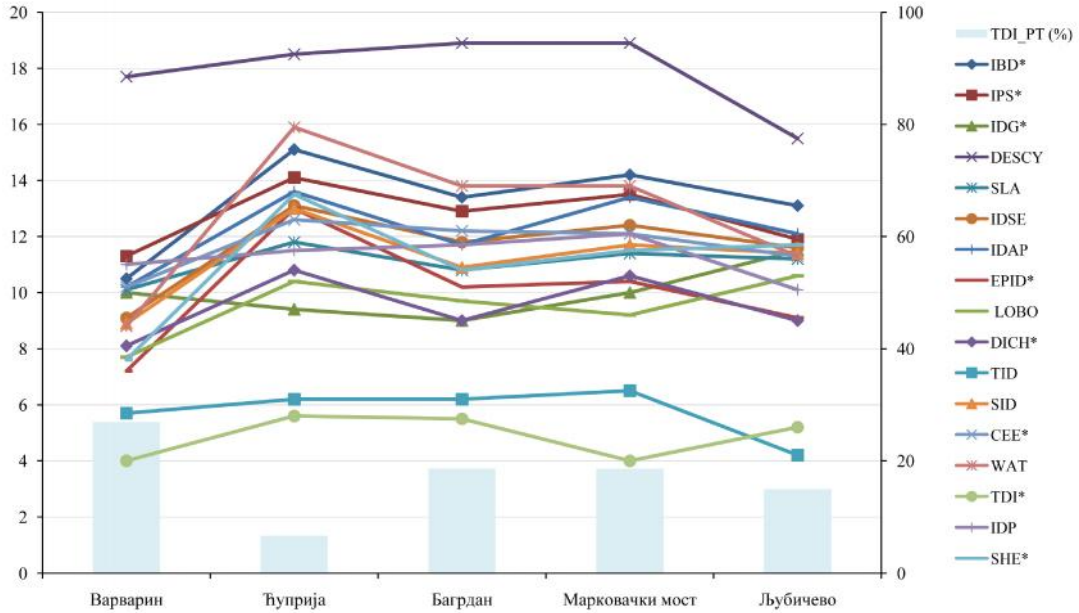
TDI

TDI_PT 27 %,

TDI_PT

20 %,

(17).



16.

2010.

() TDI_PT (%) ()

2010.

17).

(). , (IBD DESCY, (IPS, SLA, IDSE, IDAP, EPID, LOBO, SID, CEE, IDP SHE), (IDG, DICH, TID)

(17). TDI

(TDI_PT 18 %,

) (TDI_PT 23,9 %,

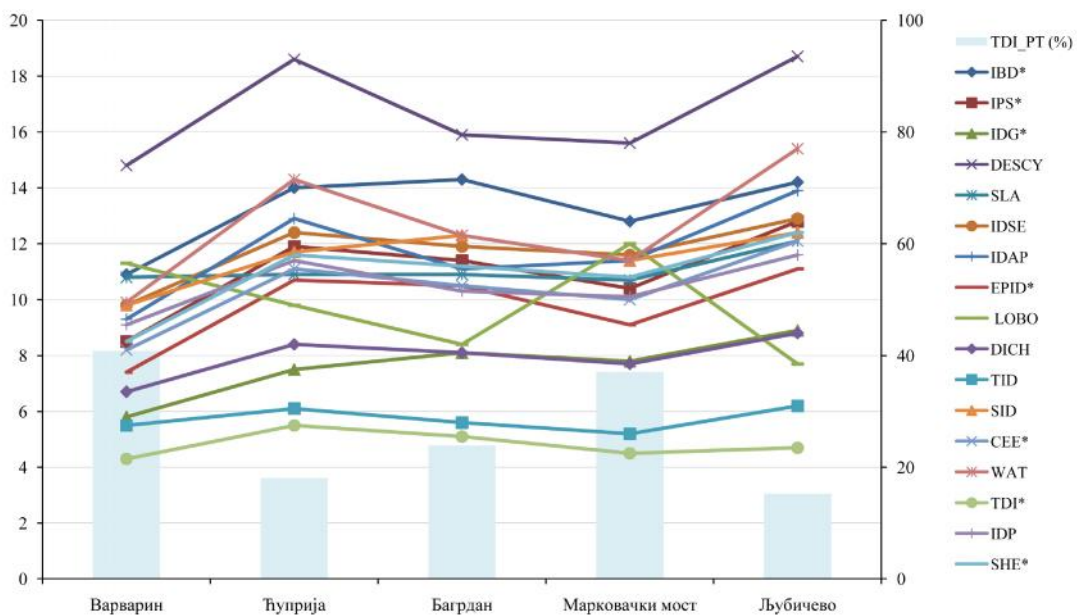
).

TDI

TDI_PT

(40,8 % 37,1 %)

(17).



17.

2010.

() TDI_PT (%) ()

2010.

(18). DESCY

IBD, IPS WAT

(17).

(SLA, IDSE, IDAP, EPID,

SID, CEE, IDP SHE),

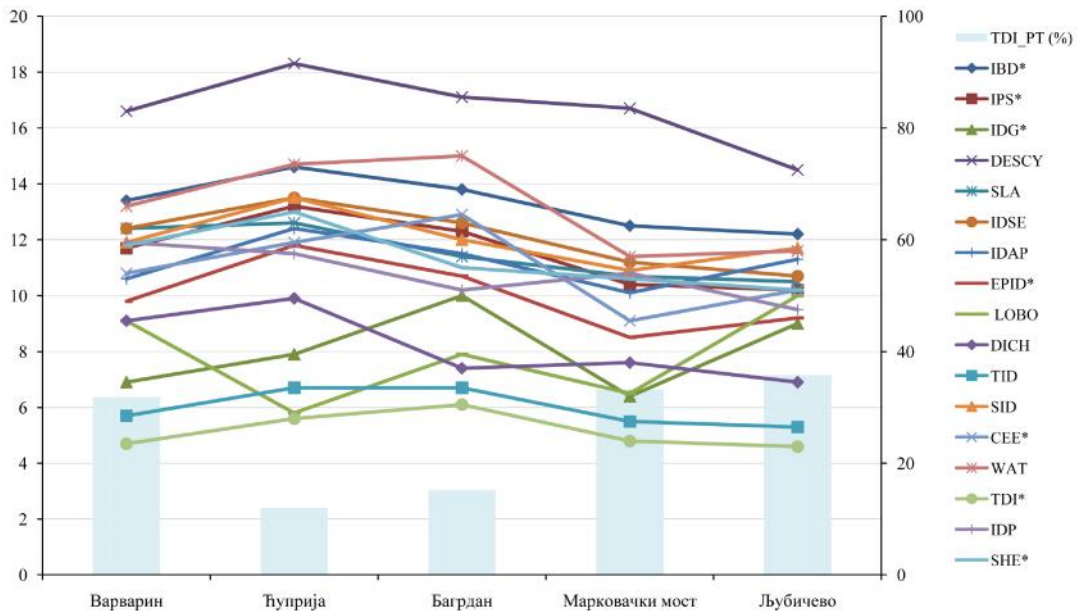
(LOBO, DICH TID).

TDI

(TDI_PT < 20 %).

TDI_PT 31,9 %, 33,2 % 35,8 %, ,

(17).



18.

2010.

() TDI_PT (%) ()

2010.

19).

, DESCY

(

(

)

,

(

17).

(IBD, SLA, IDSE, SID, WAT, IDP

SHE),

(IPS, IDG, IDAP, EPID, LOBO, DICH, TID CEE).

TDI

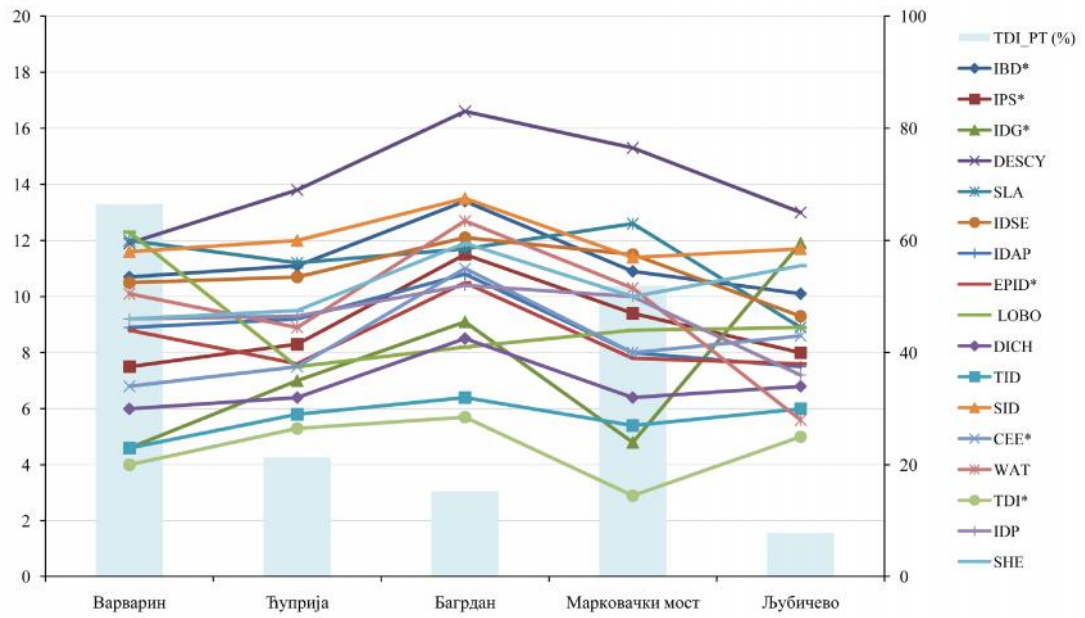
(TDI_PT < 20 %).

,

TDI_PT (21,3 %, 66,5 % 51,9 %,

)

(17).



19.

2010.

() TDI_PT (%) ()

2010.

.

(20).

IBD DESCY () (20).

a (IPS, IDG, SLA, IDSE,

IDAP, EPID, SID, CEE, IDP SHE)

(LOBO, DICH, TID).

TDI

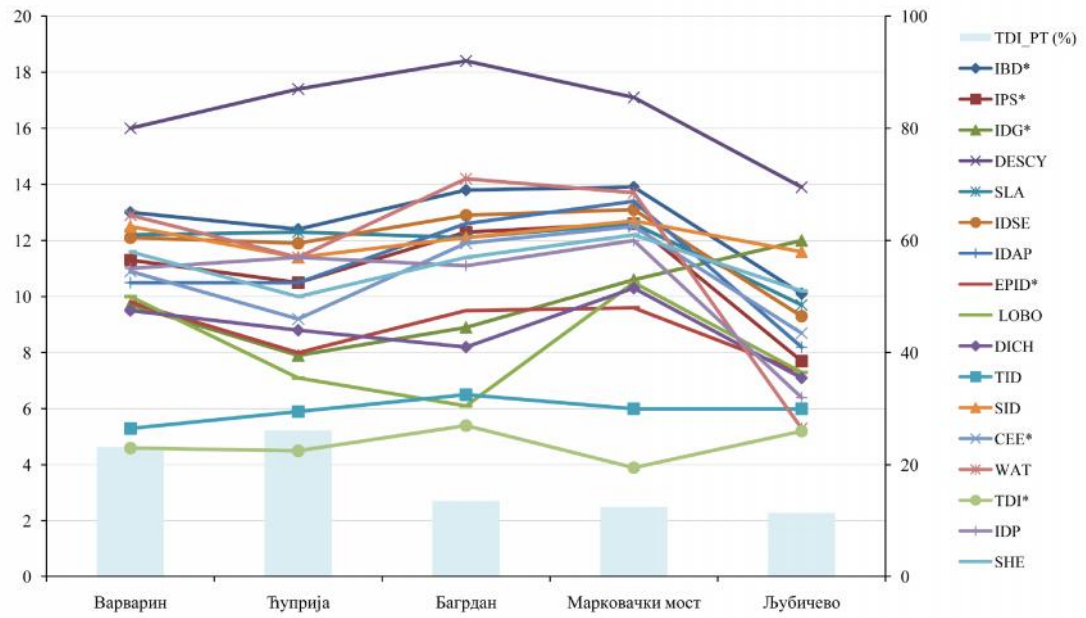
TDI_PT 23,2 %

26,1 %

(TDI_PT < 20 %):

TDI

(17).



20.

2010.

() TDI_PT (%) ()

2010.

(21).

IBD, IPS, IDSE, WAT DESCY

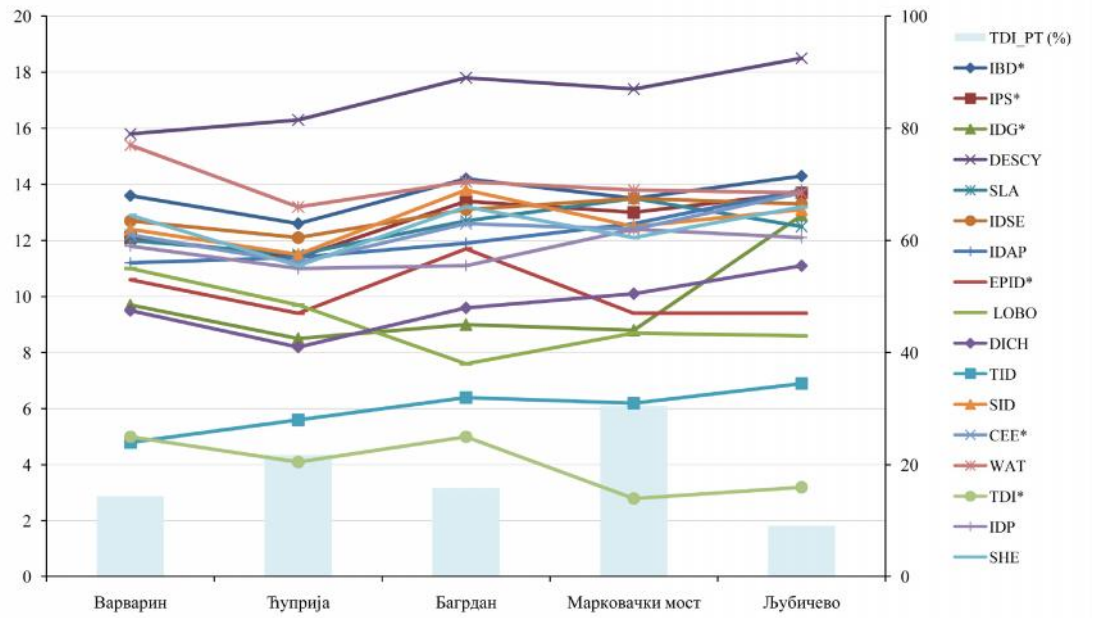
(DESCY) (17). (IDG, SLA, IDAP, EPID, DICH, SID, CEE, IDP SHE), LOBO TID

TDI ,

TDI_PT

(21,8 %)

(30,5 %) (17).



21.

2010.

() TDI_PT (%) ()

2010.

,

(22). DESCY

, IBD, IPS, SLA, IDSE,

IDAP WAT

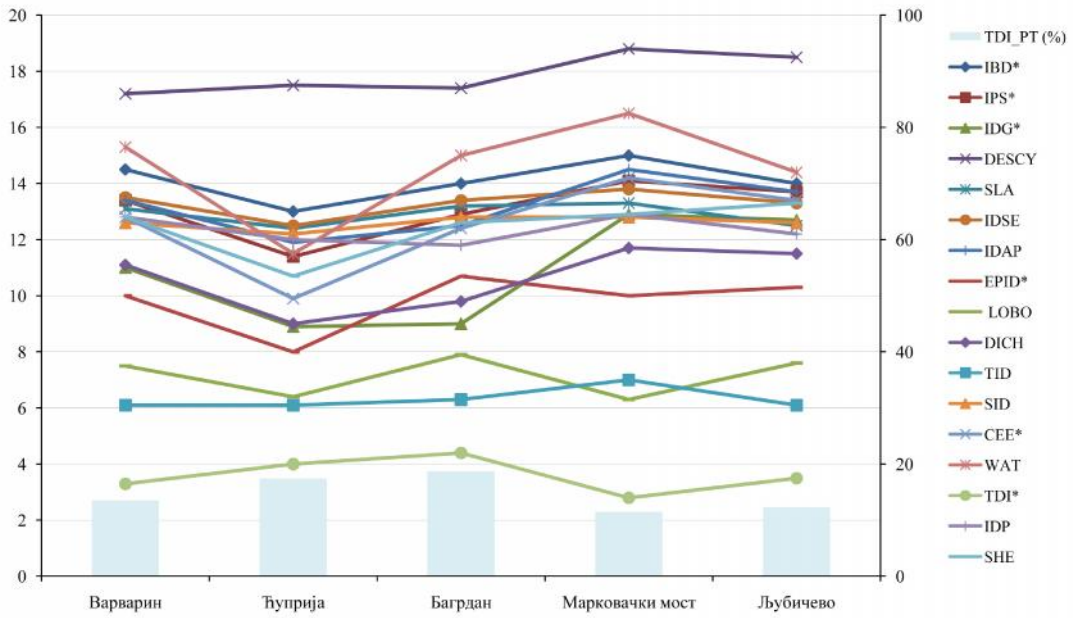
(17).

IDAP, DICH, SID, CEE, IDP SHE

LOBO TID . TDI

(TDI_PT < 20 %,)

)(17).



22.

2010.

() TDI_PT (%) ()

2010.

,

(23). DESCY

(17).

IBD, IPS,

IDG, SLA,

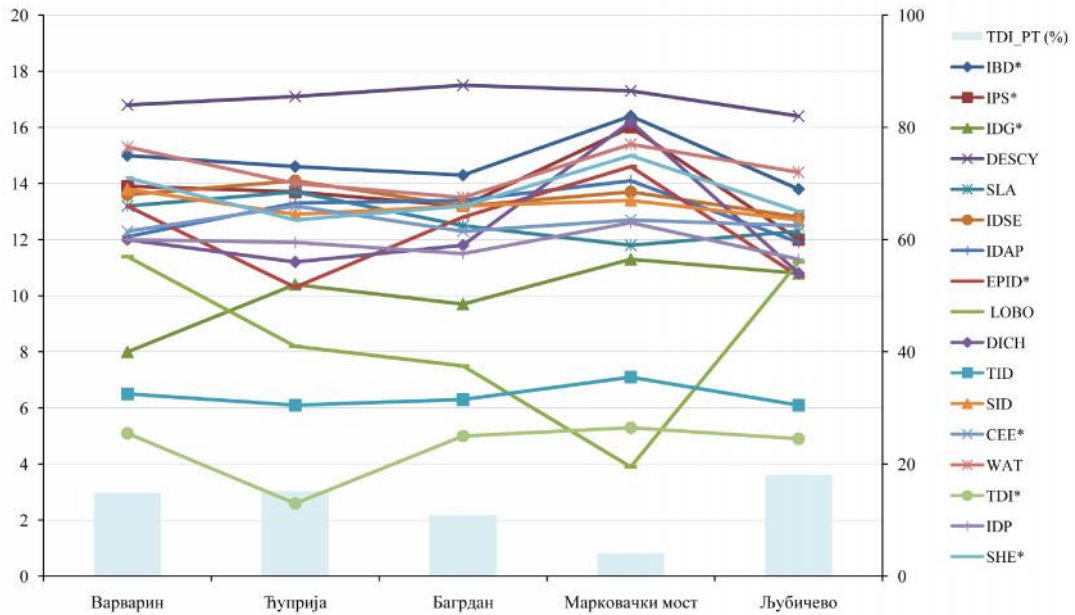
IDSE, IDAP, SID, WAT SHE

EPID, DICH, CEE IDP

TDI

(TDI_PT < 20 %,

)(17).



23.

2010.

() TDI_PT (%) ()

2010.

(IBD, IPS, SLA, IDSE, IDAP, SID, WAT

SHE)

(24).

DESCY

(

17).

EPID, DICH, CEE IDP

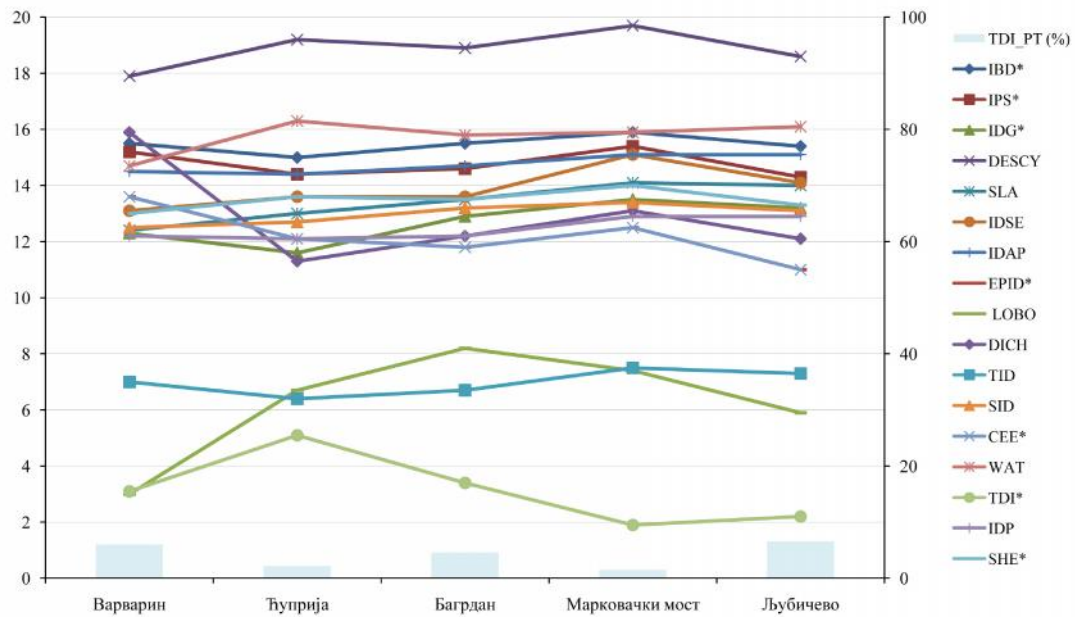
, LOBO TID

TDI

TDI

(TDI_PT < 20 %,

)(17).



24.

2011.

() TDI_PT (%) ()

2010.

(25).

(DESCY)

(IBD, IPS,

IDSE, IDAP, EPID, DICH, CEE, WAT SHE) (17).

IDG, SLA,

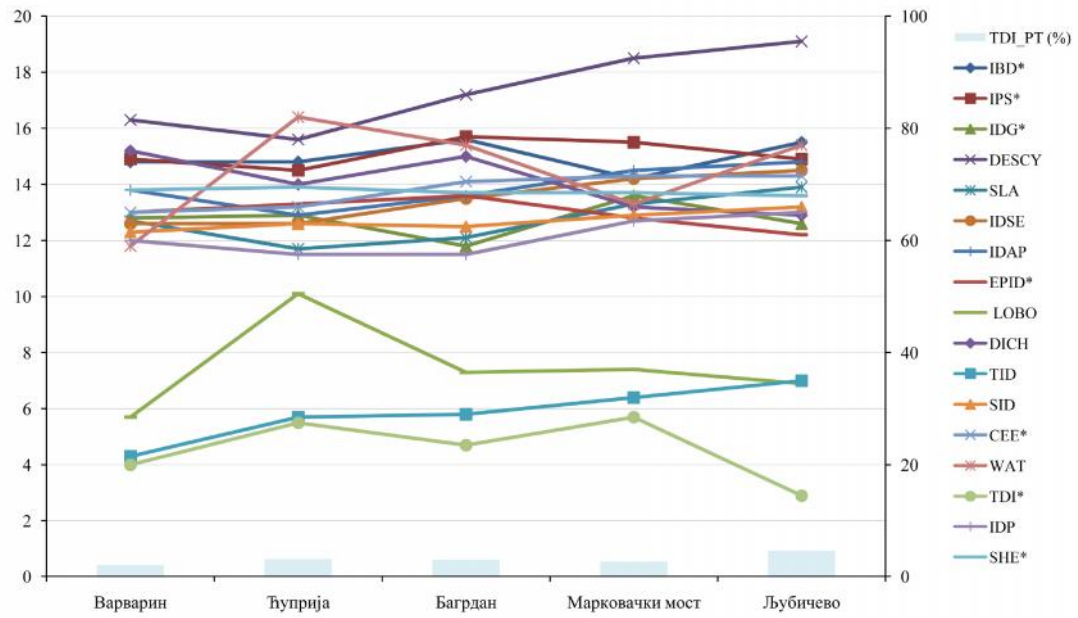
SID IDP

TDI

TDI

(TDI_PT < 20 %,

)(17).



25.

2011.

() TDI_PT (%) ()

2011.

(26).

IBD, IPS, DESCY, DICH, CEE SHE

, IDG, SLA, IDSE, IDAP, EPID, SID, WAT IDP

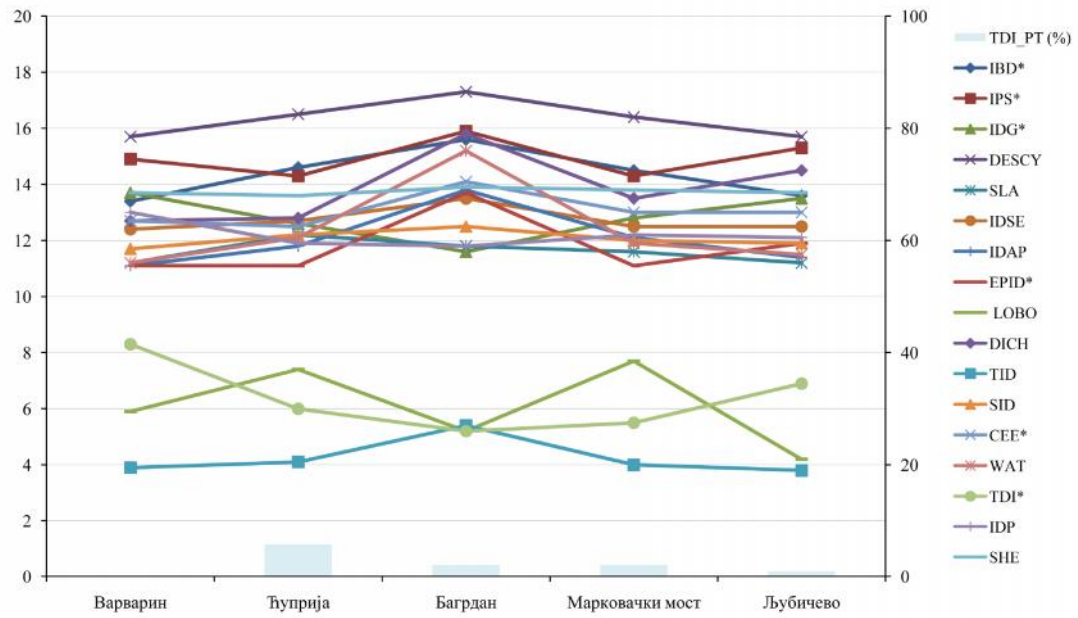
(17).

TID

TDI

(TDI_PT < 20 %,

)(17).



26.

2011.

() TDI_PT (%) ()

4.4.1. Статистичка анализа односа дијатомних индекса и одабраних физичких и хемијских параметара за реку Велику Мораву

Shapiro Wilk-

Spearman-

Spearman-

(18), (TID) , LOBO , pH As (LOBO, TID, SID TDI), IPS, EPID DICH EC, SID TH. TID Mg²⁺ Pb. **18.** (Spearman- ; p<0,05) (2010. 2011.)

/	T	TH	NO ₃	Mg ²⁺	pH	EC	As	Pb
IBD	-0,80	-0,07	0,05	-0,07	-0,51	-0,30	-0,53	0,18
IPS	-0,81	-0,13	0,00	-0,07	-0,55	-0,35	-0,58	0,17
IDG	-0,51	0,10	0,18	0,20	-0,46	-0,18	-0,42	0,04
DESCY	-0,41	0,08	-0,26	-0,06	-0,37	0,01	-0,34	0,10
SLA	-0,61	0,31	-0,28	0,27	-0,36	0,24	-0,36	0,27
IDSE	-0,68	0,22	-0,09	0,12	-0,42	0,03	-0,53	0,17
IDAP	-0,70	0,06	-0,07	0,02	-0,59	-0,14	-0,55	0,15
EPID	-0,77	-0,19	0,12	-0,19	-0,45	-0,40	-0,45	0,16
LOBO	0,42	-0,14	0,08	-0,17	0,04	0,03	0,31	0,13
DICH	-0,88	-0,12	0,05	-0,04	-0,58	-0,36	-0,54	0,27
TID	-0,16	0,34	-0,15	0,23	-0,03	0,27	-0,16	0,04
SID	-0,54	0,34	-0,02	0,16	-0,32	0,11	-0,34	0,14
CEE	-0,77	0,10	-0,03	0,10	-0,53	-0,14	-0,61	0,06
WAT	-0,53	0,18	-0,16	0,00	-0,37	0,03	-0,38	0,13
TDI	0,35	-0,17	0,32	-0,31	0,31	-0,22	0,24	-0,11
IDP	-0,69	0,12	-0,35	0,08	-0,50	-0,01	-0,44	0,26
SHE	-0,79	-0,04	0,15	-0,05	-0,60	-0,32	-0,53	0,24

*

4.5. Индикативни еколошки потенцијал реке Велике Мораве

(WFD, 2000)

-

(19).

CEE,

(, 74/2011).

VMOR_3

2010. 2011.

2010.

VMOR_3, IPS

CEE, 2011.

VMOR_3

2010,

VMOR_2

CEE IPS 2011.

19.

, 2010. 2011.

B		VMOR_3		VMOR_2	
		IPS	CEE	IPS	CEE
		III	I	I	I
		III	II	II	II
		IV	II	II	I
		IV	II	II	II
		IV	III	IV	III
		IV	II	IV	III
		III	I	II	I
		III	I	II	II
		II	I	II	I
		II	I	I	II
		II	I	I	I
		I	I	I	I

4.6. Бентосна заједница силикатних алги реке Саве

4.6.1. Флористички састав

184 55 (20).
Navicula (26), *Nitzschia* (17) *Gomphonema* (16).
27.
75 % , *A. pyrenaicum*, *A. pediculus*, *C. pediculus*, *C. placentula* var. *euglypta*, *D. vulgaris*, *N. antonii*, *N. cryptotenella*, *N. capitatoradiata*, *N. tripunctata*, *N. dissipata* *N. fonticola*.
50 75 % , : *A. minutissimum*, *E. silesiacum*, *F. vaucheriae*, *G. minutum* *N. reichardtiana*.
C. lancettula, *C. subminuscula*, *E. minima*, *G. sciotense*, *H. montana* *N. amphibia*, .
25 % , : *A. minutissimum*, *A. pyrenaicum*, *A. subatomus*, *A. pediculus*, *C. placentula* var. *euglypta*, *E. minima*, *L. mutica*, *M. cahabaensis*, *N. capitatoradiata*, *N. cryptotenella*, *N. recens*, *N. abbreviat* , *N. fonticola* *N. frustulum* var. *inconspicua*.

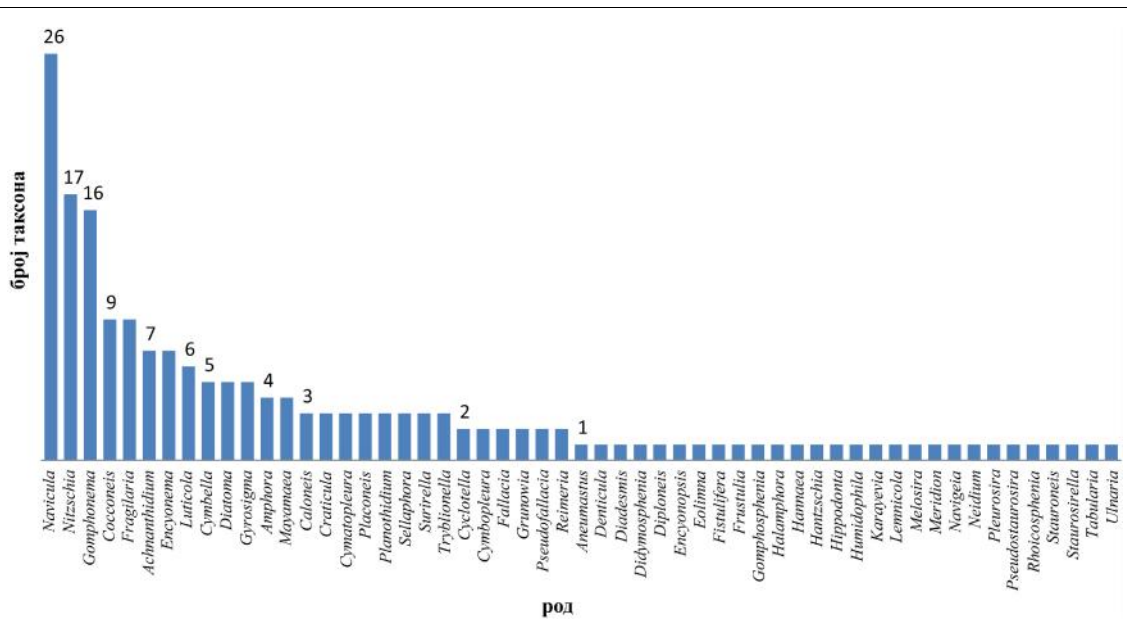
20.

2011., 2012., 2014.

2015. (, 7) (+ ; +* o ;)

Bacillariophyta																													
<i>Achnantheidium eutrophilum</i> (Lan.-Bert.) Lange-Bertalot	ADEU																												+
<i>A. exiguum</i> (Grun.) Czarnecki	ADEG																												+
<i>A. latecephalum</i> Kobayasi	ADLA																												+
<i>A. minutissimum</i> (Kütz.) Czarnecki	ADMI	+	+	+		+	+	+	+	+	+	+	+		+				+		+	+	+	+	+	+	+	+	
<i>A. pyrenaicum</i> (Hust.) Kobayasi	ADPY	+	+	+	+	+	+	+	+	+	+	+	+		+			+	+	+	+	+	+	+	+	+	+	+	
<i>A. saprophilum</i> (Kobay. & May.) Round & Bukhtiyarova	ADSA																											+	
<i>A. subatomus</i> (Hust.) Lange-Bertalot	ADSU				+		+	+		+	+	+																	
<i>Amphora copulata</i> (Kütz.) Schoeman & Archibald	ACOP				+		+	+	+	+	+	+	+		+			+	+		+	+					+	+	
<i>A. inariensis</i> Krammer	AINA	+		+	+	+	+	+	+	+	+	+							+	+	+	+	+	+	+	+	+	+	
<i>A. ovalis</i> (Kütz.) Kützing	AOVA						+	+		+	+									+	+		+	+					
<i>A. pediculus</i> (Kütz.) Grunow	APED	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	
<i>Aneumastus stroesei</i> (Østr.) Mann	ANSS	+	+																										
<i>Caloneis bacillum</i> (Grun.) Cleve	CBAC						+																					+	
<i>C. lancettula</i> (Sch.) Lange-Bertalot & Witkowski	CLCT	+		+	+		+			+		+	+		+			+	+	+			+	+	+	+	+		
<i>C. silicula</i> (Ehr.) Cleve	CSIL																											+	
<i>Cocconeis disculus</i> (Schum.) Cleve	CDIS		+																										
<i>C. neodiminuta</i> Krammer	CNDI			+		+				+																		+	
<i>C. pediculus</i> Ehrenberg	CPED	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	

<i>N. palea</i> (Kütz.) W. Smith	NPAL			+		+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	
<i>N. sigmoidea</i> (Nitzs.) W.Smith	NSIO																												+
<i>N. sociabilis</i> Hustedt	NSOC																												+
<i>N. subtilis</i> (Kütz.) Grunow	NISU																												+
<i>Placoneis gastrum</i> (Ehr.) Mereschkowsky	PGAS														+														
<i>P. paraelginensis</i> Lange-Bertalot	PPAE																												+
<i>P. symmetrica</i> (Hust.) Lange-Bertalot	PSYM															+	+		+	+	+	+	+						+
<i>Planothidium dubium</i> (Grun.) Round & Bukhtiyarova	PTDU	+		+																									
<i>P. frequentissimum</i> (Lan.-Bert.) Lange-Bertalot	PLFR	+			+		+				+								+	+	+					+	+	+	+
<i>P. lanceolatum</i> (Bréb. ex Kütz.) Bukhtiyarova	PTLA		+		+		+			+																			+
<i>Pleurosira laevis</i> (Ehr.) Compère	PLEV																												+
<i>Pseudofallacia monoculata</i> (Hust.) Liu, Kociolek & Wang	PMOC																												+
<i>P. tenera</i> (Hust.) Liu, Kociolek & Wang	PFTN																												+
<i>Pseudostaurosira parasitica</i> (Sm.) Morales	PPRS																												+
<i>Reimeria sinuata</i> (Greg.) Kociolek & Stoermer	RSIN	+	+	+	+	+	+			+	+	+							+	+						+	+	+	+
<i>R. uniseriata</i> Sala, Guerrero & Ferrario	RUNI		+	+	+	+	+			+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
<i>Rhoicosphenia abbreviata</i> (Agar.) Lange-Bertalot	RABB				+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
<i>Sellaphora bacillum</i> (Ehr.) D.G.Mann	SEBA																												+
<i>S. pupula</i> (Kütz.) Mereschkovsky	SPUP				+		+			+		+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
<i>S. seminulum</i> (Grun.) Mann	SSEM														+		+	+	+		+		+	+	+	+	+	+	+
<i>Stauroneis smithii</i> Grunow	SSMI																												+
<i>Staurosirella pinnata</i> (Ehr.) D.M.Williams & Round	SPIN															+													
<i>Surirella angusta</i> Kützing	SANG					+																							+
<i>S. brebissonii</i> var. <i>kuetzingii</i> Krammer & Lange-Bertalot	SBKU					+	+	+		+	+																		+
<i>S. minuta</i> Brébisson ex Kützing	SUMI									+		+																	
<i>Tabularia fasciculata</i> (Agar.) Williams & Round	TFAS																											+	+
<i>Tryblionella angustata</i> W. Smith	TANG									+																		+	+
<i>T. apiculata</i> W.Gregory	TAPI																												+
<i>T. levidensis</i> W. Smith	TLEV																												+
<i>Ulnaria ulna</i> (Nitz.) Compère	UULN	+				+		+		+	+	+	+	+	+														+

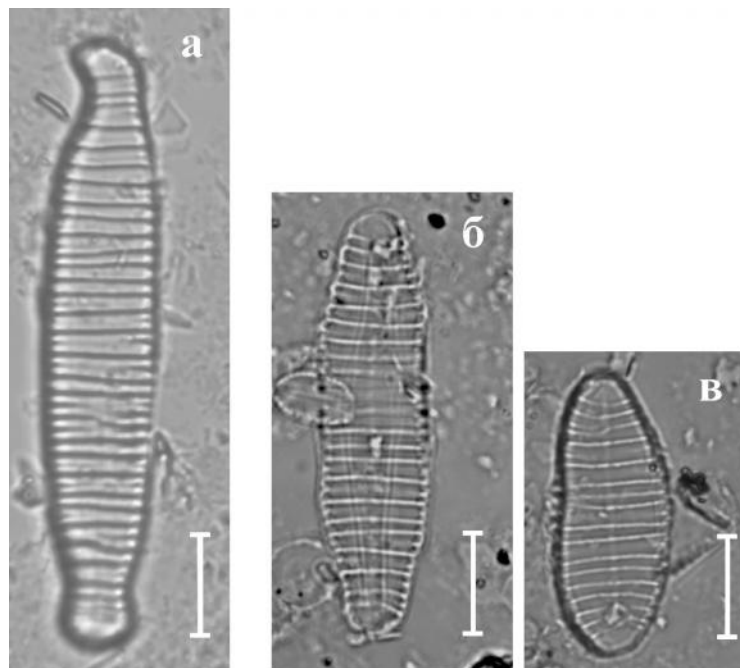


27.

,
Didymosphenia geminata *Diadesmis confervacea*. *D. geminata*,
 () (),
 (),
 0,2 % 1,1 %. *D. confervacea*
 () (), 0,3 % 0,6 %.
Mayamaea cahabaensis Morales & Manoylov (10),
 (20).
 (28) . *Diatoma ehrenbergii* *D. vulgare*, (1),
D. vulgare ()
 2). *D. ehrenbergii*
 () 2014. (0,99 %
), *D. vulgare*
 2012. (0,31 %).

0,95 %

() 2014.
(*D. ehrenbergii* 0,71 % *D. vulgaris* 0,24 %).



28. , 10 μm: *Diatoma ehrenbergii* () *D. vulgaris* (,)

4.6.2. Динамика у септембру 2011., 2012., 2014. и 2015. године

, : 100 (38)
) 2011, 110 (40)
 2012, 111 (39) 2014. 131 (37)
 2015.
 (24), (25), () (32) (35).
 () (47), () (46), () (56)
 () (65).
 2011. 23
 5 % (21).

: *A. pediculus*, *C. placentula* var. *euglypta*,
E. minima, *M. cahabaensis*, *N. cryptotenella*, *N. rostellata* *N. abbreviat* .

21.

(%)

2011.

						()	
						.	()
	42	24	34	41	37	47	47	38
	(%)*							
<i>Achnanthidium pyrenaicum</i>		11,7						
<i>Amphora pediculus</i>	26,9		12,6	5,9				
<i>Cocconeis pediculus</i>				5,6				
<i>C.placentula</i> var. <i>euglypta</i>		6,6	6,4	19,7			5,7	
<i>Cyclotella meneghiniana</i>	9,4					5,9		
<i>Eolimna minima</i>		19,9	10,4				13,6	20,9
<i>Gyrosigma sciotense</i>					6,0			
<i>Halamphora montana</i>							6,8	
<i>Mayamaea cahabaensis</i>	8,0	37,0	13,8	7,4				
<i>M. permitis</i>	7,6							
<i>Navicula cryptotenella</i>	5,6		12,3		30,9	6,2		
<i>N. recens</i>						5,9		13,5
<i>N. rostellata</i>				9,7	18,3	17,7	5,4	9,2
<i>N. tripunctata</i>			6,1					
<i>Nitzschia abbreviat</i>							17,3	
<i>N. capitellata</i>						7,8		
<i>N. dissipata</i>					9,3	9,3		
<i>N. filiformis</i> var. <i>conferta</i>								7,7
<i>N. fonticola</i>							5,7	
<i>N. palea</i>				6,5				
<i>Reimeria uniseriata</i>		6,0						
<i>Rhoicosphenia abbreviat</i>			6,1					
<i>Sellaphora seminulum</i>								16,3
(%)	57,6	81,3	67,8	54,7	64,6	52,8	54,5	67,5

*

5%

5 %

(81,3 %).

Mayamaea cahabaensis

(37 %).

() (52,8 %)

N. rostellata (17,7

%),

2011.,

1 %.

(21), *S. seminulum* (4,7 %) *C. placentula* var. *euglypta* (4,5 %) , *N. reichardtiana* (4,8 %) *N. antonii* (4,5 %).

2012.

21 5 % (22).

: *A. minutissimum*, *A. pediculus*, *C. placentula* var. *euglypta*, *E. minima*, *H. montana*, *M. cahabaensis*, *N. capitatoradiata*, *N. cryptotenella*, *N. tripunctata*, *N. abbreviat* , *N. dissipata*, *N. frustulum* var. *inconspicua* *N. palea*.

5 %

() (83,8 %).

N. frustulum var. *inconspicua* (29 %), *N. abbreviata* (20,9 %) *H. montana* (20,3 %),

() (50,3 %) *N. palea* (16,8 %).

5 % (22), *E. silesiacum* (4,6 % ; 3,1 % ; 4,4 %), *A. pediculus* (4,9 %), *N. fonticola* (4,0 % ; 4,9 %) *N. tripunctata* (4,7 %), *N. palea* *N. frustulum* var. *inconspicua* (4,9 % 4,6 %), *N. erifuga* *E. minima* (4,83 % 4,55 % ,) *M. permitis* (4,4 % ,).

22.

(%)

2012.

		((((((
(%)*														
<i>Achnanthidium minutissimum</i>		11,3	17,6		5,0	9,3								
<i>Amphora copulat</i>		7,7												
<i>A. pediculus</i>		20,8				9,9		25,0		9,1				
<i>Cocconeis placentula</i> var. <i>euglypta</i>	12,3	5,9				17,7	66,1	13,4	6,8					
<i>Eolimna minima</i>								16,6	23,2	21,7	13,2			14,4
<i>Gomphonema pumilum</i> var. <i>rigidum</i>						7,0								
<i>Halamphora montana</i>						13,7						20,3	5,4	11,7
<i>Mayamaea cahabaensis</i>								11,6	18,9					
<i>Navicula capitatoradiata</i>	12,0	5,1	10,8	25,8	12,1	6,7								
<i>N. cryptotenella</i>	9,8	5,9	17,9	12,3	6,5					7,7	29,9			
<i>N. germainii</i>											5,2			
<i>N. recens</i>												6,7	9,7	15,5
<i>N. reichardtiana</i>				5,7	8,0									
<i>N. rostellata</i>									13,0	10,0	15,2	7,0	12,5	
<i>N. tripunctata</i>	7,7	8,5	17,0	8,1	8,9									
<i>Nitzschia abbreviata</i>									11,1	10,6		20,9		5,9
<i>N. dissipata</i>					18,9									
<i>N. fonticola</i>	19,1	9,5			7,7									
<i>N. frustulum</i> var. <i>inconspicua</i>												29,0	6,0	31,9
<i>N. palea</i>				6,8									16,8	
<i>Reimeria uniseriata</i>								6,6						
(%)	60,9	74,6	62,8	58,7	67,2	57,3	73,0	73,1	73,1	59,1	63,5	83,8	50,3	79,5

*

5%

2014.

20

5 %

(23).

: *A. pyrenaicum*, *C. placentula* sensu lato,*E. minima*, *H. contenta*, *L. mutica*, *N. cryptotenella* *N. recens*.

5 %

() (70,9 %),

A. pyrenaicum

(30,0 %),

() (38,8 %) *L. mutica*
 (13,5 %) *C. placentula* sensu lato (11,3 %) () (38,9 %)
N. recens (13,5 %) *A. inariensis* (10,6 %).

23. (%)
 2014.

	36	33	32	45	42	42	43	45	43	44	44	56	36
	(%)*												
<i>Achnanthydium minutissimum</i>	8,4	13,0											
<i>A. pyrenaicum</i>	30,0	29,8	38,7		8,7	15,8							
<i>Amphora inariensis</i>						5,2						10,6	
<i>A. pediculus</i>				8,7	5,5	6,1							
<i>Cocconeis placentula</i> sensu lato	8,9	5,2	8,6	11,3	11,1	16,3	5,4	18,7	7,6	7,4			12,7
<i>Encyonema silesiacum</i>	5,9	5,8	7,0										
<i>E. ventricosum</i>	7,1	9,2	9,6										
<i>Eolimna minima</i>							25,2					8,0	11,6
<i>Gomphonema parvulum</i>													12,4
<i>Gyrosigma obtusatum</i>								5,7					
<i>Hantzschia amphioxys</i>									5,7				
<i>Humidophila contenta</i>							21,5			15,7			
<i>Luticola mutica</i>				13,5			7,2	7,6	40,8	25,9			
<i>L. ventricosa</i>									5,9				
<i>Navicula cryptotenella</i>					12,0								
<i>N. recens</i>											35,6	13,5	29,5
<i>N. reichardtiana</i>					5,5								
<i>N. tripunctata</i>				5,2	8,7			8,1					
<i>Nitzschia dissipata</i>	7,3	7,8								5,2	12,8	6,8	
<i>N. palea</i>											5,6		
(%)	67,7	70,9	63,9	38,8	51,6	43,4	59,4	40,1	60,0	54,2	54,0	38,9	66,2

* 5%

2014. 5 % 2 %
 , ()
 23). *A. minutissimum* (3,8 % , ; 3,0 %
 , ; 3,8 % ,), *D. vulgaris* (3,3 % ; 3,8 %
 , ; 3,2 % , ; 4,6 % ,), *N.*

cryptotenella (4,5 % , ; 3,8 % , 3,2 , ; 4,0 % , ; 3,4 % ,), *N. dissipata* (3,3 % , ; 4,7 % , ; 3,5 % , ; 3,8 %), *S. seminulum* (4,9 % , 2,7 5 ,).

24.

(%)

2015.

	35	37	37	36	65	37	41	36	41	51	48	50	41
	(%)*												
<i>Achnanthydium minutissimum</i>		43,7	9,0				7,0						
<i>A. pyrenaicum</i>	13,4	19,3				10,9	19,7						
<i>A. subatomus</i>			27,7	13,8		37,6	33,1						
<i>Amphora pediculus</i>				17,9	7,9					5,7		12,6	
<i>Cocconeis pediculus</i>						6,2						7,25	
<i>C. placentula</i> var. <i>euglypta</i>	9,0					7,9	5,0	7,0	5,6				
<i>C. placentula</i> var. <i>lineata</i>	6,2												
<i>C. pseudolineata</i>	25,9												
<i>Craticula subminuscula</i>								8,9			7,1		5,5
<i>Denticula tenuis</i>		6,1											
<i>Diatoma vulgaris</i>						5,3							
<i>Encyonema minutum</i>		5,4											
<i>Eolimna minima</i>					8,5			9,2	5,6		13,7		
<i>Gomphonema lagenula</i>													7,3
<i>Grunowia solgensis</i>					17,3								
<i>Halamphora montana</i>											11,1		6,6
<i>Mayamaea cahabaensis</i>								7,4	8,4				
<i>Navicula antonii</i>										7,0		12,1	
<i>N. capitatoradiata</i>					5,6								
<i>N. cryptotenella</i>				9,3	6,1	7,5		26,0		6,3			
<i>N. recens</i>													36,1
<i>N. rostellata</i>									8,6	7,9			
<i>N. tripunctata</i>			5,5	15,0									
<i>Nitzschia clausii</i>													8,6
<i>N. fonticola</i>			27,7	8,0			7,0						
<i>N. frustulum</i> var. <i>inconspicua</i>									8,4	12,0	26,6		
<i>Reimeria sinuat</i>	5,0												
<i>Sellaphora seminulum</i>								22,9	23,7	17,7	7,1	5,4	
(%)	59,5	74,4	69,9	66,1	45,4	75,4	71,8	81,4	60,4	56,6	65,6	37,4	63,9

*

5%

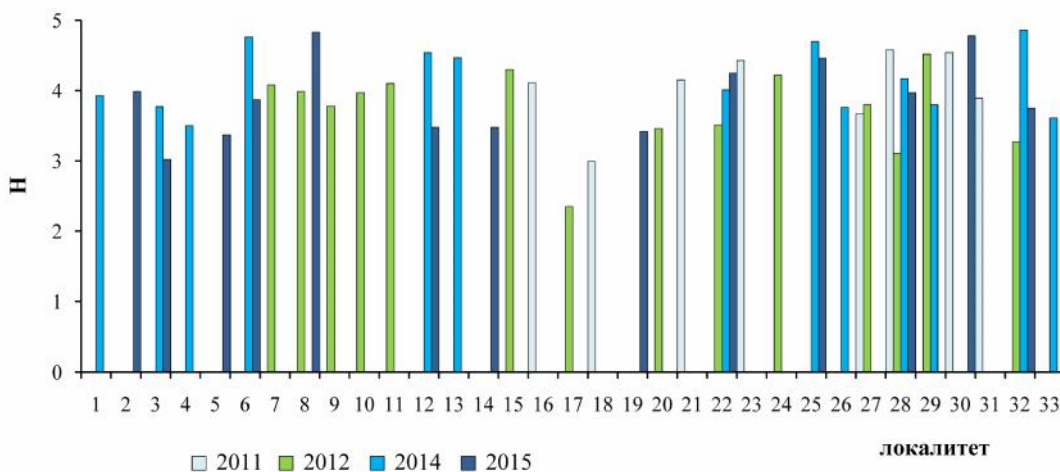
2015. 28
5 %
(24). : *A. minutissimum*, *A. pyrenaicum*, *A. subatomus*,
A. pediculus, *C. pseudolineata*, *G. solgensis*, *N. cryptotenella*, *N. recens*, *N. tripunctata*, *N. fonticola*, *N. frustulum* var. *inconspicua* *S. seminulum*.
5 %
() (71,8 %),
N. cryptotenella (26,0 %) *S. seminulum* (22,9 %),
() (37,4 %)
C. pediculus (12,6 %) *N. antonii* (12,1 %).
5 % (
24), *E. ventricosum* (3,4 %
; 4,8 % ,), *N. fonticola* (4,1 % , ; 4,2
% , ; 4,3 % ,) *N. palea* (4,4 % ,
)
R. abbreviata (4,5 % ,), *H. montana* *N. upsaliensis*
(4,8 % ,) *N. rostellata* (4,2 % ,).

4.6.3. Вредности Shannon-овог индекса диверзитета

Shannon-
(29) 2011. 2,99 (
18) 4,58 (. 28 ,). 2012.
2,35 (. 17) 4,52 (. 29
,). 2014.
3,5 (. 4 ,) 4,86 (. 32 ,
) 2015. 3,02 (. 3 ,)
4,83 (. 8).
Shannon-
2012. , .

Sh nnon-

(. 17 – 20), 2,35 3,46, .
 (. 1 16),
 3,02 (, ; . 3) 4,83 (, ; . 8),
 (. 21-33) 3,11 (,
) 4,86 (,).



29. Sh nnon- (H)

(1 33, 7)

4.6.4. Статистичка анализа бентосне заједнице силикатних алги реке Саве

(2011., 2012., 2014. 2015.),
 (CA) „FLORA” ,
 30. CA ,
 (48) (30) ,
 (184) (30). (CA1) 11,38 % ,
 9,24 % .
 (I) ,

(II)

(III)

2014.

, ()

().

/

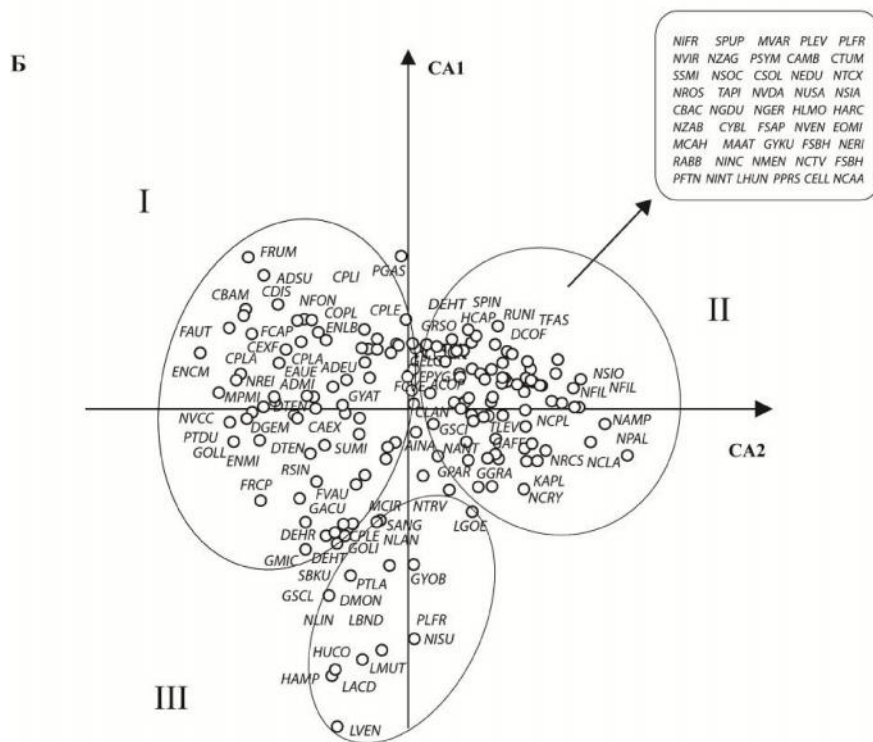
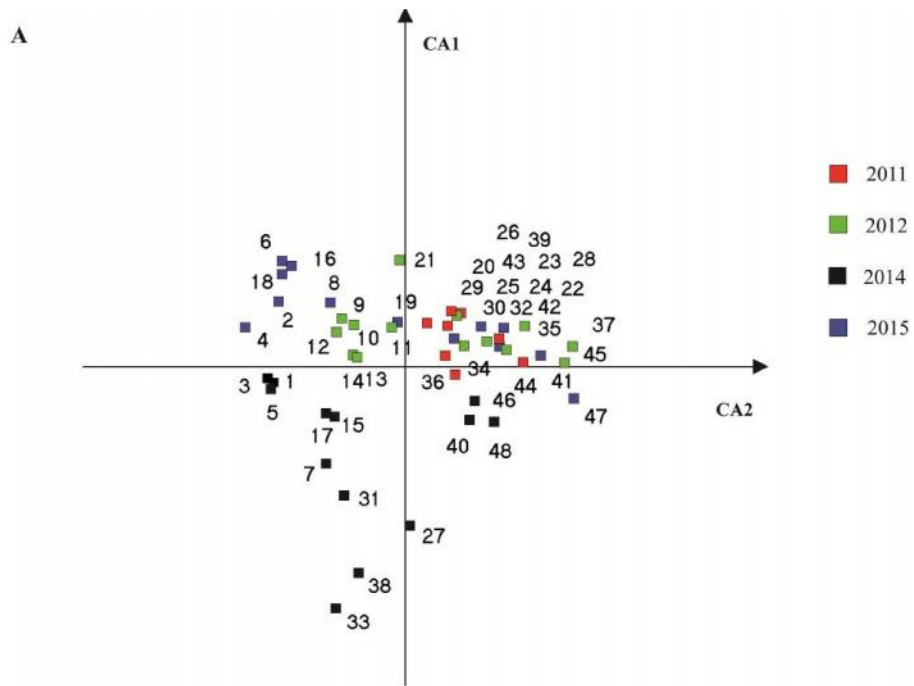
: *A. minutissimum*, *A. subatomus*,

Cocconeis (*C. placentula* var. *placentula*, *C. placentula* var. *klinoraphis*, *C. placentula* var. *lineata* *C. pseudolineata*), *Encyonema* (*E. auerswaldii*, *E. c espitosum* *E. langebertalotii*, *E. minutum*), *Fragilaria* (*F. acus*, *F. austriaca* *F. capucina* var. *capucina*), *D. tenuis*, *G. pumilum* var. *rigidum*, *G. tergestinum*.

, *G. olivaceolacuum*, *N. reinhardtii*, *N. splendidula*
N. viridulacalcis.

Mayamaea (*M. atomus*, *M. cahabaensis*, *M. permitis*), *Nitzschia*, *Sellaphora* (*S. bacillum*, *S. pupula* *S. seminulum*) *Navicula* (*N. recens*, *N. erifuga*, *N. germainii*, *N. rostellata*, *N. viridula* /).

Humidophila contenta *Luticola*: *L. acidoclinata*, *L. binodeformis*, *L. mutica* *L. ventricosa*.



30. CA (CA1 – 11,38 %, CA2 – 9,24 %);
(2011., 2012., 2014. 2015.);

4.7. Физичке и хемијске карактеристике воде Саве

2014. () 2015. () (25 26).
2014.

2014. 2015.
2014. 10,84
°C (), 2015. 9,9 °C
2014. 17,26 °C
(), 2015. 23,8 °C
() 23,7 °C ().

pH
2014. pH 7,55
() 8,71 ().
2015. pH 7,1
8,8 ().

2014.
, 381 μS/cm (,
) 277 μS/cm (,). 2015.

. 427 μS/cm
(), 100 μS/cm ().
2014.
, 1,75 meq/kg
() 2,76 meq/kg ().
2015. ,
) 2014, ()
) 3 meq/kg. 2,908 meq/kg

meq/kg (), 4,08
().

2014. ,
4,83 mg/l
(), 2,41 mg/l ().
, 130 %
() 74 % ().

2015. ,
. ()
(5,9 mg/l) (69 %),
() (9,2 mg/l) () (9,1)
() (69 %).

2014. ,
2,41mg/l C (,) 3,87 mg/l C (,).

2015.
, 1,41 mg/l C ()
3,08 mg/l C (,).

2014. -
() (-21,6 mV),
() (21,4 mV). 2015.

-
() (-74,7 mV),
() (90 mV).

2014.
1,69 mg/l () 3,86 mg/l
(). 2015. , ()
(), 4 mg/l.
1,44 mg/l ,
6,69 mg/l ().

. 2014.

2,53 mg/l (,) 13,4 mg/l (,),
2015. 8,6 mg/l () 26,38 mg/l (,).

2014. 2,27
µg/l (,) 12,53 µg/l (,), 2015.
5,24 µg/l () 203 µg/l (,).

2014. (20,9 mg/l)
(3,1 mg/l) (),
(63 mg/l) ()
(12,1 mg/l) ().

2015. ,
(42,29 mg/l), () (74,62
mg/l).
() (10,01 mg/l),
() (16,74).

2014. 0,7
mg/l () 6,01 mg/l (),
2015. 1,76 mg/l 18,03 mg/l
(). 2014. 0,3
mg/l () 1,55 mg/l
(), 2015. 0,28 mg/l
2,05 ().

2014. ,
0,8 mg/l () 3,79 mg/l (,
), 2015. 0,74 mg/l (,) 2,14 mg/l
(,). 2014.
2015.

2014.
0,22 µg/l (,
) 1,27 µg/l (,), 2015. 0,145
µg/l () 2,09 µg/l (,). 2014.
0,08 µg/l (,) 2,97 µg/l (,),

2015. 0,298 µg/l () 74,3 µg/l ()
 ,). 2014. 0,0005 µg/l
 (,) 0,38 µg/l (,) 2015.
 0,045 µg/l () 1,8 µg/l (,).
 2015. 1,51 µg/l (,) 7,11 µg/l ()
 ,), 2015. 0,758 µg/l
 () 7,87 µg/l (,).
 2014. 3,16 µg/l (,) 13,88 µg/l (,)
), 2015. 2,56 µg/l () 228 µg/l
 (,).

25.

2014.

(T)	°C	10,84	11,52	13,53	14,07	13,33	15,25	15,73	16,12	16,49	16,66	17,26	17,16
pH		7,7	8,44	7,81	7,85	7,73	7,66	7,55	7,95	7,61	8,71	7,57	8,03
(EC)	µS/cm	277	286	290	340	381	320	350	367	327	320	340	334
(Alk)	meq/kg	1,75	1,90	1,83	2,19	2,15	2,34	1,88	1,91	2,50	2,34	2,76	2,71
(DO)	mg/l	9,78	9,44	8,2	13	12,22	8,04	7,63	7,3	7,73	7,7	7,3	7,31
(O ₂ %)	%	93,3	91,2	82,6	130	117	81,6	74	75,2	79,8	80	77,5	76,2
(DOC)	mg/l C	3,15	2,49	2,41	2,91	2,74	3,64	2,98	3,87	2,98	4,83	2,95	2,59
(ORP)	mV	146	68,3	-21,6	-2	-5,1	17,1	4	6,7	21,4	3,7	19,4	14,3
(NO ₃ ⁻)	mg/l	2,11	2,33	3,27	3,77	1,69	1,82	3,22	2,55	3,52	3,60	2,26	3,86
(SO ₄ ²⁻)	mg/l	6,60	6,34	5,68	8,17	2,53	4,94	11,90	3,32	12,22	12,14	11,24	13,40
(Ca ²⁺)	mg/l	42,5	43,5	37,9	28,1	63,0	20,9	52,4	24,8	48,1	52,4	52,7	52,9
(Mg ²⁺)	mg/l	9,4	10,0	12,1	6,0	11,2	3,1	8,9	4,3	8,2	9,1	9,5	9,6
(Na ⁺)	mg/l	0,8	1,11	2,33	0,70	2,25	0,00	2,82	0,46	2,21	2,53	6,01	5,84
(K ⁺)	mg/l	0,27	0,44	1,28	0,41	1,09	0,30	1,45	0,52	1,48	1,55	0,72	0,56
(Si)	mg/l	0,8	1,02	2,84	1,18	2,20	1,17	3,63	1,75	3,43	3,78	3,79	3,71
(Cl)	mg/l	2,85	3,14	3,60	3,97	1,55	1,45	6,57	1,56	6,22	6,20	3,99	6,21
(Br)	mg/l	0,000	0,001	0,000	0,009	0,000	0,015	0,006	0,000	0,006	0,013	0,000	0,010
(F)	mg/l	0,05	0,04	0,03	0,04	0,03	0,04	0,04	0,05	0,04	0,04	0,07	0,05
(Ag)	µg/l	0,02	0,00	0,00	0,03	0,01	0,02	0,0005	0,13	0,08	0,04	0,0005	0,04
(As)	µg/l	0,22	0,22	0,26	0,30	0,35	0,53	0,59	0,70	1,02	1,17	1,27	1,10
(Ba)	µg/l	0,01	0,01	0,02	0,01	0,01	0,01	0,02	0,02	0,02	0,02	0,02	0,02
(Cd)	µg/l	0,0005	0,0005	0,0005	0,0005	0,0005	0,0005	0,0005	0,0005	0,0005	0,0005	0,0005	0,0092
(Co)	µg/l	0,0025	0,01	0,03	0,01	0,01	0,02	0,05	0,02	0,03	0,03	0,02	0,03
(Cr)	µg/l	0,06	0,07	0,10	0,07	0,07	0,31	0,23	0,26	0,28	0,27	0,18	0,30
(Cu)	µg/l	0,39	0,51	0,67	0,72	0,65	1,14	1,26	2,08	2,53	1,43	1,50	3,71
(Fe)	µg/l	3,24	3,16	4,44	4,45	3,40	13,88	10,04	8,44	10,91	10,92	4,79	8,08
(Mn)	µg/l	0,24	0,23	0,08	0,20	0,31	0,50	2,88	0,12	0,79	2,97	1,35	2,49
(Mo)	µg/l	0,83	0,71	0,23	0,50	0,41	0,29	0,30	0,29	0,36	0,30	0,39	0,34
(Ni)	µg/l	0,025	0,025	0,025	0,025	0,025	1,135	1,881	2,082	2,095	2,122	1,584	1,734
(P)	µg/l	2,27	3,13	6,05	9,46	3,90	5,04	12,53	4,56	6,00	3,80	7,35	4,51
(Pb)	µg/l	0,0005	0,08	0,01	0,05	0,08	0,23	0,11	0,05	0,09	0,38	0,11	0,23
(Rb)	µg/l	0,56	0,57	1,00	0,91	0,88	0,60	0,64	0,67	0,67	0,62	0,65	0,58
(Se)	µg/l	0,06	0,04	0,04	0,02	0,05	0,05	0,06	0,05	0,06	0,06	0,08	0,04
(Sr)	µg/l	69,65	72,53	55,37	91,35	80,90	101,11	94,97	110,62	121,60	121,81	116,37	115,12
(V)	µg/l	0,13	0,14	0,22	0,33	0,43	0,61	0,70	0,69	0,73	0,76	0,79	0,86
(Zn)	µg/l	1,51	3,54	3,58	3,41	3,64	3,26	2,86	2,92	7,11	3,17	2,47	1,92

”

”

,

26.

2015.

(T)	°C	9,9	14,5	17,6	17,7	20,2	23,1	22,6	23,8	24,8	24,1	22,6	22,4	23,7
pH		7,1	8,2	8,8	7,4	8,4	7,9	7,9	7,4	7,8	8,3	8,48	8,08	8,44
(EC)	µS/cm	210	230	260	100	280	320	285	270	336	427	293	336	324
(Alk)	meq/kg	2,921	2,908	3,790	3,882	3,843	3,924	4,077	4,080	3,969	3,716	3,229	3,389	3,635
(DO)	mg/l	8,8	10,5	9,2	8,4	8,4	8,9	9,1	5,9	8,2	6,85	7,69	8,18	7,28
(O2%)	%	80,7	102,7	101	94,6	96,6	105,8	107,4	69	100	82,2	89,2	94,2	85,9
(DOC)	mg/l C	1,41	1,66	1,93	1,93	2,12	2,14	2,32	2,84	2,78	3,08	2,29	2,58	2,40
(ORP)	mV	44	16	-13	90	-1	3	20	85	-16,6	-74,7	-27,3	4,1	-44,8
(NO ₃)	mg/l	1,44	2,92	6,48	6,69	6,53	4,57	4,23	5,39	4,32	4,56	2,11	2,21	2,42
(SO ₄ ²⁻)	mg/l	8,60	8,94	12,43	12,01	19,46	17,51	17,88	16,56	24,66	26,38	15,96	18,71	17,82
(Ca ²⁺)	mg/l	42,29	46,80	62,16	62,37	61,43	61,77	64,11	63,30	66,27	74,62	60,63	62,48	58,07
(Mg ²⁺)	mg/l	14,00	10,71	14,58	15,11	15,61	15,93	16,74	16,55	15,95	15,97	10,01	11,66	9,99
(Na ⁺)	mg/l	1,76	2,29	6,24	6,15	8,23	7,60	8,41	8,74	7,64	18,03	7,76	9,58	7,54
(K ⁺)	mg/l	0,28	0,50	1,29	1,22	1,54	1,45	1,81	1,95	1,66	2,05	1,18	1,27	1,25
(Si)	mg/l	0,92	0,74	0,98	0,91	1,12	1,16	1,21	1,91	2,01	2,04	2,01	2,11	2,14
(Cl)	mg/l	2,82	3,56	9,07	9,05	9,84	9,24	10,16	10,21	9,19	47,71	18,66	24,36	18,29
(Br)	mg/l	*	0,03	0,07	0,08	0,08	0,09	0,09	0,09	.	0,08	.	.	.
(F)	mg/l	0,07	0,06	0,06	0,05	0,06	0,06	0,06	0,08	0,08	0,07	0,05	0,06	0,06
(Ag)	µg/l	0,00	0,00	0,044	0,049	0,021	0,00	0,00	1,18	0,03	0,137	0,00	7,73	0,00
(As)	µg/l	0,145	0,285	0,378	0,388	0,743	0,615	0,692	1,42	1,36	1,62	1,46	1,66	2,09
(Ba)	µg/l	0,0094	0,00968	0,0179	0,0182	0,0233	0,0189	0,0213	0,0248	0,0222	0,027	0,0201	0,0223	0,0199
(Cd)	µg/l	0,000	0,000	0,005	0,005	0,034	0,006	0,007	0,03	0,018	0,016	0,01	0,005	0,015
(Co)	µg/l	0,017	0,086	0,107	0,093	0,303	0,116	0,111	0,196	0,313	0,218	0,136	0,105	0,16
(Cr)	µg/l	0,115	0,382	0,47	0,584	0,558	0,319	0,293	0,386	0,417	0,517	0,416	0,486	0,506
(Cu)	µg/l	0,79	2,44	1,12	1,76	1,81	1,47	2,38	1,69	1,57	1,36	1,03	1	2,26
(Fe)	µg/l	2,56	26,5	23,4	19,2	228	28,5	30,7	172	182	114	58,1	43,2	86,5
(Mn)	µg/l	0,298	3,48	16,8	5,18	50,5	13,4	11,9	50,9	74,3	49,9	23,5	19,7	27,8
(Mo)	µg/l	0,559	1,24	0,612	0,667	1,44	1,09	1,13	0,833	0,762	0,774	0,469	0,608	0,901
(Ni)	µg/l	1,05	0,279	2,38	2,01	5,03	0,497	0,61	5,61	19,2	22,5	1,07	1,23	33,1
(P)	µg/l	5,24	45,4	112	106,3	116	54,9	60,7	203	115	125	47,4	54,0	92,6
(Pb)	µg/l	0,045	0,207	0,106	0,328	1,8	0,19	0,197	0,703	0,556	0,385	0,464	0,307	0,516
(Rb)	µg/l	1,59	3,32	5,85	5,73	7,6	6,7	7,8	8,69	6,5	8,68	5,5	6,27	5,88
(Se)	µg/l	0,095	0,088	0,149	0,085	0,093	0,096	0,089	0,146	0,093	0,129	0,101	0,113	0,124
(Sr)	µg/l	139	90	113	112	128	121	127	123	154	159	121	134	118
(V)	µg/l	0,191	0,243	0,338	0,342	0,719	0,554	0,579	1,11	1,26	1,22	0,756	0,84	1,17
(Zn)	µg/l	0,758	5,85	7,15	2,52	8,2	1,63	1,63	7,87	4,89	2,9	3,04	1,63	4,19

*

”

”

,

4.8. Статистичка анализа утицаја физичких и хемијских параметара на састав заједнице бентосних силикатних алги реке Саве током септембра 2014. и 2015. године

2014. ()
 2015. ().
 Mantel- ()
 (r=0,188; p=0,092)

(FS) (p 0,05)
 11 (27),
 – (, NO₃⁻ , Si, Mg²⁺ Alk)
 – (As, Fe, F , Pb, Rb Mo).

27. (p 0,05)

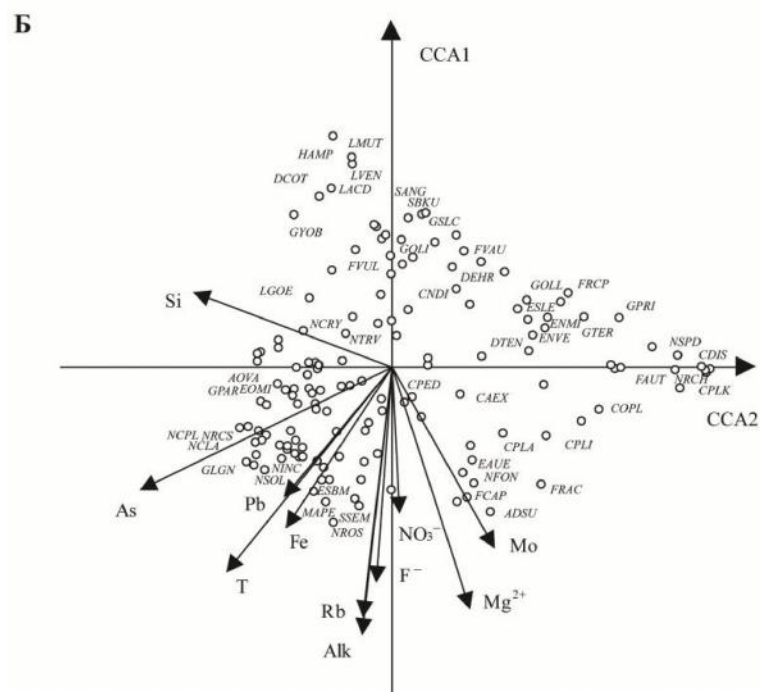
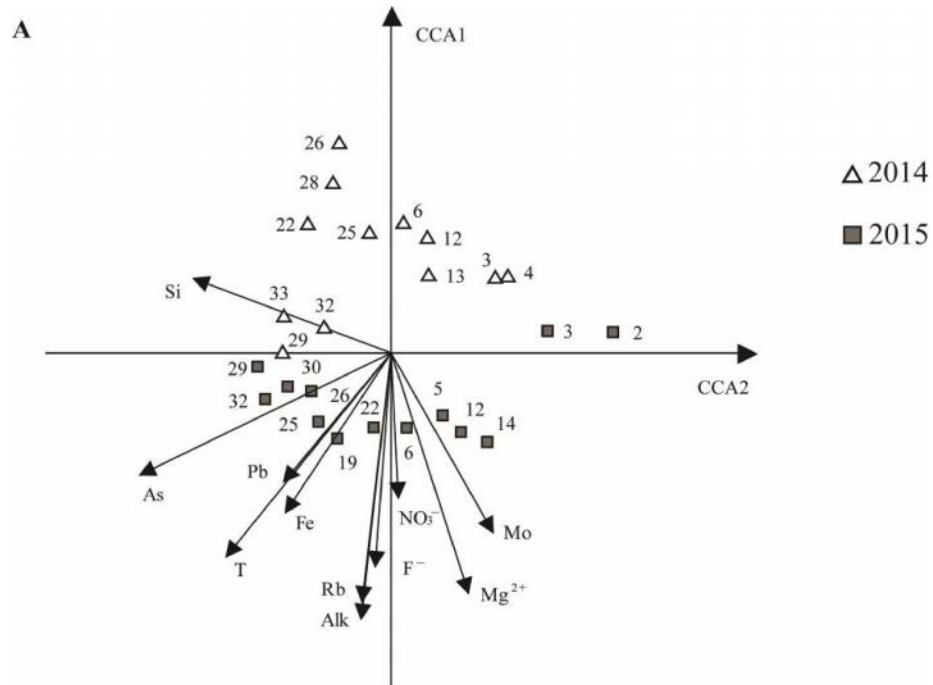
		F	
(T)	0,458	2,903	0,000
(NO ₃ ⁻)	0,231	1,380	0,012
(Si)	0,243	1,457	0,022
(Mg ²⁺)	0,219	1,305	0,044
(Alk)	0,418	2,624	0,050
(As)	0,553	3,596	0,000
(Fe)	0,268	1,616	0,000
(F)	0,229	1,368	0,028
(Pb)	0,231	1,380	0,034
(Rb)	0,444	2,803	0,040
(Mo)	0,252	1,516	0,044

(CCA)

11 (31).
 CCA (24,2 %), ,
 ().
 ()

CCA As
 Si, CCA (21,1%

) 2014. (;
) 2015. (;).
 CCA , , ,
 .
 ()
 CCA , ().
Achnanthydium sp., *Cocconeis* sp., *Encyonema* sp.,
Fragilaria sp. *Gomphonema* sp.,
Cymatopleura, *Eolimna*, *Mayamaea*, *Nitzschia* *Navicula* ,
Humidophila contenta *Luticola* 2014.
 CCA , , *N.*
recens *E. minima*. , *N. capitellata* *N.*
clausii, , *N.*
solgensis . , , Pb, Fe
 T , *N. frustulum* var.
inconspicua, *E. subminuscula* *M. permitis* . ()
) .
 As, Pb Fe.
A. ovalis, *G. parvulum*, *L. goeppertiana*, *N. cryptocephala*, *N. trivialis* *E.*
minima Si, *C. pediculus*, *C. placentula* var.
lineata, *C. placentula* var. *placentula*, *C. pseudolineata*, *C. excisa* *E. auerswaldii*
 , Si.
 , *S. seminulum* *N. rostellata*,
 CCA , Rb F.
 Mg²⁺, Mo NO₃ , *N. fonticola*, *A. subatomus*, *F.*
capucina var. *capucina* *F. acus* ,
N. fonticola ()
) *A. subatomus* () .
 Mg²⁺, Mo NO₃
 2014., : *L. acidoclinata*, *L. ventricosa*,
L. mutica, *H. contenta*, *H. amphioxys*, *F. vulgaris* *G. obtusatum*.



31. CCA

(
; 24,2% (CCA1) 21,1% (CCA2)
, Monte Carlo
);
(4);
(18)

4.9. Квалитет

на основу дијатомних индекса

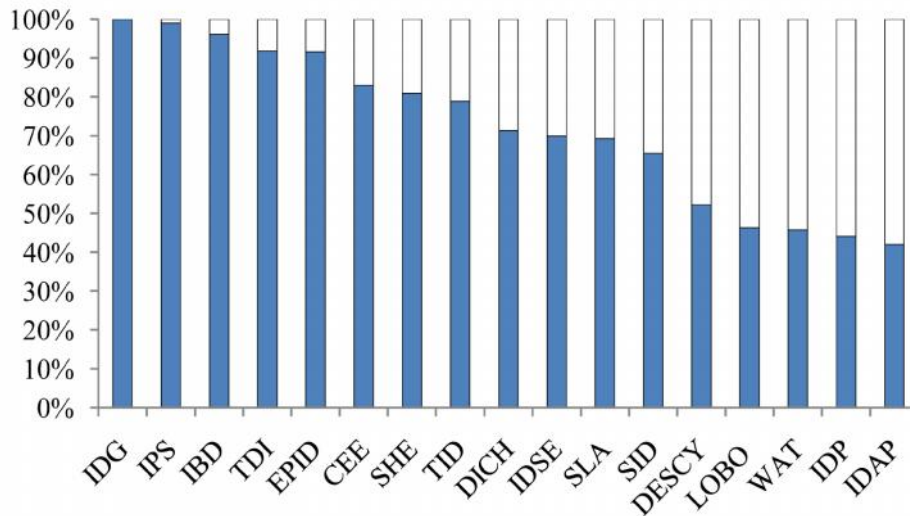
2011., 2012., 2014. 2015. ,

(32).

80%

(33 36,
80%

).



32.

(%)

2011.

(

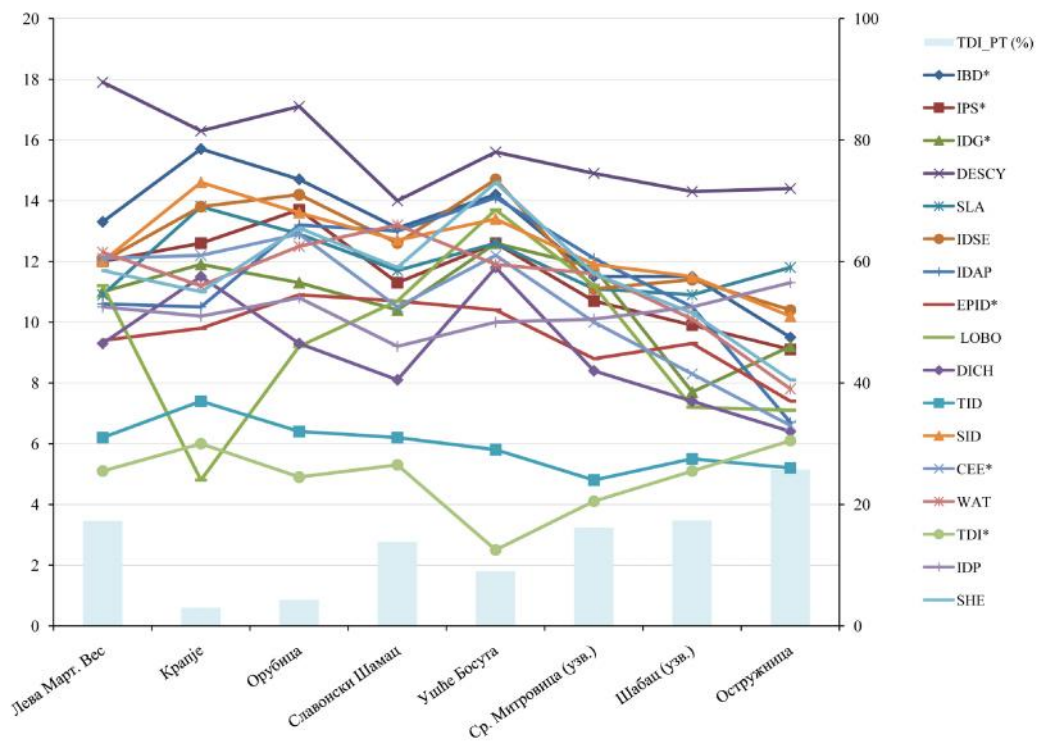
),

(33).

(IPS, IDG, SLA, IDSE, EPID, CEE, WAT IDP) (28).

DESCY () IBD (

), DICH TID. TID
 (,
 , ,)
 (,)
 (TDI_PT < 20 %),
 (TDI_PT 25,8 %) (28).



33.

2011.

() TDI_PT (%) ()

2012.

(34).

(,)

()

(). DESCY

, DICH, TID IDP (28).

TDI

(), ,

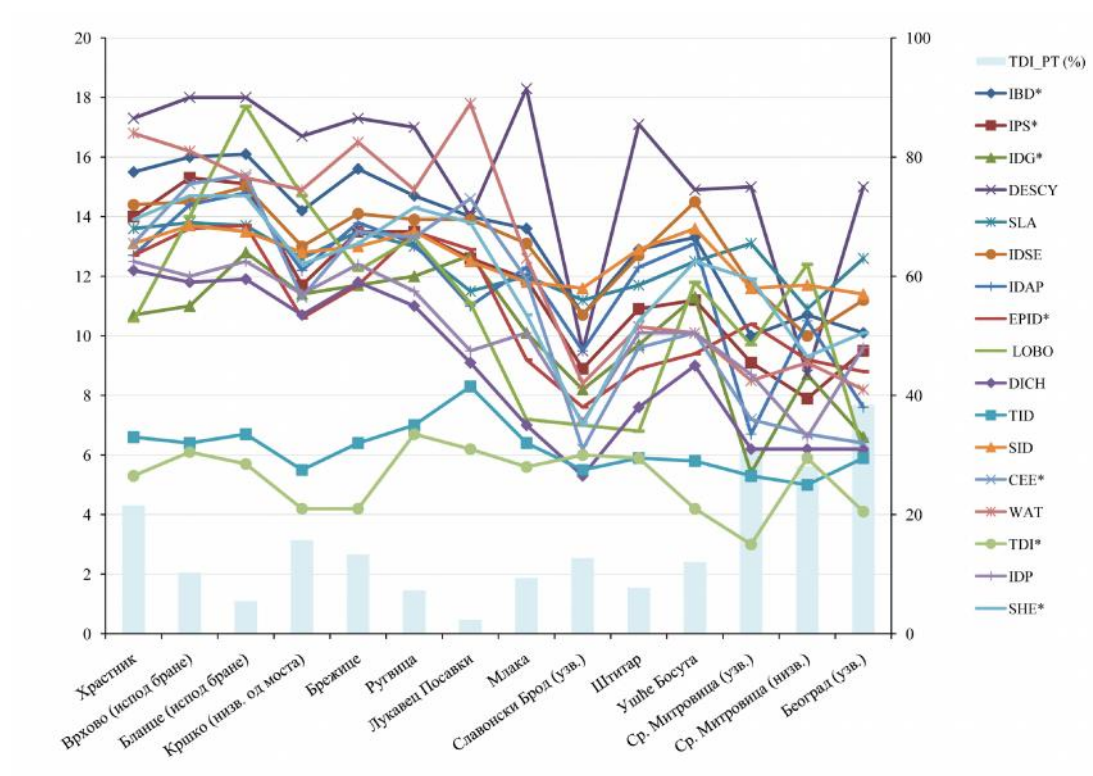
() (),

() ()

(), TDI_PT

(21,5 %, 30,7 %, 28,1 % 38,4 %,).

(IBD, IPS, IDG, SLA, IDAP, EPID, SID SHE) (LOBO) (28).



34.

2012.

() TDI_PT (%) ()

2014. , ,

(35). ()

, (), 2012.

, .

() ().

DESCY SID

, TID LOBO (28). TDI

() ()

(),

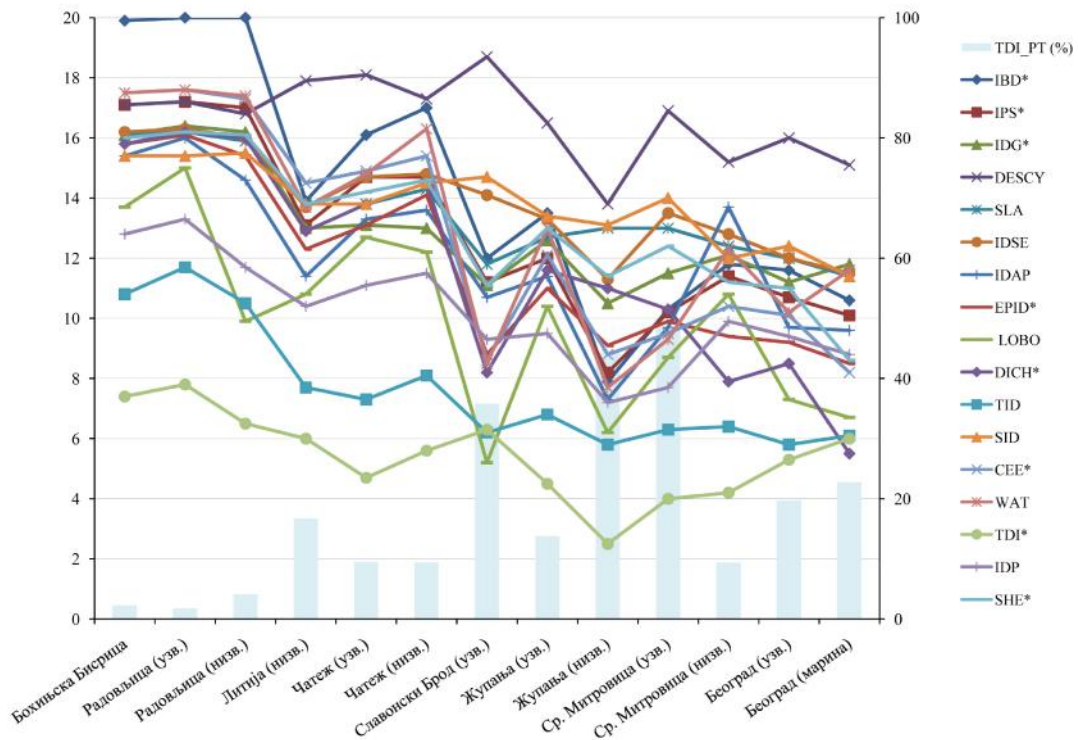
() (), TDI_PT

(35,8 %, 44,4 % 48,6 %,).

() ,

() (IBD, IPS, IDG,

SLA, IDAP, EPID, IDP SHE) (DICH) (28).



35.

2014.

() TDI_PT (%) ()

2015.

(36).

()

()

()

()

DESCY

, TID IDP

(28).

TDI

()

().

()

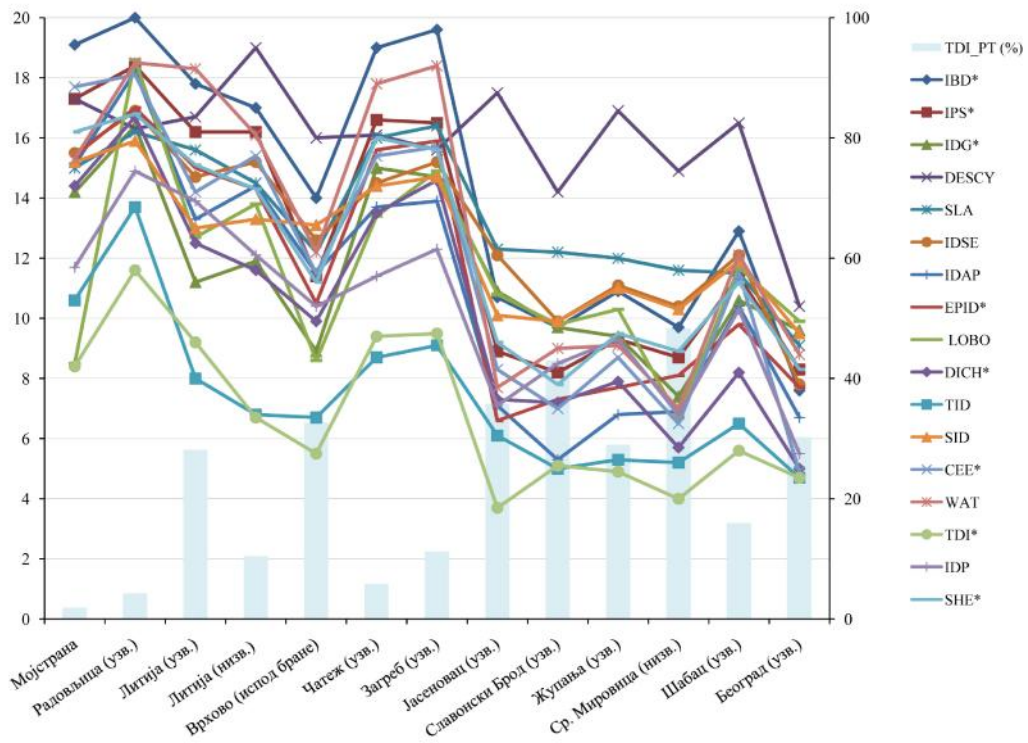
(),

(),

()

() (),
 TDI_PT (28,1%, 32,7 %, 35,8 %, 43,0 %, 29,0 %, 48,3 % 30,2 %,).

() ()
 () (IBD, IDG, SLA, IDSE, SID)
 (IPS, IDAP, EPID, DICH, CEE, WAT, SHE) (28).



36. 2015.

() TDI_PT (%) ()

4.9.1. Статистичка анализа односа дијатомних индекса и одабраних физичких и хемијских параметра

Shapiro Wilk-

- (p<0,05),

2014.

2015.

(

)(29).

29.

- (p<0,05)

2014. (

)

2015.

(

)

	IBD	IPS	IDG	DESCY	SLA	IDSE	IDAP	EPID	LOBO
p	0,877	0,991	0,141	0,282	0,861	0,301	0,460	0,919	0,215
	DI-CH	RTI	RSI	CEE	WAT	TDI	IDP	SHE	
p	0,343	0,797	0,076	0,617	0,985	0,121	0,896	0,539	

Pearson-

a

, As, Fe Si

2014.

(

),

Mo

(

30).

2015.

(),

, As, Fe Si

30.

* (Pearson- ; p<0,05)

2014. ()

2015.

()

2014/ 2014	Alk	As	F ⁻	Fe	Mg ²⁺	Mo	NO ₃ ⁻	Pb	Rb	Si	T
IBD	-0,51	-0,73	-0,26	-0,68	0,41	0,81	-0,35	-0,40	0,09	-0,58	-0,91
IPS	-0,53	-0,74	-0,30	-0,69	0,42	0,81	-0,29	-0,41	0,14	-0,58	-0,91
IDG	-0,58	-0,66	-0,22	-0,65	0,45	0,87	-0,21	-0,40	-0,08	-0,52	-0,90
DESCY	-0,18	-0,59	-0,41	-0,13	0,02	0,20	-0,23	-0,27	0,41	-0,43	-0,50
SLA	-0,72	-0,76	-0,27	-0,70	0,33	0,82	-0,35	-0,63	0,11	-0,65	-0,94
IDSE	-0,54	-0,78	-0,40	-0,45	0,20	0,75	-0,38	-0,39	0,09	-0,68	-0,91
IDAP	-0,42	-0,55	-0,34	-0,48	0,40	0,72	-0,19	-0,08	0,03	-0,42	-0,77
EPID	-0,66	-0,80	-0,32	-0,76	0,42	0,80	-0,32	-0,60	0,22	-0,61	-0,95
LOBO	-0,52	-0,54	-0,33	-0,60	0,50	0,58	0,03	-0,38	0,33	-0,26	-0,68
DICH	-0,78	-0,85	-0,33	-0,70	0,27	0,68	-0,36	-0,74	0,31	-0,66	-0,90
TID	-0,63	-0,71	-0,21	-0,64	0,38	0,88	-0,36	-0,47	-0,08	-0,61	-0,94
SID	-0,62	-0,81	-0,34	-0,32	-0,04	0,62	-0,59	-0,56	0,07	-0,78	-0,85
CEE	-0,66	-0,79	-0,32	-0,79	0,48	0,73	-0,27	-0,57	0,30	-0,55	-0,92
WAT	-0,50	-0,64	-0,33	-0,75	0,60	0,72	-0,14	-0,34	0,19	-0,38	-0,82
TDI	-0,14	-0,44	-0,05	-0,41	0,37	0,60	-0,38	-0,14	-0,12	-0,37	-0,64
IDP	-0,46	-0,65	-0,22	-0,68	0,44	0,75	-0,33	-0,29	0,14	-0,51	-0,85
SHE	-0,74	-0,82	-0,34	-0,64	0,31	0,71	-0,35	-0,65	0,26	-0,62	-0,92
2015/ 2015	Alk	As	F ⁻	Fe	Mg ²⁺	Mo	NO ₃ ⁻	Pb	Rb	Si	T
IBD	-0,18	-0,91	-0,28	-0,61	0,24	0,28	0,14	-0,39	-0,41	-0,90	-0,66
IPS	-0,25	-0,90	-0,34	-0,69	0,13	0,22	0,10	-0,46	-0,50	-0,91	-0,72
IDG	-0,20	-0,68	0,00	-0,58	0,15	0,35	-0,10	-0,52	-0,42	-0,70	-0,52
DESCY	0,00	-0,62	-0,06	-0,22	0,46	-0,13	0,41	-0,13	-0,05	-0,52	-0,43
SLA	-0,05	-0,89	-0,15	-0,55	0,36	0,22	0,23	-0,45	-0,30	-0,84	-0,53
IDSE	-0,22	-0,92	-0,23	-0,55	0,25	0,22	0,20	-0,35	-0,39	-0,89	-0,70
IDAP	-0,35	-0,88	-0,40	-0,62	0,01	0,32	0,07	-0,31	-0,54	-0,92	-0,76
EPID	-0,25	-0,88	-0,40	-0,70	0,08	0,25	0,07	-0,45	-0,51	-0,90	-0,68
LOBO	0,01	-0,46	-0,13	-0,45	0,06	0,41	0,19	-0,41	-0,09	-0,54	-0,22
DICH	-0,24	-0,90	-0,17	-0,55	0,21	0,36	0,09	-0,37	-0,45	-0,90	-0,68
TID	-0,50	-0,78	-0,13	-0,53	-0,07	0,35	-0,17	-0,37	-0,60	-0,79	-0,74
SID	-0,33	-0,88	-0,29	-0,56	0,13	0,38	0,02	-0,29	-0,49	-0,89	-0,71
CEE	-0,27	-0,91	-0,27	-0,61	0,19	0,24	0,09	-0,38	-0,50	-0,90	-0,74
WAT	-0,08	-0,83	-0,36	-0,63	0,18	0,31	0,21	-0,41	-0,36	-0,87	-0,56
TDI	-0,27	-0,79	-0,26	-0,63	0,03	0,34	0,00	-0,48	-0,48	-0,82	-0,59
IDP	-0,21	-0,86	-0,28	-0,52	0,19	0,25	0,25	-0,34	-0,37	-0,86	-0,64
SHE	-0,26	-0,88	-0,35	-0,70	0,13	0,23	0,07	-0,46	-0,48	-0,89	-0,70

*

4.10. Индикативни еколошки потенцијал реке Саве на делу тока у Србији

IPS CEE

74/2011).

SA_2. 2014.

SA_1

2011. 2014.

2012. 2015.

SA_1

CEE

31.

2011., 2012., 2014. 2015.

		SA_2		SA_1	
		IPS	CEE	IPS	CEE
2011		II	II	III	IV
2012		III	III	III	IV
2014		II	II	II	II
2015		II	III	III	V

5. Дискусија

(Andreji , 2012; Krizmani , 2013; Vidakovi , 2013; Vasiljevi , 2014; Jakovljevi , 2016 , 2016), (Jakovljevi , 2014) (Predojevi , 2017).

(Descy Gosselain, 1994; Köhler, 1993; Wu , 2010; Grabowska Mazur-Marzec, 2016; a o , 2006 ; a o , 2006 ; Ržani anin , 2005; Obuškovi Kalafati , 1979; , 2010), (Vannote , 1980)

(Werner Köhler, 2005).

(Obuškovi , 1979; Obuškovi Kalafati 1979, Obuškovi Martinovi , 1987; Martinovi - Vitanovi , 1994, 1996; Lauševi , 1998; a o , 2006), (Simi , 2015; Vasiljevi , 2017).

Simi (2015), (81,7 %), (11,11 %) (6,54 %).

(Makovinska Hlubikova, 2015; Simi , 2010). (65 %), (35 %) (, 2010).

162

184

177 (Andreji ., 2012),

188 (De Jonge ., 2008),

145 (Szabó ., 2005).

(WFD, 2000),

(JDS1, 2 3) (Makovinska Hlubikova, 2015; Liška ., 2015).

JDS2 , 68

(., 2010).

(391) 2.345 km,

JDS2 .

17, 11 ,

75 % : *A. pediculus*, *C. placentula* var. *euglypta*, *D. vulgaris*, *N. cryptotenella*, *N. tripunctata* *N. dissipata*.

: *C. subminuscula*, *G. parvulum*, *G. pumilum* var. *rigidum*, *N. lanceolata*, *N. abbreviata*, *N. amphibia*, *N. frustulum* var. *inconspicua*, *R. sinuata*, *R. uniseriata*, *R. abbreviata* *U. ulna*, : *A. pyrenaicum*, *C. pediculus*, *N. antonii*, *N. capitatoradiata* *N. fonticola*.

(

75 %): *A. pediculus*, *C. placentula* var. *euglypt* , *N. capitatoradiata*, *N. cryptotenella* *N. fonticola*, : *A. pediculus*, *C. placentula* var. *euglypt* , *C. subminuscula*, *N. lanceolata*, *N. abbreviata*, *N. dissipata* *N. frustulum* var. *inconspicua*.

(Besse-Lototskaya ., 2011;

Hofmann ., 2011).

Amphora, *Cocconeis*, *Eolimna*, *Gyrosigma*, *Luticola*, *Navicula*, *Nitzschia*, *Rhoicosphenia* *Reimeria*

(Makovinska Hlubikova, 2015).

() (), -
 (6 7),
 .
 ,
 (, 2010.)
 , 40 (,) 57
 (,). (2010, ,
 2011.) ,
 24 51 (,),
 24 39.
 , , 32 45 (,
) 31 43 (,).
 (DAPC) ()
 12). :
 , .
 , ,
 .
Amphora pediculus ,
 , ()
 53,32 %).
 , 2010, *A. pediculus*
N. lanceolata, *G. olivaceum*, *N. dissipata*,
F. saprophila (24,2 %) (9). , ,
 2010. , *A.*
pediculus, *N. abbreviata*, *N. frustulum* var. *inconspicua* (10),
 (11), 2010. *N. dissipata*.
 , *C. atomus*
 (22,97 % 40,43 %) *C. meneghiniana* (38,42 % 21,29 %)
N. frustulum var. *inconspicua*.
 (12), 2010. 2011. ,

A. pediculus, *N. dissipata* *G. olivaceum*. 2011.
D. problematica (36,73 %
 30,25 %), *R. abbreviat* *N. lanceolata*.
C. atomus *C. meneghiniana*
 . Makovinska Hlubikova
 (2015) ,
C. meneghiniana, 92 % 85 %
 .
 ,
 (Ács ., 2003 Makovinska Hlubikova, 2015).
 ,
 2011., 2012., 2014. 2015. 33 ,
 ,
 ,
 2011, 2012. 2015. (, 2012, 2013,
 2016; Meteorološki i hidrološki bilten br. 9, 2011, 2012, 2015; Pregled hidroloških razmer
 površinskih voda v Sloveniji, 2011, 2012, 2015). ,
 ,
 .
 ,
 (21 24). ,
 24 () 35 (),
 , 47 (,) 56 (,
), () , 65
 .
 ,
 . CA (30

)
,
(937 rkm,
507 m . .) (657 rkm, 98 m . .) (622 rkm, 95 m . .)
) (2 rkm, 69 m . .). T
(360 rkm, 82 m . .) (118 rkm, 72 m . .),
2014. ().

,
(),
,
() (30).
” (. *ecological guild*) . Passy (2006)

,
” (. *low profile*) „ ”
,

” (. *high profile*) „ ”
,

,
” (. *motile*)
,

- (Passy, 2001; Potapova Charles, 2002;
Soinien ., 2004).
(7, 30),

(*Achnantheidium*, *Cocconeis*, *Encyonema*, *Fragilaria*, *Gomphonema*).

Mayamaea, *Nitzschia*, *Sellaphora* *Navicula*.

Borojevi (2017)

CA

(2014.

). *H. contenta* *Luticola*: *L. acidoclinata*, *L. binodeformis*, *L. mutica* *L. ventricosa* *H. contenta* (21,5 %)

(23) *E. minima* (25,2 %). *L. mutica* 40,84%

(Hofmann ., 2011).

2014.

CA

TDI

TDI_PT (Kelly Whitton, 1995; Kelly ., 2001).

Passy (2006),

(Fore Grafe, 2002).

Kelly (2003).
 TDI_PT (%) CA .

(7). - (8)

,
 TDI_PT (%),

(35,8 % 66,5 % ,
 17 19).

- , - - ,
 (Van Dam ., 1994).

(13 15),
 2010., 2011. .

(, 2011, 2012).

(

, ,
) (

,). Passy

(2006) ,

(13, 14

15), (Kolarevi ., 2012).

F. saprophila (24,16 %),
 9 % *F. saprophila* - -
 (Van Dam ., 1994).

M. atomus (5 %) *M. permitis*

(8,5 %),

F. saprophila
(*M. permitis*) (Lange-Bertalot, 2001).

(Krebs, 2001).

Sh nnon-

) (11),

Sh nnon-

2011.

, Sh nnon-

2012.) (29),

Sh nnon-

2015.

2014.

Sh nnon-

H. contenta

Luticola,

(Peterson, 1996).

(Grimm Fisher, 1989).

(13).
 (13 15).
 CCA ,
C. placentula var. *lineata*, *N. capitellata*, *E. minima*, *A. granulata*, *L. comta*, *C. atomus* *C. meneghiniana* pH,
D. problematica, *E. leibleinii*, *E. silesiacum*, *G. olivaceum*,
N. reichardtiana *N. tripunctata*.
 , Potapova Charles (2002)
 :
 pH .
 (Patrick, 1971; Patrick .,
 1969; Lowe, 1974). *Cocconeis*
 (Vinson Rushforth, 1989;
 Patrick, 1971), *D. mesodon* *E. prostratum* (*E. leibleinii*)
 (Potapova Patrick, 2002).
 (DeNicola, 1996).
 . Anderson (2000)
 ,
 pH
 pH,
 (pH 7) (Kovács
 ., 2006).
 (8),

5 µg/l,

II

(TNMN Yearbook, 2004).

CCA

NO₃ Pb,

(13).

(Vrzel ., 2016).

Kolarevi

(2012)

CCA , Pb

NO₃

(Leland, 1995;

Carpenter Waite, 2000).

().

(13 15)

(Potapova Charles, 2003).

CCA a

(2014.)

(2015.),

CCA (31).

(25

26).

CCA

(2014.)

(2015.)

(31). CCA ,

N. recens *E. minima* ,

(25 26).

12 km (Popovic ., 2001),

(7).

CCA ,

(30) ().

(Vidmar ., 2016; Vrzel ., 2016). *N. solgensis*

E. minima As, Pb Fe

() (26).

N. recens, *E. minima*

As Morin (2012),

. *N. recens*

(Várbíró ., 2012). Makovinska Hlubikova (2015)

(Besse-Lototskaya ., 2011).

N. recens *E. minima*

2014.

(Vidmar ., 2016),

(25).

(Vidmar et al., 2016; Vrzel et al., 2016).

CCA, (*L. goeppertiana*, *N. cryptocephala*, *N. trivialis*, *E. minima*), *Cocconeis*, (Fore-Grafe, 2002).

(Bondoc et al., 2016).

(De Jonge et al., 2008; Fore-Grafe, 2002; Gold et al., 2003; Ivorra et al., 1999; Morin et al., 2008, Sabater, 2000). De Jonge (2008), (Round, 1991), (Falasco et al., 2009).

(9-28). (: *F. recapitellata*, *F. vaucheriae*, *D. moniliformis*, *D. vulgaris*, *U. ulna*. (1), *D. vulgaris* (2) (Falasco et al., 2009). (), (). *D. ehrenbergii*, *D. vulgaris*, (1), *D. vulgaris* (2). Morin (2012), 10 ‰ (1 ‰)

(1,98 %
).
 (U.
ulna D. vulgaris), *D. vulgaris* (1,78 %
).
 2010.
 (0,4 µg/l)
 0,07 µg/l,
 (EU, 2013).
 (Fernández ., 2017),
 10 ‰ (Morin ., 2012),
 ,
 ,
Didymosphenia geminata *Diademsia confervacea*.
 ,
 . *D. geminata*
 (Krammer
 Lange-Bertalot, 1986).
 (Whitton
 ., 2009).
 (Blanco Ector, 2009; Kawecka Sanecki, 2003; Kilroy ., 2009; Kumar
 ., 2008).
 ,
 (Kilroy ., 2008) (Bhatt
 ., 2008). - (Uroševi , 1994)
 (Obuškovi Maslikovi , 1997; a o ., 2007), (Pujin
 ., 1999; Martinovi -Vitanovi Kalafati , 2002; Subakov-Simi Cvijan, 2004),
 (a o ., 2006 , 2008; Marinkovi ., 2016)
 (Krizmani , 2015).

D. confervacea,
 (Coste Ector, 2000),
 (Lai .., 2010) (Krizmani ..,
 2015; Simi .., 2016; Predojevi .., 2017).

D. geminata
 (),
 (), 0,2 % 1,1 %. *D.*
confervacea
 (), 0,3 % 0,6 %.

Mayamaea cahabaensis (
 10). (),
 5,53 % () 9,54 %
 (), ()
 7,38 % () 37,05 % (). , *M. cahabaensis*
 ” ”

(Morales Manoylov, 2009). *Eolimna comperei*
 Ector, Coste & Iserentant, (Coste Ector,
 2000; Blanco .., 2010; Novais, 2011). Falasco Bona (2013),

M.
cahabaensis *E.comperei*. *M. cahabaensis*
 1 % 40 % (Morales Manoylov, 2009).
 50%

(Falasco Bona, 2013),
. Morales Manoylov (2009)
N. amphibia, *S. seminulum*, *A. minutissimum*, *A. pediculus*, *C. placentula* var. *lineat* *R. abbreviat* . *M. cahabaensis* (37 %),
. *M. cahabaensis* (Morales Manoylov, 2009)
(Falasco Bona, 2013), (6 7).
((EN 13946, 2003),
(EN 14407, 2004).
(Smucker Vis, 2010).
(Kahlert ., 2016; Brabcová ., 2017; Werner ., 2016; Poikane ., 2016).
(Poikane ., 2016).

OMNIDIA.

80%

: IPS, IDG, IBD, TDI, EPIDI, CEE SHE (14 32).

IPS IDG 93 % 100 % .

IPS (Kelly, 2013;

Eloranta Andersson 1998; Kwadrans ., 1998; Goma ., 2005; Ács ., 2003, 2004; Trábert ., 2017; Makovinska Hlubikova, 2015),

(. 74/2011). IDG

IPS,

Kelly (1995)

IDG ,

IPS .

(Bennett ., 2014).

(Rimet Bouchez, 2012).

(2010.

2011.),

(16 19, 17).

17).

TDI ,

(17). TDI_PT,

TDI,

20 %

DESCY, IDP, IPS WAT,

LOBO, TID, DICH TDI.

LOBO (Lobo et al., 2002)

pH (LOBO, TID, SID TDI),

(33 36).

CA (30). DESY, TID, IDP

TDI (TDI).

TDI_PT, 20 %,

(Lecointe et al., 1993).

(Besse-Lototskaya et al., 2011).

(WFD, 2000),

(Commission Decision, 74/2011).

: IPS CEE,

VMOR_3

2

IPS CEE

IPS

(Annex 31).

VMOR_2,

1

IPS CEE

IPS

2010.

2011,

(Annex 31).

VMOR_3

VMOR_2.

VMOR_3.

1

SA_1 (

,) SA_2 (,).

1 ,

IPS CEE ,

, 74/2011,

CEE .

CEE

,

2

2

CCA ,

pH,

,

,

,

,

.

,

.

,

.

/

,

,

.

,

(

., 2010).

6. Закључци

162

•

•

, *Amphora pediculus*, *Cocconeis placentula* var. *euglypt*, *Navicula capitatoradiata*, *N. cryptotenella*, *Nitzschia fonticola*, *Amphora pediculus*, *Cocconeis placentula* var. *euglypta*, *Craticula subminuscula*, *Navicula lanceolata*, *Nitzschia abbreviata*, *N. dissipata*, *N. frustulum* var. *inconspicua*.

•

(DAPC),

(),

•

pH,

•

•

Sh nnon-

2010.),

(,

Sh nnon-

(,

2011.).

• , C , (),
(,).
, - ()
, ,
(,).

• , , , .

• *Navicula recens*, *Eolimna minima* *Nitzschia solgensis*

, .
• , Sh non-

. ,
, .
2014.

Sh non-

, .

• *Mayamaea cahabaensis*,

(,) , .

, ,
, .

• *Didymosphenia geminata* *Diadlesmis confervacea*, .

• T

2 %

()

,

.

,

.

•

17

,

,

IPS, IDG, IBD, TDI,

EPIDI, CEE SHE,

(80 %),

IPS IDG

93 % 100 %

.

,

IPS

,

,

.

•

As Fe,

.

,

.

•

,

.

•

IPS

- CEE,

.

•

/

,

,

.

,

CEE

2

2

.

• , .
• , .
• , .
• , .
• , .
• , .

7. Литература

-
- Ács, É., Szabó, K., Kiss, K. T., Hindák, F. (2003). Benthic algal investigations in the Danube river and some of its main tributaries from Germany to Hungary. *Biologia Bratislava*, 58 (4): 545–554.
- Ács, É., Szabó, K., Tóth, B., Kiss, K. T. (2004). Investigation of benthic algal communities, especially diatoms of some Hungarian streams in connection with reference conditions of the Water Framework Directives. *Acta Botanica Hungarica*, 46 (3 4): 255–278.
- Allan, J. D., Castillo, M. M. (2007). *Stream ecology: structure and function of running waters*. Springer Netherlands, 436 pp.
- Anderson, N. J. (2000). Miniview: diatoms, temperature and climatic change. *European Journal of Phycology*, 35 (4): 307–314.
- Andreji, J. Z. (2012). Floristi ko-ekološka analiza silikatnih algi (Bacillariophyta) reke Nišave i pritoka Jerme i Temske. Doktorska disertacija. Univerzitet u Beogradu, Biološki fakultet, 279 str.
- Andrén, C., Jarlman, A. (2008). Benthic diatoms as indicators of acidity in streams. *Fundamental and Applied Limnology/Archiv für Hydrobiologie*, 173 (3): 237–253.
- AQEM (2002). Manual for the application of the AQEM system. A comprehensive method to assess European streams using benthic macroinvertebrates, developed for the purpose of the Water Framework Directive. Version 1.0, Contract No: EVK1-CT1999-00027.
- Barber, H.G, Haworth, E. Y. (1981). *A Guide to the Morphology of the Diatom Frustule*. Freshwater Biological Association. Scientific Publication No. 44, Cumbria, 112 pp.
- Barnese, L. E., Lowe, R. L. (1992). Effects of substrate, light, and benthic invertebrates on algal drift in small streams. *Journal of North American Benthological Society*, 11 (1): 49–59.
- Bellinger, B. J., Cocquyt, C., O'Reilly, C. M. (2006). Benthic diatoms as indicators of eutrophication in tropical streams. *Hydrobiologia*, 573 (1): 75–87.
- Bellinger, E. G., Sigeo, D.C. (2010). *Freshwater algae: identification and use as bioindicators*. John Wiley and Sons, Ltd., United Kingdom, 271 pp.
- Bennett, J. R., Sisson, D. R., Smol, J. P., Cumming, B. F., Possingham, H. P., Buckley, Y. M. (2014). Optimizing taxonomic resolution and sampling effort to design cost-effective
-

-
- ecological models for environmental assessment. *Journal of applied ecology*, 51 (6): 1722–1732.
- Bennion, H., Juggins, S., Anderson, N. J. (1996). Predicting epilimnetic phosphorus concentrations using an improved diatom-based transfer function and its application to lake eutrophication management. *Environmental Science and Technology*, 30 (6): 2004–2007.
- Besse-Lototskaya, A., Verdonshot, P.F., Coste, M., Van de Vijver, B. (2011). Evaluation of European diatom trophic indices. *Ecological Indicators*, 11 (2): 456–467.
- Bhatt, J. P., Bhaskar, A., Pandit M. K. (2008). Biology, distribution and ecology of *Didymosphenia geminata* (Lyngbye) Schmidt an abundant diatom from the Indian Himalayan rivers. *Aquatic Ecology*, 47 (3): 347–353.
- Blanco, C. Cejudo-Figueiras, I., Álvarez-Blanco, E., Bécáres, L., Hoffmann, L., Ector, L. (2010). Atlas de las diatomeas de la cuenca del Duero. Área de Publicaciones. Universidad de León, 386 pp.
- Blanco, S., Ector, L. (2009). Distribution, ecology and nuisance effects of the freshwater invasive diatom *Didymosphenia geminata* (Lyngbye) M. Schmidt: a literature review. *Nova Hedwigia*, 88 (3 4): 347 422.
- Blažen i , J. (2000). *Sistematika algi*. IV izdanje. NNK, Beograd, 298 str.
- Bondoc, K.G.V., Heuschele, J., Gillard, J., Vyverman, W., Pohnert, G. (2016). Selective silicate-directed motility in diatoms. *Nature Communications*, 7.
- Borojevi , K. K., Udovi , M. G., Žutini , P., Várбірó, G., Plenkovi -Moraj, A. (2017). Do benthic diatom assemblages reflect abiotic typology: a case study of Croatian streams and rivers. *Acta Botanica Croatica*, 76 (1): 80 90.
- Brabcová, B., Marvan, P., Opat ilová, L., Brabec, K., Fránková, M., Heteša, J. (2017). Diatoms in water quality assessment: to count or not to count them? *Hydrobiologia*, 795 (1): 113 127.
- Burkholder, J. M., Wetzel, R. G. (1990). Epiphytic alkaline phosphatase on natural and artificial plants in an oligotrophic lake: Re-evaluation of the role of macrophytes as a phosphorus source for epiphytes. *Limnology and Oceanography*, 35 (3): 736 747.
-

-
- Burkholder, J.M. (1996). Interactions of benthic algae with their substrata. In: R.J. Stevenson, M.L. Bothwell, R.L. Lowe (eds.). *Algal Ecology: Freshwater Benthic Ecosystems*. Academic, San Diego, pp. 253–297.
- Čađović, S., Miletić, A., Čurković, A. (2007). Phytoplankton, physicochemical and saprobiological characteristics of the Danube river, on the stretch through Serbia. BALWOIS Conference, Ohrid, Republic of Macedonia.
- Čađović, S., Miletić, A., Čurković, A. (2008). The composition and biomass of phytoplankton of the Sava River. BALWOIS Conference, Ohrid, Republic of Macedonia, pp. 1–11.
- Čađović, S., Miletić, A., Bugarski, R. (2006 a). Phytoplankton biomass of the Danube river (Serbia). *Jugoslovensko društvo za zaštitu voda*, pp. 67–72.
- Čađović, S., Miletić, A., Đopuša-Glišić, T., Đeniš, L. (2006 b). Physical-chemical characteristics and phytoplankton composition of the Sava River on its lower flow stretch through Serbia. In 36th Conference of the International Association for Danube Research-IAD, Proceedings, Vienna-Klosterneuburg, pp. 184–188.
- Carpenter, K. D., Waite, I. R. (2000). Relations of habitat-specific algal assemblages to land use and water chemistry in the Willamette Basin, Oregon. *Environmental Monitoring and Assessment*, 64 (1): 247–257.
- Cemagref (1982). *Etude des méthodes biologiques quantitative d'appréciation de la qualité des eaux*. Rapport Division Qualité des Eaux Lyon – Agence financière de Bassin Rhône–Méditerranée–Corse, Pierre-Bénite, 218 pp.
- Coste, M., Boutry, S., Tison-Rosebery, J., Delmas, F. (2009). Improvements of the Biological Diatom Index (BDI): Description and efficiency of the new version (BDI-2006). *Ecological indicators*, 9 (4): 621–650.
- Coste, M., Ector, L. (2000). Diatomées invasives exotiques ou rares en France: principales observations effectuées au cours des dernières décennies. *Systematics and Geography of Plants*, 70: 373–400.
- Cox, E. J. (2011). Morphology, cell wall, cytology, ultrastructure and morphogenetic studies. In: Seckbach, J., Kociolek, P. (eds.). *Cellular origin, life in extreme habitats and astrobiology Vol. 19: The Diatom World*. Springer Netherlands, pp. 21–45.
-

-
- Cumming, B. F., Smol, J. P. (1993). Development of diatom-based salinity models for paleoclimatic research from lakes in British Columbia (Canada). Twelfth international diatom symposium. Springer, Dordrecht, pp. 179–196.
- De Jonge, M., Van de Vijver, B., Blust, R., Bervoets, L. (2008). Responses of aquatic organisms to metal pollution in a lowland river in Flanders: a comparison of diatoms and macroinvertebrates. *Science of the Total Environment*, 407(1): 615–629.
- Dell'Uomo, A. (2004). *L'Indice Diatomico de Eutrofizzazione/Polluzione (EPI-D) nel Monitoraggio delle Acque Correnti*. Linee Guida. APAT: Roma, 101 pp.
- DeNicola, D. M. (1996). Periphyton responses to temperature at different ecological levels. In: Stevenson, R. J., Bothwell, M. L., Lowe, R. L. (eds.). *Algal Ecology: Freshwater Benthic Ecosystems*. Academic, San Diego, pp. 150–176.
- Descy, J. P., Gosselain, V. (1994). Development and ecological importance of phytoplankton in a large lowland river (River Meuse, Belgium). *Hydrobiologia*, 289: 139–155.
- Descy, J.P. (1979). A new approach to water quality estimation using diatoms. *Nova Hedwigia*, 64: 305–323.
- Descy, J.P., M. Coste (1991). A test methods for assessing water quality based on diatoms. *Verhandlung Internationale Vereinigung de Limnologie*, 24: 2112–2116.
- Dudley, T. L. (1992). Beneficial effects of herbivores on stream macroalgae via epiphyte removal. *Oikos*, 65 (1): 121–127.
- ikanovi, V., Tomovi, J., Zori, K., Vrankovi, J., Vasiljevi, B., Zindovi, B., Todorovi, A. (2010). Biological data and WFD implementation in Serbia. Conference Proceedings. Balkans regional young water professionals conference, Belgrade, pp. 44-47.
- Eldund, M. B., Stoermer, E. F. (1997). Ecological, evolutionary, and systematic significance of diatom life histories. *Jornal of Phycology*, 33 (6): 897–918.
- Eloranta, J.P., Andersson, K. (1998). Diatom indices in water quality monitoring of some South Finnish rivers. *Verhandlungen des Internationalen Verein Limnologie*, 26:1213–1215.

-
- EN 13946 (2003). Water quality – Guidance standard for the routine sampling and pretreatment of benthic diatoms from rivers. Geneva: Comité European de Normalisation.
- EN 14407 (2004). Water quality – Guidance standard for the identification, enumeration and interpretation of benthic diatom samples from running waters. Geneva: Comité European de Normalisation.
- EU (2013). Directive 2013/39/EU of the European Parliament and of the Council of 12 August 2013 amending Directives 2000/60/EC and 2008/105/EC as regards priority substances in the field of water policy.
- Falasco, E., Bona, F. (2013). Recent findings regarding non-native or poorly known diatom taxa in north-western Italian rivers. *Journal of Limnology*, 72 (1).
- Falasco, E., Bona, F., Badino, G., Hoffmann, L., Ector, L. (2009). Diatom teratological forms and environmental alterations: a review. *Hydrobiologia*, 623 (1): 1–35.
- Falkowski, P.G., Raven, J.A. (1997). *Aquatic Photosynthesis*. Malden, MA, Blackwell Science, 375 pp.
- Fernández, M. R., Martín, G., Corzo, J., de la Linde, A., García, E., López, M., Sousa, M. (2017). Design and testing of a new diatom-based index for heavy metal pollution. *Archives of Environmental Contamination and Toxicology*, <https://doi.org/10.1007/s00244-017-0409-6>.
- Fore, L. S., Grafe, C. (2002). Using diatoms to assess the biological condition of large rivers in Idaho (USA). *Freshwater Biology*, 47 (10): 2015–2037.
- Gaiser, E. E., Bachmann, R. W. (1993). The ecology and taxonomy of epizoic diatoms on Cladocera. *Limnology and Oceanography*, 38 (3): 628–637.
- Gaul, U., Geissler, U., Henderson, M., Mahoney, R., Reimer, C. W. (1993). Bibliography on the fine-structure of diatom frustules (Bacillariophyceae). *Proceedings of the Academy of Natural Sciences of Philadelphia*, 144: 69–238.
- Geitler, L. (1932). Der Formwechsel der pennaten Diatomeen. *Archiv für Protistenkunde*, 78: 1–226.
- Gold, C., Feurtet-Mazel, A., Coste, M., Boudou, A. (2003). Impacts of Cd and Zn on the development of periphytic diatom communities in artificial streams located along a

-
- river pollution gradient. *Archives of Environmental Contamination and Toxicology*, 44: 189–197.
- Gomà, J, Ortiz, R., Cambra, J., Ector, L. (2004). Water quality evaluation in Catalanian Mediterranean rivers using epilithic diatoms as bioindicators. *Vie Milieu*, 54 (2–3): 81–90.
- Gómez, N., Licursi, M. (2001). Pampean Diatom Index (IDP) for assessment of rivers and streams in Argentina. *Aquatic Ecology*, 35 (2): 173–181.
- Grabowska, M., Mazur-Marzec, H. (2016). The influence of hydrological conditions on phytoplankton community structure and cyanopeptide concentration in dammed lowland river. *Environmental monitoring and assessment*, 188 (8): 488.
- Greenacre, M. J. (1984). *Theory and applications of correspondence analysis*. Academic Press, London, 364 pp.
- Grimm, N. B., Fisher, S. G. (1989). Stability of periphyton and macroinvertebrates to disturbance by flash floods in a desert stream. *Journal of the North American Benthological Society*, 8 (4): 293–307.
- Gross, E. M. (2003). Allelopathy of aquatic autotrophs. *Critical Reviews in Plant Sciences*, 22 (3–4): 313–339.
- Guiry, M. (2012). How many species of algae are there? *Journal of Phycology*, 48: 1057–1063.
- Guiry, M.D., Guiry, G.M. (2017). *AlgaeBase*. World-wide electronic publication, National University of Ireland, Galway. <http://www.algaebase.org> (searched on 26 June 2017).
- Hall, R. I, Smol, J. P. (1992). A weighted—averaging regression and calibration model for inferring total phosphorus concentration from diatoms in British Columbia (Canada) lakes. *Freshwater Biology*, 27 (3): 417–434.
- Harwood, D. M., Gersonde, R. (1990). Lower Cretaceous diatoms from ODP Leg 113 Site 693 (Weddell Sea) part 2, resting spores, chrysophycean cysts, and endoskeletal dinoflagellates, and notes on the origins of diatoms. *Proceedings of the Ocean Drilling Program, Scientific Results*, 113: 403–425
- Hlúbiková, D. Hindáková, A., Haviar, M., Miettinen, J. (2007). Application of diatom water quality indices in influenced and non-influenced sites of Slovak rivers (Central
-

Europe). *Archiv für Hydrobiologie. Supplementband. Large rivers*, 17 (3 4): 443 464.

Hofmann, G., Werum, M., Lange-Bertalot, H. (2011). *Diatomeen im Süßwasser-Benthos von Mitteleuropa*. Ganter Verlag K. G., Germany, 908 pp.

http://www.hidmet.gov.rs/ciril/hidrologija/karakteristicne_v.php.

(16.09.2017.)

Hürlimann, J., P. Niederhauser (2002). *Méthodes d'étude et d'appréciation de l'état de Santé des Cours d'eau: Diatomées–Niveau R (Région)*. Office Fédéral de l'Environnement, des Forêts et du Paysage: Berne.

Illies, J. (1978). *Limnofauna Europaea*. 2. Auflage., Gustav Fischer Verlag: Stuttgart, New York, 532 pp.

Ivorra, N., Hettelaar, J., Tubbing, G.M.J., Kraak, M.H.S., Sabater, S., Admiraal, W. (1999). Translocation of microbenthic algal assemblages used for in situ analysis of metal pollution in rivers. *Archives of Environmental Contamination and Toxicology*, 37: 19–28.

Jakovljevi , O. S., Krizmani , J. Ž., Cvijan, M. V. (2014). Water quality assessment of the DTD canal system by diatom indices. *Matica Srpska Journal for Natural Sciences*, 127: 23 33.

Jakovljevi , O. S., Popovi , S. S., Vidakovi , D. P., Stojanovi , K. Z., Krizmani , J. Ž. (2016). The application of benthic diatoms in water quality assessment (Mlava River, Serbia). *Acta Botanica Croatica*, 75 (2): 199 205.

Jakovljevi , O., Popovi , S., Živi , I., Stojanovi , K., Krizmani , J. (2016). Benthic diatoms of the Vrla River (Serbia) and their application in the water ecological status assessment. *Oceanological and Hydrobiological Studies*, 45 (3): 304 315.

Jovanovi , M, Rosi , N. (2010). Prora un prelivanja kamenog praga u re nom koritu pri malim vodama. *Vodoprivreda* 0350-0519, 42 (243 245): 55 60.

Kahlert, M., Ács, É., Almeida, S. F., Blanco, S., Dreßler, M., Ector, L., Karjalainen, S.M, Liess, A., Mertens, A., van der Wal, J., Vilbaste, S. (2016). Quality assurance of diatom counts in Europe: towards harmonized datasets. *Hydrobiologia*, 772 (1): 1 14.

-
- Karadži , B. (2013). FLORA: A Software Package for Statistical Analysis of Ecological Data. *Water Research and Management*, 3 (2): 45–54.
- Karadži , B., Marinkovi , S. (2009). Kvantitativna ekologija. Insitut za biološka istraživanja „Siniša Stankovi “, Beograd, 489 str.
- Kawecka, B., Sanecki, J. (2003). *Didymosphenia geminata* in running waters of southern Poland—symptoms of change in water quality? *Hydrobiologia*, 495 (1–3): 193–201.
- Keithan, E. D., Lowe, R. L., DeYoe, H. R. (1988). Benthic diatom distribution in a Pennsylvania stream: role of pH and nutrients. *Journal of Phycology*, 24 (4): 581–585.
- Kelly, M. (2013). Data rich, information poor? Phytobenthos assessment and the Water Framework Directive. *European Journal of Phycology*, 48 (4): 437–450.
- Kelly, M. G. (2003). Short term dynamics of diatoms in an upland stream and implications for monitoring eutrophication. *Environmental Pollution*, 125 (2): 117–122.
- Kelly, M. G., Penny, C. J.,Whitton, B. A. (1995). Comparative performance of benthic diatom indices used to assess river water quality. *Hydrobiologia*, 302: 179–188.
- Kelly, M.G, Adams, C., Graves, A. C., Jamieson, J., Krokowski, J., Lycett, E. B., Murray-Bligh, J., Pritchard, S., Wilkins, C. (2001). *The Trophic Diatom Index: A User's Manual. Revised Edition.* Environment Agency, Bristol, 135 pp.
- Kelly, M.G., Whitton B.A. (1995). The Trophic Diatom Index: a new index for monitoring eutrophication in rivers. *Journal of Applied Phycology*, 7: 433–444.
- Kilroy, C., Larned, S. T. , Biggs, B. F. J. (2009). The non-indigenous diatom *Didymosphenia geminata* alters benthic communities in New Zealand rivers. *Freshwater Biology*, 54 (9): 1990–2002.
- Kireta, A.R., Reavie, E.D., Sgro, G.V., Angradi, T.R., Bolgrien, D.W., Hill, B.H., Jicha, T.M. (2012). Planktonic and periphytic diatoms as indicators of stress on great rivers of the United States: Testing water quality and disturbance models. *Ecological Indicators*, 13 (1): 222–231.
- Köhler, J. (1993). Growth, production and losses of phytoplankton in the lowland River Spree. I. Population dynamics. *Journal of Plankton Research*, 15 (3): 335–349.
- Kolarevi S., Kneževi -Vuk evi J., Paunovi M., Vasiljevi B., Kra un Margareta, Ga i Z., Vukovi -Ga ic B. (2012). Seasonal Variations of Microbiological Parameters of
-

-
- Water Quality of the Velika Morava River, Serbia. Archives of Biological Sciences, 64 (3): 1017–1027.
- Komatina, D., Grošelj, S. (2015). Transboundary Water Cooperation for Sustainable Development of the Sava River Basin. In: Milačić R., Ščanar J., Paunović M. (eds.). The Handbook of Environmental Chemistry Vol. 31: The Sava River. Springer, Heidelberg, New York, Dordrecht, London, pp. 229–248.
- Kovács, C., Kahlert, M., Padisák, J. (2006). Benthic diatom communities along pH and TP gradients in Hungarian and Swedish streams. Journal of Applied Phycology, 18 (2): 105–117.
- Krammer, K. (1997 a). Die cymbelloiden Diatomeen, Teil 1. Bibliotheca Diatomologica 36. J. Cramer, Berlin-Stuttgart, 382 pp.
- Krammer, K. (1997 b). Die cymbelloiden Diatomeen, Teil 2. Bibliotheca Diatomologica 37. J. Cramer, Berlin-Stuttgart, 469 pp.
- Krammer, K. (2000). Diatoms of Europe. Volume 1: The genus *Pinnularia*. A.R.G. Gantner Verlag K.G., 703 pp.
- Krammer, K. (2002). Diatoms of Europe. Volume 3: *Cymbella*. A.R.G. Gantner Verlag K.G., 584 pp.
- Krammer, K. (2003). Diatoms of Europe. Volume 4: *Cymbopleura*, *Delicata*, *Navicymbula*, *Gomphocymbellopsis*, *Afrocybella*. A.R.G. Gantner Verlag K.G., 530 pp.
- Krammer, K., Lange-Bertalot, H. (1986). Bacillariophyceae 1, Teil: Naviculaceae. In: Ettl, H., Gerloff, J., Heynig, H., Mollenhauer, D. (eds.). Süßwasser flora von Mitteleuropa, Band 2/1. Gustav Fischer Verlag, Jena, 876 pp.
- Krammer, K., Lange-Bertalot, H. (1988). Bacillariophyceae 2/2, Teil: Bacillariaceae, Epithemiaceae, Surirellaceae. In: H. Ettl, J. Gerloff, H. Heynig, D. Mollenhauer, (eds.). Süßwasserflora von Mitteleuropa, Band 2/2. Gustav Fischer Verlag, Stuttgart, 596 pp.
- Krammer, K., Lange-Bertalot, H. (2004). Bacillariophyceae 2/3, Teil: Centrales, Fragilariaceae, Eunotiaceae. In: H. Ettl, J. Gerloff, H. Heynig, D. Mollenhauer (eds.). Süßwasserflora von Mitteleuropa, Band 2/3. Gustav Fischer Verlag, Stuttgart, 598 pp.
-

-
- Krammer, K., Lange-Bertalot, H. (2011). Bacillariophyceae 2/4, Teil: Achnantheaceae, Kristische Ergänzungen zu Navicula (Lineolate) und Gomphonema. In: H. Ettl, J. Gerloff, H. Heynig, D. Mollenhauer (eds.). Süßwasserflora von Mitteleuropa, Band 2/4. Gustav Fischer Verlag, Stuttgart, 468 pp.
- Krebs, C. J. (2001). Ecology: the experimental analysis of distribution and abundance. Benjamin Cummings, an imprint of Addison Wesley Longman, Inc. 5th Ed., 695 pp.
- Krebs, C.J. (2014). Ecological Methodology, 3rd ed. (in prep). Chapter 13. Species Diversity Measures. <http://www.zoology.ubc.ca/~krebs/books.html>
- Krejci, M.E., Lowe, R.L. (1986). The importance of sand grain mineralogy and topography in determining micro-spatial distribution of epipsammic diatoms. Journal of the North American Benthological Society, 5: 221–229.
- Krizmani, J. (2009). Floristi ka, taksonomska i ekološka istraživanja silikatnih algi sa rafom (Bacillariohyceae, Bacillariophycideae, Bacillariophyta) Srbije. Doktorska disertacija. Univerzitet u Beogradu, Biološki fakultet, 595 str.
- Krizmani, J., Predojevi, D., Trbojevi, I., Vidakovi, D., Jakovljevi, O., Subakov-Simi, G. (2015). Expansion of invasive diatom species *Didymosphenia geminata* (Lyngb.) M.Schmidt and *Diadlesmis confervacea* (Grun.) Hustedt in the waters of Serbia. Abstract Book. 6th Balkan Botanical Congress. Rijeka, Hrvatska, pp. 81.
- Krizmani, J., Subakov Simi, G., Predojevi, D. (2013). Algae as water quality bioindicators of the River Djetinja. Proceedings of 6th International Conference “Water & Fish”. Belgrade, Zemun, pp. 342–348.
- Kumar, S., Spaulding, S. A., Stohlgren, T. J. (2008). Potential habitat distribution for the freshwater diatom *Didymosphenia geminata* in the continental US. Frontiers in Ecology and the Environment. 7 (8): 415–420.
- Kupferberg, S. (1997). Facilitation of periphyton production by tadpole grazing: functional differences between species. Freshwater Biology, 37: 427–439.
- Kwandrans, J., Eloranta, P., Kawecka, B., Woitan, K. (1998). Use of benthic diatom communities to evaluate water quality in rivers of Southern Poland. Journal of Applied Phycology, 10 (2): 193–201.
- Lai, G. G., Padedda, B. M., Pulina, S., Viridis, T., Sechi, N., Lugliè, A. (2010). Diatoms and quality of watercourses in North-Central Sardinia. Vie et Milieu, 60 (3): 209–216.
-

-
- Lamberti, G. A. (1996). The role of periphyton in benthic food webs. In: Stevenson, R. J., Bothwell, M. L., Lowe, R. L. (eds.). *Algal Ecology: Freshwater Benthic Ecosystems*. Academic Press, San Diego, CA., pp. 533–573.
- Lange-Bertalot, H. (1993). 85 Neue Taxa und über 100 weitere neu definierte Taxa ergänzend zur Süßwasserflora von Mitteleuropa, Volume 2/1-4. *Bibliotheca Diatomologica*, 27. J. Cramer, Berlin-Stuttgart, 454 pp.
- Lange-Bertalot, H. (2001). *Diatoms of Europe. Volume 2: Navicula sensu stricto, 10 genera separated from Navicula sensu lato, Frustulia*. A.R.G. Gantner Verlag K.G, 526 pp.
- Laušević, R., Nikitović, J., Tomašević, V. (1998). Phytoplankton in River Sava near Belgrade. *Ekologija*, 33 (1–2): 29–40.
- Leclercq, L., Maquet, B. (1987). Deux nouveaux indices chimique et diatomique de qualité d'eau courante. Application au Samson et à ses affluents. Comparaison avec d'autres indices chimiques, biocénologiques et diatomiques. Institut Royal des Sciences Naturelles de Belgique, 113 pp.
- Lecointe, C., Coste, M., Prygiel, J. (1993). Omnidia: software for taxonomy, calculation of diatom indices and inventories management. *Hydrobiologia*, 269/270: 509–513.
- Leland, H. V. (1995). Distribution of phytobenthos in the Yakima River basin, Washington, in relation to geology, land use and other environmental factors. *Canadian Journal of Fisheries and Aquatic Sciences*, 52 (5): 1108–1129.
- Lenoir, A., Coste, M. (1996). Development of a practical diatom index of overall water quality applicable to the French National Water Board network. In: Whitton B. A., Rott E. (eds.). *Use of Algae for Monitoring Rivers II*. Institut für Botanik, Universität Innsbruck, 29–43.
- Levkov, Z., Metzeltin, D., Pavlov, A. (2013). Luticola and Luticolopsis. In: Lange-Bertalot, H. (ed.). *Diatoms of Europe: Diatoms of the European Inland Waters and Comparable Habitats. Volume 7*. Koeltz Scientific Books, Königstein, 698 pp.
- Liška, I., Wagner, F., Sengl, M., Deutsch, K., Slobodnik, J. (2015). *Joint Danube Survey 3. A Comprehensive Analysis of Danube Water Quality*. International Commission for the Protection of the Danube River, Vienna, Austria, 369 pp.
-

-
- Lobo, E. A., Callegaro, V. L., Bender, P. (2002). Utilização de algas diatomáceas epilíticas como indicadores da qualidade da água em rios e arroios da Região Hidrográfica do Guaíba, RS, Brasil. Santa Cruz do Sul: EDUNISC, 126 pp.
- Lowe, R. L. (1974). Environmental requirements and pollution tolerance of freshwater diatoms. USEPA 670/4-74-005. USEPA. Cincinnati, OH. <https://nepis.epa.gov>
- Lowe, R. L. (2011). The importance of scale in understanding the natural history of diatom communities. In: Seckbach, J., Kociolek, P. (eds.). The Diatom World. Vol. 19 of the series Cellular Origin, Life in Extreme Habitats and Astrobiology. Springer Netherlands, pp. 295–311.
- Makovinska, J., Hlubikova, D. (2015). Phytobenthos of the River Danube. In: Liska, I. (ed.). The Handbook of Environmental Chemistry, Vol. 39: The Danube River Basin. Springer Berlin Heidelberg, pp. 317–340.
- Mann D.G., Droop S.J.M. (1996) Biodiversity, biogeography and conservation of diatoms. In: Kristiansen J. (ed.). Biogeography of Freshwater Algae. Developments in Hydrobiology, Vol. 118. Springer, Dordrecht, pp 19–32.
- Manoylov, K. M. (2009). Intra- and interspecific competition for nutrients and light in diatom cultures. *Journal of Freshwater Ecology*, 24 (1): 145–157.
- Mantel, N., (1967). The detection of disease clustering and a generalized regression approach. *Cancer Research*, 27 (2): 209–220.
- Marinković, N., Krizmani, J., Karadžić, V., Karadžić, B., Vasiljević, B., Paunović, M. (2016). Algal diversity along the Serbian stretch of the Sava River. *Water Research and Management*, 6 (2): 27–33.
- Marković, V., Atanacković, A., Tubić, B., Vasiljević, B., Simić, V., Tomović, J., Nikolić, V., Paunović, M. (2011). Indicative status assessment of the Velika Morava River based on aquatic macroinvertebrates. *Water Research and Management*, 1 (3): 47–53.
- Martinović-Vitanović, V. (1996). Ekološka studija Obedske bare. Javno preduzeće za gazdovanje šumama „Srbija šume”, Beograd, Geokarta.
- Martinović-Vitanović, V., Kalafatić, V. (2002). Limnological investigations of the Tisa River in Yugoslavia. Joint Investigations of the Danube River on the Territory of the FR Yugoslavia within the International JDS-ITR Program. Public Report of Serbia,
-

-
- Ministry for Protection of Natatural Resources and Environmental and Federal Hydrometeorological Institute, Belgrade, pp. 29 52.
- Martinovi -Vitanovi , V., Kalafati , V. (2004). Kvalitet vode Save na podru ju Beograda u 2003. godini – Saprobiološka analiza. *Vodoprivreda*, 36: 385 391.
- Maznah, W. O., Mansor, M. (2002). Aquatic pollution assessment based on attached diatom communities in the Pinang River Basin, Malaysia. *Hydrobiologia*, 487 (1): 229 241.
- McCormick, P., Cairns, J. (1994). Algae as indicators of environmental change. *Journal of Applied Phycology*, 6: 509 526.
- McGarrigle, ., Lucey, ., Ó Cinnéide, . (2010). Water quality in Ireland 2007-2009. Apendix 3.1 of the Report. Environmenat Protection Agency Ireland. <http://www.epa.ie/pubs/data/water/riverstatusassessment.html>
- Medley, C. N., Clements, W. H. (1998). Responses of diatom communities to heavy metals in streams: the influence of longitudinal variation. *Ecological applications*, 8 (3), 631 644.
- Medlin, L. K. (2014). Evolution of the Diatoms: VIII. Re-examination of the SSU-Rrna gene using multiple outgroups and a cladistic analysis of valve features. *Journal of Biodiversity, Bioprospecting and Development*, 1 (3): 1 16.
- Medlin, L. K. (2016). Evolution of the diatoms: major steps in their evolution and a review of the supporting molecular and morphological evidence. *Phycologia*, 55 (1): 79 103.
- Medlin, L. K., Kaczmarska, I. (2004). Evolution of the diatoms: V. Morphological and cytological support for the major clades and a taxonomic revision. *Phycologia*, 43 (3): 245 270.
- Meteorološki i hidrološki bilten br. 9 (2011). Državni hidrometeorološki zavod Republike Hrvatske, Zagreb.
- Meteorološki i hidrološki bilten br. 9 (2012). Državni hidrometeorološki zavod Republike Hrvatske, Zagreb.
- Meteorološki i hidrološki bilten br. 9 (2014). Državni hidrometeorološki zavod Republike Hrvatske, Zagreb.
- Meteorološki i hidrološki bilten br. 9 (2015). Državni hidrometeorološki zavod Republike Hrvatske, Zagreb.
-

-
- Morales, E. A., Manoylov, K. M. (2009). *Mayamaea cahabaensis* sp. nov. (Bacillariophyceae), a new freshwater diatom from streams in the southern United States. *Proceedings of the Academy of Natural Sciences of Philadelphia*, 158 (1): 49–59.
- Morin, S., Cordonier, A., Lavoie, I., Arini, A., Blanco, S., Duong, T. T., Tornés, E., Bonet, B., Corcoll, N., Faggiano, L., Laviale, M., Pérès, F., Becares, E., Coste, M., Feurtet-Mazel, A., Fortin, C., Guasch, H., Sabater, S. (2012). Consistency in diatom response to metal-contaminated environments. In: Guasch, H., Ginebreda, A., Geislinger, A. (eds.). *Emerging and priority pollutants in rivers*. Springer Berlin Heidelberg, pp. 117–146.
- Morin, S., Duong, T.T., Dabrin, A., Coynel, A., Herlory, O., Baudrimont, M., Delmas, F., Durrieu G., Schafer, J., Winterton, P., Blanc, G., Coste, M. (2008). Long-term survey of heavy-metal pollution, biofilm contamination and diatom community structure in the Riou Mort watershed, SouthWest France. *Environmental Pollution*, 151: 532–542.
- Moss, B. (1977). Adaptations of epipelic and epipsammic freshwater algae. *Oecologia*, 28: 103–108.
- Navarro-Ortega, A., Acuña, V., Bellin, A., Burek, P., Cassiani, G., Choukr-Allah, R., Dolédec, S., Elosegi, A., Ferrari, F., Ginebreda, A., Grathwohl, P., Jones, C., Rault, P.K., Kok, K., Koundouri, P., Ludwig, R.P., Merz, R., Milacic, R., Munoz, I., Nikulin, G., Paniconi, C., Paunovi, M., Petrovi, M., Sabater, L., Sabaterb, S., Skoulikidis, N.T., Slob, A., Teutsch, G., Voulvoulis, N., Barcelo, D. (2015). Managing the effects of multiple stressors on aquatic ecosystems under water scarcity. The GLOBAQUA project. *Science of the Total Environment*, 503: 3–9.
- Ndiritu, G. G., Gichuki, N. N., Triest, L. (2006). Distribution of Epilithic Diatoms in Response to Environmental Conditions in an Urban Tropical Stream, Central Kenya. *Biodiversity and Conservation*, 15 (10): 3267–3293.
- Newall, P., Walsh, C.J. (2005). Response of epilithic diatom assemblages to urbanization influences. *Hydrobiologia*, 532: 53–67.
- Novais, M. H. B. D. C. (2011). Estudo das diatomáceas bênticas em sistemas lóticos de Portugal Continental. PhD Thesis, Universidade de Évora, 50 pp.
-

-
- Obuškovi , Lj. D. (1979). Pojava „vodenog cveta“ vrste *Stephanodiscus astrea* var. *minutula* (Kg.) Grun. u Savskom jezeru kod Beograda. *Biosistematika*, 5 (2): 127–138.
- Obuškovi , Lj., Kalafati , V. (1979). Ispitivanje planktona Morave u uslovima poja anog zaga enja. Prethodno saopštenje. Drugi Kongres ekologija Jugoslavije, Zagreb, str. 1889–1903.
- Obuškovi , Lj., Kalafati , V., Markovi , L. (1985). Višegodišnja ispitivanja kvaliteta vode reke Save u regionu Beograda na osnovu planktona kao bioindikatora. „Zaštita voda ’85“, Sarajevo. Knjiga 1:109–112.
- Obuškovi , Lj., Markovi , L. (1987). Fitoplankton i saprobiološke karakteristike reke Save u 1984. Rijeka Sava, Zaštita i koriš enje voda ’87, Zagreb. Zbornik radova, pp. 426–431.
- Obuškovi , Lj., Maslikovi , M. (1997). Studies of phytoplankton and some chemical parameters of the river Danube waters at river section km 1162–1115. *Archives of Biological Sciences*, 49: 37–42.
- Pascher, A. (1921). Über die einstimmungen zwischen Diatomeen, Heterokonten und Chrysoomonaden. *Berichte der Deutsch Botanisches Gesellschaft*, 39: 236–240.
- Passy, S. (2007). Diatom ecological guilds display distinct and predictable behavior along nutrient and disturbance gradients in running waters. *Aquatic Botany*, 86: 171–178.
- Patrick, R. (1971). The effects of increasing light and temperature on the structure of diatom communities. *Limnology and Oceanography*, 16 (2): 405–421.
- Patrick, R., Crum, B., Coles, J. (1969). Temperature and manganese as determining factors in the presence of diatom or blue-green algal floras in streams. *Proceedings of the National Academy of Sciences*, 64 (2): 472–478.
- Paunovi , M. (2007). Struktura zajednica makroinvertebrata kao indikator tipova teku ih voda Srbije. Doktorska disertacija. Univerzitet u Beogradu, Biološki fakultet, 218 str.
- Peterson, C. G. (1996). Response of benthic algal communities to natural physical disturbance. In: Stevenson, R. J., Bothwell, M. L., Lowe, R. L. (eds.). *Algal Ecology: Freshwater Benthic Ecosystems*. Academic Press, San Diego, CA., pp. 375–402.
-

-
- Poikane, S., Kelly, M., Cantonati, M. (2016). Benthic algal assessment of ecological status in European lakes and rivers: Challenges and opportunities. *Science of the Total Environment*, 568: 603–613.
- Popovic, A., Djodjevic, D., Polic, P. (2001). Trace and major element pollution originating from coal ash suspension and transport processes. *Environment International*, 26 (4): 251–255.
- Potapova, M., Charles, D. F. (2003). Distribution of benthic diatoms in US rivers in relation to conductivity and ionic composition. *Freshwater Biology*, 48 (8): 1311–1328.
- Potapova, M.G., Charles, D.F. (2002). Benthic diatoms in USA rivers: distributions along spatial and environmental gradients. *Journal of Biogeography*, 29 (2): 167–187.
- Predojevi, D. (2017). Procena ekološkog statusa reke Zasavice na osnovu algoloških parametara. Doktorska disertacija. Univerzitet u Beogradu, Biološki fakultet, 329 str.
- Pregled hidroloških razmer površinskih voda v Sloveniji (2011). Poročilo o monitoringu za leto 2011. Agencija Republike Slovenije za okolje, Ljubljana.
- Pregled hidroloških razmer površinskih voda v Sloveniji (2012). Poročilo o monitoringu za leto 2012. Agencija Republike Slovenije za okolje, Ljubljana.
- Pregled hidroloških razmer površinskih voda v Sloveniji (2014). Poročilo o monitoringu za leto 2014. Agencija Republike Slovenije za okolje, Ljubljana.
- Pregled hidroloških razmer površinskih voda v Sloveniji (2015). Poročilo o monitoringu za leto 2015. Agencija Republike Slovenije za okolje, Ljubljana.
- Prygiel, J., Coste, M. (1993). The assessment of water quality in the Artois Picardie water basin (France) by the use of diatom indices. *Hydrobiologia*, 269/270: 343–349.
- Prygiel, J., Carpentier, P., Almeida, S., Coste, M., Druart, J.-C., Ector, L., Guillard, D., Honoré, M. A., Iserentant, R., Ledgeanck, P., Lalanne-Cassou, C., Lesniak, C., Mercier, I., Moncaut, P., Nazart, M., Nouchet, N., Peres, F., Peeters, V., Rimet, F., Rumeau, A., Sabater, S., Straub, F., Torrisi, M., Tudesque, L., v.d. Vijver, B., Vidal, H., Vizinet, J., Zydek, N. (2002). Determination of the biological diatom index (IBD NF T 90-354): results of an intercalibration exercise. *Journal of Applied Phycology*, 14: 19–26.
-

-
- Prygiel, J., Coste, M. (2000). Guide méthodologique pour la mise en oeuvre de l'Indice Biologique Diatomées. Agences de l'Eau—Cemagref, Bordeaux, 178 pp.
- Prygiel, J., Leveque, L., R. Iseretant (1996). Un nouvel indice diatomique pratique pour l'évaluation de la qualité des eaux en réseau de surveillance. *Revue des Sciences de l'Eau*, 9: 97–113.
- Pujin, V., Stojkovi, S., Duku, N., Miljanovi, B., Maletin, S., Sekuli, A. Todorovi, I. (1999). Hidrobionti pokazatelji kvaliteta vode reke Tise. 28. Konferencija o aktuelnim problemima zaštite voda "Zaštita voda '99", Soko Banja, str. 230–242.
- Rimet, F., Bouchez, A. (2012). Biomonitoring river diatoms: implications of taxonomic resolution. *Ecological indicators*, 15 (1): 92–99.
- Rott, E., Hofmann, G., Pall, K., Pfister, P., Pipp, E., (1997). Indikationslisten für Aufwuchsalgen, Teil 1: Saprobielle Indikation. Bundesministerium für Land- und Forstwirtschaft, Wien.
- Rott, E., Pfister, P., Van Dam, H., Pipp, E., Pall, K., Binder, N., Ortler, K. (1999). Indikationslisten für Aufwuchsalgen. Teil 2: Trophieindikation sowie geochemische Präferenz, taxonomische und toxikologische Anmerkungen. Bundesministerium für Land und Forstwirtschaft, Wien.
- Round, F. E. (1990). Diatom communities their response to changes in acidity. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, pp. 243–249.
- Round, F. E. (1991). Diatoms in river water-monitoring studies. *Journal of Applied Phycology*, 3 (2): 129–145.
- Round, F. E., Lee, K. (1989). Studies on freshwater Amphora species IV. The Amphora epiphytic on other diatoms. *Diatom Research*, 4 (2): 345–349.
- Round, F.E., Crawford, R.M., Mann, D.G. (1990). *The Diatoms. Biology and Morphology of the Genera*. Cambridge University press, Cambridge, 747 pp.
- Ruck, E. C., Theriot, E. C. (2011). Origin and evolution of the canal raphe system in diatoms. *Protist*, 162 (5): 723–737.
- Rumeau, A., M. Coste (1988). Initiation à la systématique des diatomées d'eau douce. *Bulletin Francais de la Peche et de la Pisciculture*, 309: 1–69.
-

-
- Ržanić, A. M., Cvijan, M. V., Krizmanić, J. (2005). Phytoplankton of the Tisa river. *Archives of Biological Sciences*, 57 (3): 223–235.
- Sabater, S. (2000). Diatom communities as indicators of environmental stress in the Guadiamar River, SW. Spain, following a major mine tailings spill. *Journal of Applied Phycology*, 12 (2): 113–124.
- Sava River Basin Analysis Report (2009). International Sava River Basin Commission, Zagreb, Croatia, 289 pp. <http://www.savacommission.org/publication>
- Sava River Basin Management Plan (2013). Background paper No. 1. Surface water bodies in the Sava River Basin. International Sava River Basin Commission, Zagreb, Republic of Croatia, 90 pp.
- Sava River Basin Management Plan (2014). International Sava River Basin Commission, Zagreb, Republic of Croatia, 240 pp.
- Schöll, F., Birk, S., Böhmer, J. (2012). XGIG Large River Intercalibration Exercise – WFD Intercalibration Phase 2: Milestone 6 Report., 73 pp.
- Schwarz, U. (2016). Sava White Book. The River Sava: Threats and Restoration Potential. Radolfzell/Wien: EuroNatur/Riverwatch, 144 pp.
- Shannon, C.E. (1948). A mathematical theory of communication. *Bell System Technical Journal*, 27: 379–423.
- Shapiro, S.S., Wilk, M.B., Chen, H.J. (1968). A comparative study of various tests for normality. *Journal of the American Statistical Association*, 63 (324): 1343–1372.
- Simić, B. S., Karadžić, R. V., Cvijan, V. M., Vasiljević, M. B. (2015). Algal Communities Along the Sava River. In: Milačić, R., Štanić, J., Paunović, M. (eds.). *The Handbook of Environmental Chemistry Vol. 31: The Sava River*. Springer Berlin Heidelberg. pp 229–248.
- Simić, B. S., Čorović, N., Mitrović, A. (2016): Diversity of algae in protected natural habitats – the Mlava springhead and the Krupaj springhead. 2nd International Symposium on Nature Conservation “Nature Conservation-experiences and perspectives”, Book of Proceedings, Institute for nature Conservation of Vojvodina Province, Novi Sad, pp. 105–119.
-

-
- Simi , B. S., Simi , M. V. (2012). Ekologija kopnenih voda. Hidrobiologija I. Drugo izdanje. Univerzitet u Beogradu, Biološki fakultet i Univerzitet u Kragujevcu, Prirodno-matemati ki fakultet. Zemun. Alta Nova, 295 str.
- Simi , S., Pantovi , N. (2010). Observations on the rare alga *Thorea hispida* (Thore) Desvaux (Rhodophyta) from Serbia. *Cryptogamie Algologie*, 31 (3): 343–353.
- Simi , S.B., or evi , N.B., Vasiljevi , B.M. (2014). New record of red alga *Thorea hispida* (Thore) Desvaux (Rhodophyta) in the River Sava (Sremska Mitrovica, Serbia). *Water Research and Management*, 4 (1): 47–51.
- Simonovi , P., Piria, M., Zuliani, T., Ili , M., Marinkovi , N., Kra un-Kolarevi , M., Paunovi , M. (2017). Characterization of sections of the Sava River based on fish community structure. *Science of the Total Environment*, 574: 264–271.
- Sims, P. A., Mann, D. G., Medlin, L. K. (2006). Evolution of the diatoms: insights from fossil, biological and molecular data. *Phycologia*, 45 (4): 361–402.
- Sládec k, V. (1986). Diatoms as indicators of organic pollution. *Acta Hydrochimica et Hydrobiologia*, 14 (5): 555–566.
- Smucker, N. J., Vis, M. L. (2010). Using diatoms to assess human impacts on streams benefits from multiple-habitat sampling. *Hydrobiologia*, 654 (1): 93–109.
- Soininen, J., Paavola, R., Muotka, T. (2004). Benthic diatom communities in boreal streams: community structure in relation to environmental and spatial gradients. *Ecography*, 27 (3), 330–342.
- Stal, L.J. (2010). Microphytobenthos as a biogeomorphological force in intertidal sediment stabilization. *Ecological Engineering*, 36: 236–245.
- StatSoft Inc. (2001). STATISTICA for Windows, version 6.0. www.statsoft.com
- Steinberg, C., Schiefele, S. (1988). Biological indication of trophy and pollution of running waters. *Wasser Abwasser Forsch*, 21: 227–234.
- Steinman, A. D. (1996). Effects of grazers on freshwater benthic algae. In: Stevenson, R. J., Bothwell, M. L., Lowe, R. L. (eds.). *Algal Ecology: Freshwater Benthic Ecosystems*. Academic Press, San Diego, CA., pp. 341–373.
- Steinman, A. D., McIntire, C. D., Gregory, S.V., Lamberti, G. A., Ashkenas, L. R. (1987). Effects on herbivore type and density on taxonomic structure and physiognomy of
-

-
- algal assemblages in laboratory streams. *Journal of North American Benthological Society*, 6: 175–188.
- Stevenson, R. J. (1984). Epilithic and epipellic diatoms in the Sandusky River, with emphasis on species diversity and water pollution. *Hydrobiologia*, 114 (3): 161–175.
- Subakov-Simi, G., Cvijan, M. (2004). *Didymosphenia geminata* (Lyngb.) M. Schmidt (Bacillariophyta) from the Tisa River (Serbia) – its distribution and specific morphological and ecological characteristics. *Algological Studies*, 114 (1): 53–66.
- Szabó, K., Kiss, K. T., Taba, G., Ács, É. (2005). Epiphytic diatoms of the Tisza River, Kisköre Reservoir and some oxbows of the Tisza River after the cyanide and heavy metal pollution in 2000. *Acta Botanica Croatica*, 64 (1): 1–46.
- Ter Braak, C.J. (1987). The analysis of vegetation-environment relationships by canonical correspondence analysis, in *Theory and models in vegetation science*. Springer Netherlands, pp. 69–77.
- Tiffany, M. A., Lange, C. B. (2002). Diatoms provide attachment sites for other diatoms: a natural history of epiphytism from southern California. *Phycologia*, 41 (2): 116–124.
- TNMN Yearbook (2004). Water quality in the Danube River Basin 2004. International Commission for the Protection of the Danube River, 36 pp.
- Trábert, Z., Tihamér Kiss, K., Várбірó, G., Dobosy, P., Grigorszky, I., Ács, É. (2017). Comparison of the utility of a frequently used diatom index (IPS) and the diatom ecological guilds in the ecological status assessment of large rivers. *Fundamental and Applied Limnology/Archiv für Hydrobiologie*, 189 (2): 87–103.
- Tuchman, N. C. (1996). The role of heterotrophy in algae. In: Stevenson, R. J., Bothwell, M. L. and Lowe, R. L. (eds.). *Algal Ecology: Freshwater Benthic Ecosystems*. Academic Press, San Diego, CA., pp. 229–319.
- Tuchman, N. C., Stevenson, J. R. (1991). Effects of Selective Grazing by Snails on Benthic Algal Succession. *Journal of North American Benthological Society*, 10 (4): 430–443.
- Urošević, V. (1994). Alge visokoplaninskih jezera Sirini ke strane Šar-planine. Doktorska disertacija. Univerzitet u Prištini.
-

-
- Van Dam, H., Mertens, A., Sinkeldam, J. (1994). A coded checklist and ecological indicator values of freshwater diatoms from the Netherlands. *Aquatic Ecology*, 28 (1): 117–133.
- Van Dam, H., Padisák, J., Kovács, C. (2005). BQE Report Phytobenthos, Ecosurv, Ministry of Environment and Water, Hungary, EuropeAid/114951/D/SV/2002-000-180-04-01-02-02, 54 pp.
- Vannote, R. L., Minshall, G. W., Cummins, K. W., Sedell, J. R., Cushing, C. E. (1980). The river continuum concept. *Canadian Journal of Fisheries and Aquatic Sciences*, 37 (1): 130–137.
- Várbíró, G., Borics, G., Csányi, B., Fehér, G., Grigorszky, I., Kiss, K.T., Tóth, A., Ács, É. (2012). Improvement of the ecological water qualification system of rivers based on the first results of the Hungarian phytobenthos surveillance monitoring. *Hydrobiologia*, 695 (1): 125–135.
- Vasiljević B., Krizmanić J., Ilić M., Marković V., Tomović J., Zorić K., Paunović M. (2014). Water quality assessment based on diatom indices – small hilly streams case study. *Water Research and Management*, 4 (2): 31–35.
- Vasiljević B., Simić S. B., Paunović M., Zuliani, T., Krizmanić J., Marković V., Tomović J. (2017). Contribution to the improvement of diatom-based assessments of the ecological status of large rivers–The Sava River Case Study. *Science of The Total Environment*, 605-606: 874–883.
- Vidaković D. (2013). Procena ekološkog statusa reke Raške na osnovu epilitskih silikatnih algi. Master rad. Univerzitet u Beogradu, Biološki fakultet, Beograd.
- Vidmar, J., Zuliani, T., Novak, P., Drinić A., Štanić J., Milačić R. (2016). Elements in water, suspended particulate matter and sediments of the Sava River. *Journal of Soils and Sediments*, pp. 1–11.
- Vilbaste, S. (2004). Application of diatom indices in the evaluation of the water quality in Estonian streams. *Proceedings of the Estonian Academy of Sciences, Biology and Ecology*, 53 (1): 37–51.
- Vinson, D. K., Rushforth, S. R. (1989). Diatom species composition along a thermal gradient in the Portneuf River, Idaho, USA. *Hydrobiologia*, 185 (1): 41–54.
-

-
- Vrzel, J., Vuković-Gajić, B., Kolarević, S., Gajić, Z., Kraun-Kolarević, M., Kostić, J., Aborgiba, M., Farnleitner, A., Reischer, G., Linke, R., Paunović, M., Ogrinc, N. (2016). Determination of the sources of nitrate and the microbiological sources of pollution in the Sava River Basin. *Science of the Total Environment*, 573: 1460–1471.
- Wallin, M., T. Wiederholm, R. K. Johnson (2003). Guidance on establishing reference conditions and ecological status class boundaries for inland surface waters. CIS Working Group 2 (3), 93 pp.
- Washington, H. G. (1984). Diversity, biotic and similarity indices: a review with special relevance to aquatic ecosystems. *Water Research*, 18 (6): 653–694.
- Watanabe, T., Asai, K., Houki, A. (1990). Numerical simulation of organic pollution in flowing waters. *Encyclopedia of Environmental Control Technology*, Vol. 4. Hazardous Waste Containment and Treatment. P. N. Cheremisinoff., Gulf Publishing Company Houston, pp. 251–284.
- Werner, P., Adler, S., Dreßler, M. (2016). Effects of counting variances on water quality assessments: implications from four benthic diatom samples, each counted by 40 diatomists. *Journal of Applied Phycology*, 28 (4): 2287–2297.
- Werner, P., Köhler, J. (2005). Seasonal Dynamics of Benthic and Planktonic Algae in a Nutrient-Rich Lowland River (Spree, Germany). *International review of hydrobiology*, 90 (1): 1–20.
- WFD (2000). Water Framework Directive - Directive of European Parliament and of the Council 2000/60/EC – Establishing a Framework for Community Action in the Field of Water Policy. European Union, the European Parliament and Council, Luxembourg.
- WFD CIS Guidance Document No 13. (2005). Common implementation strategy for the water framework directive (2000/60/EC). Overall approach to the classification of ecological status and ecological potential. Luxembourg: Office for Official Publications of the European Communities.
- WFD CIS Guidance Document No 2. (2003). Common implementation strategy for the water framework directive (2000/60/EC). Identification of Water Bodies. Luxembourg: Office for Official Publications of the European Communities.
-

-
- Whitton, B. A., Ellwood, N. T. W., Kawecka, B. (2009). Biology of the freshwater diatom *Didymosphenia*: a review. *Hydrobiologia*, 630 (1): 1–37.
- Wu, N., Schmalz, B., Fohrer, N. (2010). Distribution of phytoplankton in a German lowland river in relation to environmental factors. *Journal of Plankton Research*, 33 (5): 807–820.
- Yendle, P. W., MacFie, H. J. (1989). Discriminant principal components analysis. *Journal of chemometrics*, 3 (4): 589–600.
- Zelinka, M., Marvan, P. (1961). Zur Präzisierung der biologischen Klassifikation der Reinheit fließender Gewässer. *Arch. Hydrobiol*, 57: 389–407.
- Ziemann, H., Nolting, E., Rustige, K. H. (1999). Bestimmung des Halobienindex. *Biologische Gewässeruntersuchung. Methoden der Biologischen Gewässeruntersuchung*, 2: 310–313.
- „...“ (2014). , 227 .
- „...“ (2015). , 232 .
- „...“ (2014). , 202 .
- 2011.
- (2011). , 633 .
- „...“ (2010). : , „...“ (). – . 157–182.

