We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,600 Open access books available 177,000





Our authors are among the

TOP 1% most cited scientists





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



The Phytoestrogens, Calcitonin and Thyroid Hormones: Effects on Bone Tissue

Branko Filipović and Branka Šošić-Jurjević University of Belgrade, Institute for Biological Research"Siniša Stanković" Serbia

1. Introduction

The skeleton is a metabolically active organ that undergoes remodeling throughout life. This involves a complex process by which old bone is continuously replaced by new tissue. Bone remodeling refers to the sequential, coupled actions of osteoclasts and osteoblasts. In conditions of sex hormone deficiency during advancing age, after the menopause or andropause, the rate of remodeling increases and bone formation is reduced relative to resorption. These alterations can cause microarchitectural deterioration of bone tissues, which increases bone loss as a predisposition to the occurrence of osteoporosis (Rehman et al., 2005). However, in contrast to postmenopausal osteoporosis in women, age-related bone loss in men is less well defined.

Numerous studies attest to the importance of estrogen in bone remodeling, evident from the finding that hormone replacement therapy (HRT) administered in a dose-dependent manner effectively prevented bone loss in postmenopausal women (Lindsay et al., 1976, 1984). However, in addition to protective effects on bone, HRT is associated with an increased risk for breast, endometrial, ovarian or prostate cancers (Davison & Davis, 2003; Loughlin & Richie, 1997; Nelson et al., 2002). Therefore, it is important to examine alternative approaches for prevention and treatment of osteoporosis without side effects. It is well known that the incidence of osteoporosis-related fractures is significantly lower in Southern and Eastern Asian women than in Western women (Tham et al., 1998). One possible reason for this difference is a high intake of phytoestrogen-rich plants, which Asian people eat more often than Western people (Ho et al., 2003). As a result, over the past decade a number of clinical trials for prevention of bone loss have assessed the effectiveness of plant derived non-steroidal phytoestrogens found in a wide variety of foods, most notably soybean. Isoflavones, which include daidzein and genistein are a class of phytoestrogens that act like estrogens. Since these compounds bind to estrogen receptors (ERs) and have estrogen-like activity (Branca, 2003), they have attracted much attention because of their potential benefit in the prevention and treatment of osteoporosis.

In addition to the phytoestrogen-mediated protective mechanisms against bone loss, recent evidence suggests that daidzein may also act on rat bone tissue through enhancement of thyroid C cell activity (Filipović et al., 2010). Namely, thyroid C cells produce the hormone, calcitonin (CT), which lowers plasma calcium concentration by suppressing osteoclast activity. Synthesis of CT and its release from C cells were decreased in conditions of gonadal hormone deficiency (Filipović et al., 2003, 2007; Isaia et al., 1989; Lu et al., 2000; Sakai et al., 2000). Due to its osteoprotective properties, CT is widely applied in the therapy of osteoporosis.

It is known that parathyroid hormone (PTH) is a major factor involved in the systemic regulation of bone resorption. Phytoestrogens may affect the parathyroid gland and reduce PTH secretion (Wong et al., 2002), suggesting that one way in which these compounds inhibit bone loss may be through reducing PTH levels.

Thyroid hormones are essential for normal bone maturation *in utero* and during early life. In adults an excess of thyroid hormones in the body affects the remodeling system in cortical and trabecular bone and may contribute to the development of osteoporosis (Kung, 1994). Receptors for these hormones are present in bone cells and they may directly increase bone resorption (Abu et al., 1997; Rizzoli et al., 1986). Additionally, thyroid-stimulating hormone (TSH), which stimulates the release of thyroid hormones, positively influences bone remodeling. Therefore, demonstrating both anabolic and antiresorptive effects, TSH may represent a promising candidate for the treatment of osteoporosis (Sendak et al., 2007).

In this chapter we will describe the known effects of phytoestrogens on bone. In addition to the direct action of these plant compounds, special attention will be paid to their influence on thyroid C and follicular cells, as producers of CT and thyroid hormones, using the latest data in the literature and our own results. These hormones, together with PTH may be involved in the indirect effects of phytoestrogens on bone tissue.

2. Bone cells and bone remodeling

Bone is a dynamic organ that undergoes remodeling throughout life. This process results from the separate action of bone forming cells called osteoblasts and bone resorbing cells called osteoclasts. Osteoblasts are responsible for the production of bone matrix constituents and are found in clusters on bone surfaces (Fig 1). They originate from multipotent mesenchymal stem cells, which have the capacity to differentiate into osteoblasts or other cells, such as adipocytes, chondrocytes, myoblasts and fibroblasts (Bianco et al., 2001). A mature osteoblast that is trapped in the bone matrix and remains isolated in lacunae becomes an osteocyte. (Fig.1). Bone formation involves production and maturation of the osteoid matrix, followed by mineralization of the matrix. Osteoblasts produce growth factors, such as insulin-like growth factor (IGF), platelet-derived growth factor (PDGF), basic fibroblast growth factor (bFGF), transforming growth factor- β (TGF- β) and bone morphometric protein (BMP) (Canalis et al., 1993, 1993a; Chen et al., 2004; Globus et al., 1989; Rydzel et al., 1994). These factors regulate osteoblast activity in an autocrine and paracrine manner.

Osteoclasts are large multinucleate cells responsible for bone resorption. They are derived from hematopoetic cells of the mononuclear lineage (Teitelbaum, 2000) (Fig.1). Osteoclasts have an abundant Golgi complex, mitochondria and transport vesicles loaded with lysosomal enzymes, such as tartrate-resistant acid phosphatase (TRAP) and cathepsin K. These enzymes are secreted via the specialized (ruffler border) plasma membrane of osteoclasts into the bone-resorbing compartment (Väänänen et al., 2000). The process of osteoclast attachment to the bone is complex and involves binding of integrins expressed in osteoclasts with specific amino acid sequences within proteins at the surface of the bone matrix and cytoskeleton activation (Davies et al., 1989; Reinholt et al., 1990). Dynamic structures, called podozomes allow movement of osteoclasts across the bone surface. Bone resorption occurs due to acidification and proteolysis of the bone matrix. As a result of this resorptive activity in contact

with the surface of calcified bone, osteoclasts create resorptive lacunae. Osteoclast function is regulated both by locally acting cytokines and by systemic hormones.

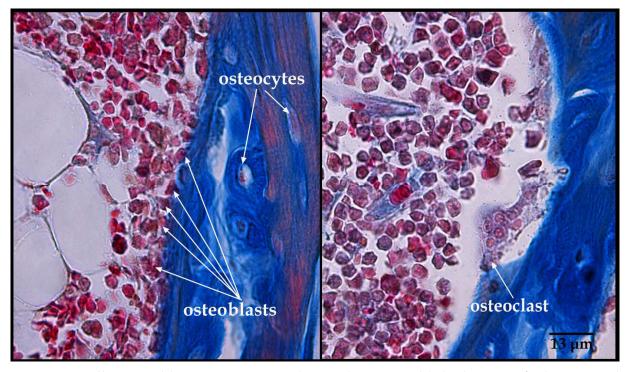


Fig. 1. Bone cells - osteoblasts, osteocytes and osteoclasts; unpublished image of Filipović et al.

In homeostatic equilibrium, bone resorption and formation are balanced. It appears that osteoclasts and osteoblasts closely collaborate in the remodeling process in what is called a "Basic Multicellular Unit", or BMU. This indicates that a coupling mechanism must exist between formation and resorption (Frost, 1964), although its nature is not known. Organization of the BMU in cortical and trabecular bone differs. Between 2% and 5% of cortical bone is remodeled each year. The remodeling process in trabecular bone is mainly a surface event. Due to the much larger surface to volume ratio, it is more actively remodeled than cortical bone, with remodeling rates that can be up to 10 times higher (Lee & Einhorn, 2001).

The remodeling cycle consists of three consecutive phases: resorption, reversal and formation. Resorption begins with the migration of partially differentiated preosteoclasts, which form multinucleated osteoclasts on the bone surface. During the reversal phase, mononuclear cells prepare the resorption lacunae for bone formation and provide signals for osteoblast differentiation and migration (Eriksen et al., 1990). Bone formation starts with activation of preosteoblasts to differentiate into osteoblasts. They secrete bone-matrix proteins to form the organic matrix, which is later mineralized. During this period, osteoblasts completely replace the resorbed bone by new tissue. After this phase, the surface is covered with flattened lining cells and a prolonged resting period ensues until a new remodeling cycle is initiated. Duration of the resorption phase is about 2 weeks, the reversal phase lasts for up to 4 or 5 weeks, while the formation phase can continue for 4 months.

At each remodeling site, bone resorption is coupled with bone formation, locally released growth factors and cytokines acting as mediators of this process (Canalis et al., 1988; Mundy, 1995). The decrease of bone mass, which may be due to different causes, is a

consequence of an imbalance between the amount of mineral and matrix removed and that subsequently incorporated into each resorption cavity (Kanis et al., 1990).

3. Phytoestrogens in bone protection

Phytoestrogens are structurally and functionally similar to estrogens and their estrogenic activity may occur through ERs. There are three main classes of phytoestrogens: isoflavonoids, coumestans and lignans (Fig. 2). Due to their estrogenic and anti-estrogenic activity, they are termed - natural selective ER modulators (SERMs). Therefore, soybean isoflavones have received great attention as alternatives to HRT for the prevention of postmenopausal osteoporosis. Genistein and daidzein, the main isoflavones in soybean, may protect against osteoporosis, because they can affect both types of bone cells.

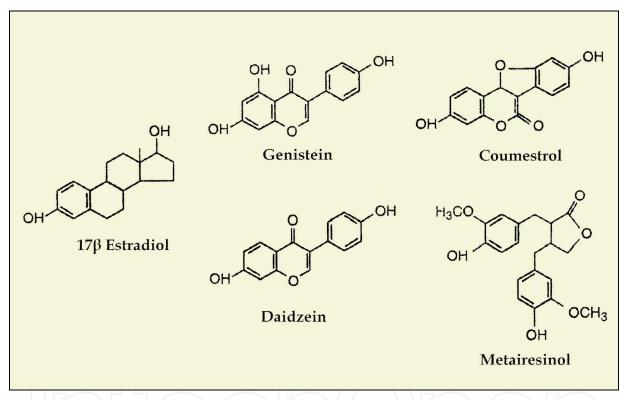


Fig. 2. Structure of 17β estradiol, isoflavones (genistein and daidzein), coumestan (coumestrol) and lignans (metairesinol); Filipović et al.

Isoflavones can stimulate the proliferation and differentiation of osteoblasts. Thus, the presence of genistein or daidzein led to a significant increase in protein synthesis, alkaline phosphatase activity, and DNA content in cultures of osteoblastic MC3T3-E1 cells (Sugimoto & Yamaguchi, 2000, 2000a; Yamaguchi & Sugimoto, 2000).

In addition to a stimulating effect on bone formation, these plant compounds may also suppress osteoclastic bone resorption in vitro. Thus, genistein was found to induce apoptosis of osteoclasts isolated from rat femoral tissues. Daidzein also decreased the number of these bone resorbing cells in rats (Gao & Yamaguchi, 1999) and their development in cultures of porcine bone marrow (Rassi et al., 2002). Osteoclast activity is regulated by phosphorylation of cell membrane constituents, involving tyrosine kinases. As a naturally tyrosine kinase inhibitor, genistein was found to suppress avian osteoclastic activity through inhibition of tyrosine kinase (Blair et al., 1996). Genistein also caused a significant increase in tyrosine phosphatase activity, which is a negative regulator of osteoclastogenesis and osteoclast-resorbing activity in mutant mice (Aoki et al., 1999; Gao &Yamaguchi, 2000) (Fig 3).

While investigations in vitro give clues about the effects of isoflavones on individual bone cells, studies in vivo provide knowledge about their influence in intact systems. Aged gonadectomized female and male rodents are suitable animal models for studying osteoporosis (Comelekoglu et al., 2007; Filipović et al., 2007; Pantelić et al., 2010; Vanderschueren et al., 1992.) Using them it has been demonstrated that isoflavones can prevent bone loss in female rats and mice after ovariectomy (Ovx) (Blum et al., 2003; Erlandsson et al., 2005; Fonseca & Ward, 2004; Ishimi et al., 1999; Lee et al., 2004; Om & Shim, 2007; Ren et al., 2007; Wu et al., 2004). The bone-preventing effects of isoflavones were also confirmed in male orchidectomized (Orx) rats and mice (Filipović et al., 2010; Ishimi et al., 2002; Khalil et al., 2005; Soung et al., 2006; Wu et al., 2003). On the contrary, some studies showed that isoflavones had minimal or no effects on bone loss in animal models (Bahr et al., 2005; Nakai et al., 2005; Picherit et al., 2001). Moreover, in the monkey, a nonhuman primate, dietary isoflavones do not effectively prevent ovariectomy-induced bone loss (Register et al., 2003). However, others suggested that soy phytoestrogens were protective against loss of bone volume (Ham et al., 2004).

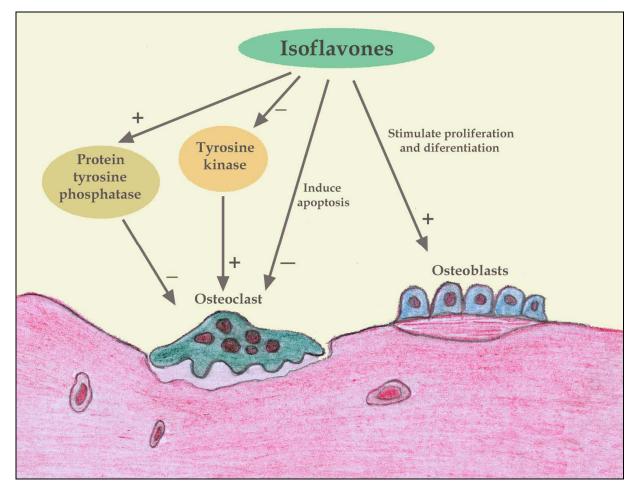


Fig. 3. Influence of isoflavones on bone cells; Filipović et al.

During recent years, numerous human studies have evaluated the effect of soy proteincontaining isoflavones or pure isoflavones on bone mass. However, the results of these observational and dietary interventional investigations have been variable and conflicting. In general, isoflavone supplementation studies indicate a beneficial effect on bone mass (Huang et al., 2006; Lydeking-Olsen et al., 2004; Newton et al., 2006), no effect (Anderson et al., 2002; Arjmandi et al., 2005; Brink et al., 2008; Wu et al., 2006) or a possible negative effect in terms of increased circulating concentrations of biochemical markers associated with bone resorption (Geppert et al., 2004; Wanger et al., 2000).

The large heterogeneity of these results may be due to study design, differences regarding hormonal status of the subjects, together with the duration, type and dose of isoflavone supplementation. In addition, bone sparing benefits may depend on the extent of conversion of isoflavones to metabolites. Thus, equol binds with greater affinity to ERs than daidzein from which it is derived (Setchell et al., 2002). Equol production is dependent on the intestinal microflora and there are large interindividual differences in this metabolism. Some people produce more equol than others. Also, production of this metabolite may at least partially explain why the beneficial effects of isoflavones observed in laboratory rodents, which consistently produce high levels of equol, have not been easily recapitulated in humans, where this is not the case. Generally, the relative importance of phytoestrogens in human health must be resolved and longer-term studies are needed to determine their effects on human bone tissue.

4. Phytoestrogens – Mechanisms of action in bone

Although the mechanisms by which soy phytoestrogens may alter bone remodeling are still not completely known, Atmaca et al. (2008) state that they act on both osteoblasts and osteoclasts through genomic and nongenomic pathways.

Due to their low molecular weight these plant compounds can pass through cell membranes and interact with receptors and enzymes (Adlercreutz et al., 1998). Phytoestrogens possess estrogenic activity and act as natural SERMs. This suggests that their effect on bone can be achieved by binding to ERs. Both α and β subtypes of ERs have been identified in bone (Arts et al., 1997; Onoe et al., 1997). The protective effect of phytoestrogens is probably achieved mainly through binding to ER- β , the expression of which is increased during bone mineralization (Arts et al., 1997; Kuiper et al., 1998). In addition to ERs, phytoestrogens can bind to androgenic receptors and act as phytoandrogens (Chen & Chang, 2007).

Both genistein and daidzein stimulate osteoblast proliferation, differentiation and activation by an ER-dependent mechanism (De Wilde et al., 2004; Pan et al., 2005). These isoflavones regulated the synthesis of core binding factor-1 (Cbfa-1) and bone morphogenic protein-2 (BMP-2), which is involved in the differentiation of osteoblasts (De Wilde et al., 2004; Jia et al., 2003; Pan et al., 2005). Genistein and daidzein activate peroxisome proliferator activator receptors (PPARs). The balance between PPAR and ER activation may govern the balance between adipogenesis and osteoblastogenesis (Dang et al., 2003, 2004).

Osteoclasts express the receptor activator of nuclear factor kappa B (RANK) (Hsu et al., 1999), while the receptor activator of nuclear factor kappa B ligand (RANK-L) and osteoprotegerin (OPG) is expressed by osteoblasts (Udagawa et al., 1999). Binding of RANKL to RANK stimulates osteoclastogenesis, whereas binding of RANK-L to OPG prevents RANK-L – RANK binding and indirectly inhibits osteoclastogenesis (Fuller et al., 1998; Theoleyre et al., 2004). The relative levels this triad of proteins are important for

738

controlling osteoclastogenesis. It was shown that isoflavones may increase the activity of osteoblasts by stimulating the secretion of OPG and RANK-L (De Wilde et al., 2004; Yamagishi et al., 2001) (Fig. 4).

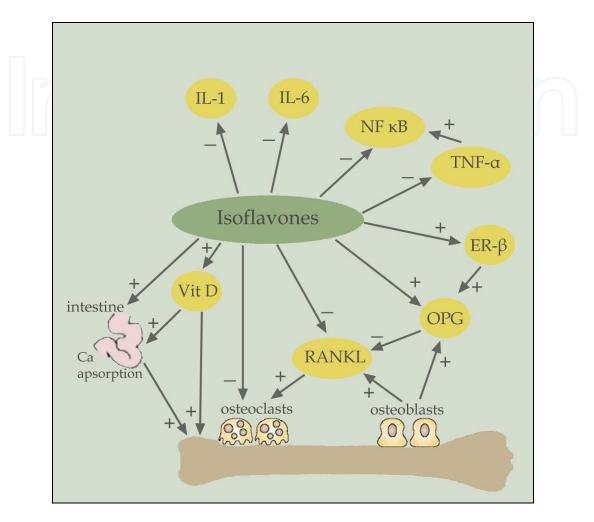


Fig. 4. Mechanisms of isoflavone action in bone; Filipović et al.

Proinflammatory cytokines, such as interleukin (IL)-1, IL-6, and tumor necrosis factor α (TNF- α), stimulate osteoclastogenesis and bone resorption. These effects can be achieved by both RANK-L dependent and RANK-L-independent mechanisms (Collin-Osbody et al., 2001; Katagiri et al., 2002). Isoflavones have been shown to inhibit IL-6 synthesis by MC3T3-E1/4 osteoblast-like cells in vitro (Chen et al., 2003; Suh et al., 2003) and to reduce serum IL-1 β and TNF- α concentrations in Ovx rats (Li, 2003). Also, a soy supplemented diet may inhibit serum concentrations of proinflammatory cytokines in postmenopausal women (Huang et al., 2005). In addition to osteoclastogenesis, isoflavones appear to influence osteoclast activity through inhibition of inward rectifier K+channels in osteoclasts. This leads to membrane depolarization, intracellular influx of Ca2+ and inhibition of bone resorption (Okamoto et al., 2001). One beneficial effect of isoflavones on bone is increased intestinal calcium absorption (Fig. 4). However, it is not known whether the mechanism(s) by which isoflavones influence calcium absorption include interactions with intestinal ER and/or vitamin D receptor-mediated calcium transport or not (Arjmandi et al., 2002).

Nongenomic effects do not involve ERs. These effects of phytoestrogens include inhibition of tyrosine kinase which directly modulate osteoclastic acid secretion (Blair et al., 1996; Williams et al., 1998) or topoisomerase I and II, which helps to regulate cell differentiation and the cell replication cycle (Okura et al., 1998; Yamagishi et al., 2001).

5. Indirect effects of phytoestrogens on bone – The role of calcitonin, parathyroid and thyroid hormones

5.1 Effects of phytoestrogens on thyroid C cells and calcitonin production

Thyroid C cells are dispersed neuroendocrine cells that produce many bioregulatory peptides, among which CT is considered the most important. This calcium regulating hormone lowers plasma calcium concentration by inhibiting osteoclast activity. In addition to sex steroids, a voluminous literature has accumulated for therapeutic use of CT in treating osteoporosis. Thus, C cells may also be very important in the pathogenesis of osteoporosis.

The C cells are mostly located in the middle of the thyroid lobes and appear in clusters or as solitary cells between follicular cells and the capillary wall. They have a round, elliptical or polygonal shape and never face the follicular lumen. The nucleus is located in the center of the cell. The most salient ultrastructural feature of C cells is the numerous round secretory granules that fill extensive areas of the cytoplasm. The Golgi complex and endoplasmic reticulum are well developed. There is a moderate number of mitochondria, which are mostly round to elongate in shape and not uniformly distributed. Lysosomes are large and contain acid phosphatase and other lysosomal enzymes (Fig. 5).

CT suppresses the number and motility of osteoclasts (Gao & Yamaguchi, 1999; Zaidi et al., 1990) and induces a change in their contractile elements (Hunter et al., 1989). Also, CT increases osteoblast proliferation by acting on components of the insulin-like growth factor system (Farley et al., 2000) and enhancing alkaline phosphatase activity, which is associated with increased synthesis and deposition of bone matrix collagen (Farley et al., 1988, 1992; Ito et al., 1987). The action of CT bone formation is at least in part, mediated via CT receptors located on osteoblasts, through the cAMP second messenger system (Farley et al., 1992; Villa et al., 2003).

It was shown that gonadal hormone deficiency affects thyroid C cell activity. Thus, synthesis of CT and its release from rat C cells were decreased after Ovx due to lack of estrogens (Filipović et al., 2002, 2003; Sakai et al. 2000). Also, the decline in testosterone level induced by Orx altered thyroid C cell structure and reduced the synthesis and release of CT (Filipović et al., 2007; Lu et al., 2000). The same effects were noticed after Orx or the natural menopause in women (Isaia et al. 1989). On the other hand, estrogen treatment was found to have a stimulatory effect on CT secretory activity of C cells in Ovx rats (Grauer et al., 1993; Filipović et al., 2003), Orx rats (Filipović et al., 2010a) and women (Isaia et al., 1992). In addition to estrogen, chronic calcium administration after Ovx increased the release of CT from C cells without affecting CT synthesis, suggesting that estrogen plays an important role in CT synthesis (Filipović et al., 2005). On the other hand, CT administration, which may be useful for treatment of osteoporosis, negatively affected rat thyroid C cells by a negative feedback mechanism (Sekulić et al., 2005).

Among the few studies concerning the potential effects of phytoestrogens on CT production, the influence of ipriflavone, a derivative of isoflavone, on CT synthesis and secretion was

740

investigated. Administration of ipriflavone to intact rats had a gender-related effect on serum CT, which increased in females, but no significant change was seen in male rats (Watanabe et al., 1992). With regard to the inhibitory effect of testosterone on the synthesis of some enzymes it is possible that testosterone inhibited ipriflavone-stimulated CT synthesis (Weiner & Dias, 1990).

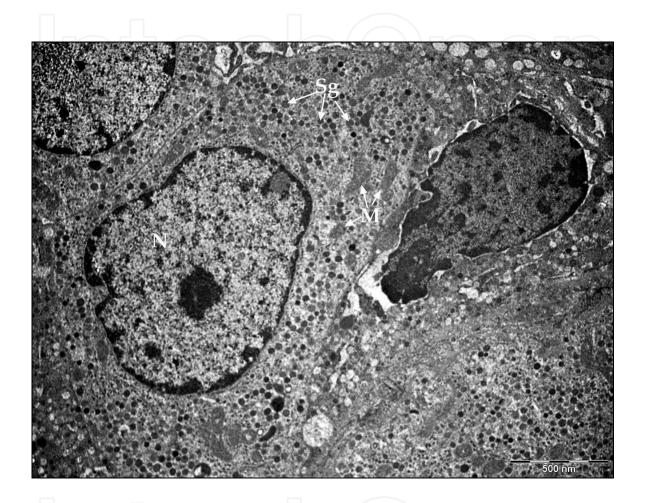


Fig. 5. Ultrastructure of a thyroid C cell; nucleus (N), mitochondria (M), secretory granules (Sg); unpublished image of Filipović et al.

Recently the first experimental data suggesting that daidzein affects thyroid C cells and stimulates CT secretory activity in Orx middle-aged rats were presented (Filipović et al., 2010). The androgen deficiency after Orx strongly affected thyroid C cell structure and reduced the synthesis and release of CT. Daidzein treatment decreased immunoreactivity for CT, significantly increased C cell volume (Fig. 6) and slightly raised serum CT concentration.

Daidzein administration also decreased bone turnover, prevented loss of cancellous bone and the plate-like structure was recovered after trabecular bone destruction caused by Orx (Fig. 7). Based on these results, the authors suggested that, besides direct action on the skeleton, daidzein may affect bone structure indirectly through enhancement of thyroid C cell activity (Filipović et al., 2010).

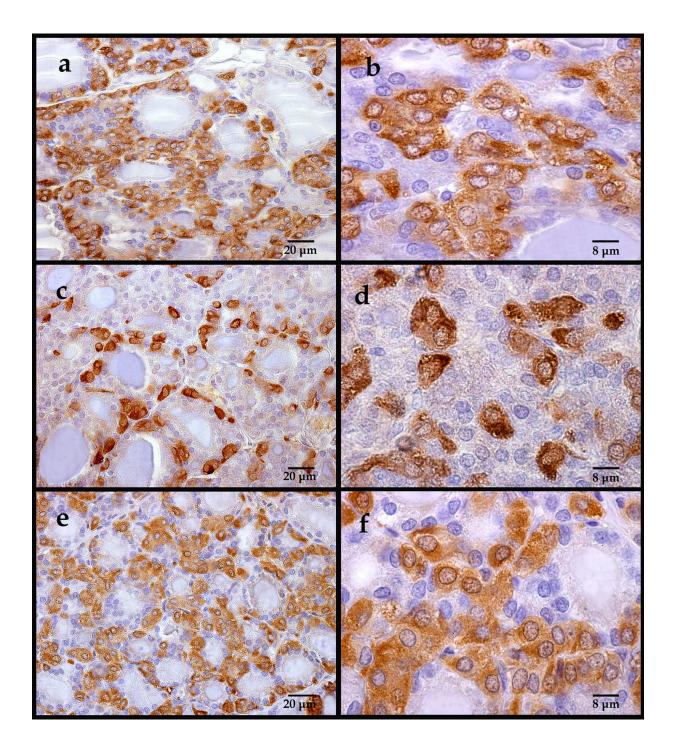


Fig. 6. Calcitonin producing thyroid C cells in control (a, b), orchidectomized (c, d) and orchidectomized rats treated with daidzein (e, f); immuno-staining for calcitonin; unpublished image of Filipović et al.

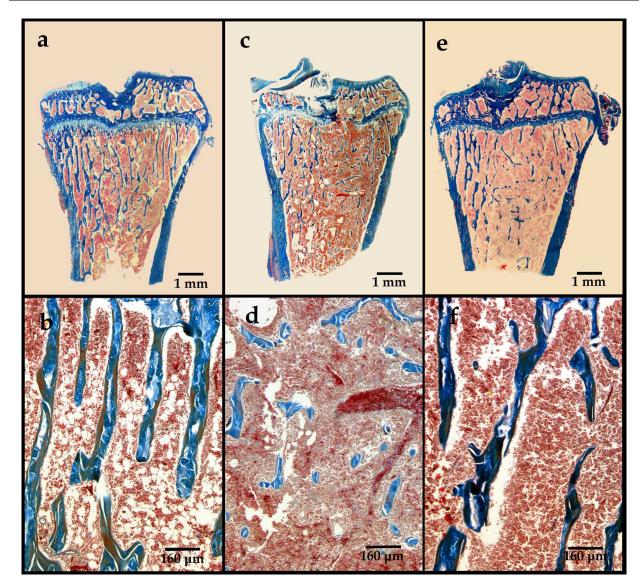


Fig. 7. Trabecular microarchitecture of the proximal tibial metaphysis in control (a, b), orchidectomized (c, d) and orchidectomized rats treated with daidzein (e, f); azan staining method; unpublished image of Filipović et al.

5.2 Effects of phytoestrogens on parathyroid hormone production

Parathyroid glands are constituted of chief, clear and oxyphilous cells. The chief cells synthesize and secrete PTH and are arranged in rather dense cords or nests around abundant capillaries. These cells are oval or polygonal in shape. The nucleus is irregularly shaped, with a few spots of chromatin located in the margin, and the nuclear membrane is infolded. The plasma membrane shows interdigitations. Mitochondria are dispersed throughout the cytoplasm. The cisternae of the rough-surfaced endoplasmic reticulum are arranged in parallel arrays or randomly distributed in the cytoplasm. The Golgi complexes are well developed. Storage granules are filled with finely particulate electron-dense material (Fig. 8).

PTH plays an important role in calcium homeostasis and has a critical role in bone turnover. It antagonizes CT produced by thyroid C cells and acts directly on bone and kidney to increase Ca influx into the blood circulation. This hormone increases the tubular re-

absorption of calcium and induces increased conversion of 25(OH)-D to 1,25(OH)2-D, which enhances intestinal calcium absorption and increases skeletal calcium mobilization.

PTH has a biphasic effect on bone, as it stimulates bone formation when given intermittently, whereas continuous infusion reduces bone mass (Kim et al., 2003). Treatment with PTH significantly increases ALP activity, which suggests that this hormone modulates SaOS-2 osteoblastic cell differentiation and has an anabolic effect on bone. However, increases in RANKL mRNA and decreased OPG mRNA expression in SaOS-2 cells due to PTH indicates induction of bone resorption (Chen & Wong, 2006).

Elevated PTH secretion contributes to the greater bone resorption in osteoporosis which is related to estrogen deficiency. Estrogen therapy prevented the increase in PTH levels associated with the menopause (Khosla et al., 1997). Similarly, phytoestrogens behave as estrogen and may prevent the bone loss caused by estrogen deficiency in female animals and women through reduction of PTH levels. It was shown that phytoestrogens from medical plants can lower serum PTH levels in aged menopausal monkeys (Trisomboon et al. 2004). Also, postmenopausal women with habitually high intakes of dietary isoflavones had significantly lower levels of serum PTH and higher BMD (Mei et al., 2001). These plant compounds bind to ERs in the kidney, gastrointestinal tract and bone and improve calcium absorption resulting in a secondary decrease in the PTH level. Moreover, phytoestrogens may directly reduce PTH secretion from the parathyroid gland (Wong et al., 2002).

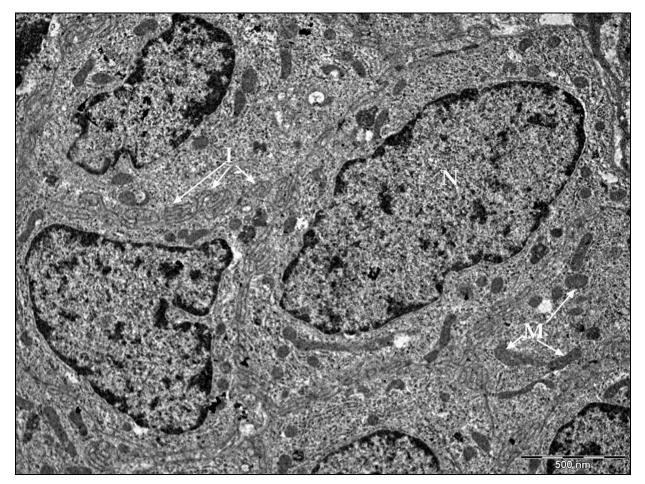


Fig. 8. Ultrastructure of parathyroid chief cells; nucleus (N), mitochondria (M), interdigitations of the plasma membrane (I); unpublished image of Pantelić et al.

www.intechopen.com

744

Mimicking the effect of estrogen, phytoestrogens can modulate the action of PTH on bone. Thus, one study in vitro showed that pre-treatment of SaOS-2 osteoblastic cells with genistein enhanced PTH-induced ALP activity and attenuated PTH up regulation of RANKL mRNA expression and PTH down regulation of OPG mRNA expression (Chen & Wong, 2006).

5.3 Effects of phytoestrogens on thyroid glands and thyroid hormones production

Hypothalamic-pituitary-thyroid axis (HPT) plays a key role in skeletal development, attainment of peak bone mass and regulation of adult bone turnover (Gogakos et al., 2010; Roef et al., 2011). Additionally, thyroid disorders are associated with alterations in bone metabolism (Lakatos, 2003).

Soy-food, soy-based infant formula, as well as dietary supplements containing purified soybean isoflavones, genistein and daidzein, are increasingly consumed in typical "Western" diet in the recent years. Commonly cited reasons for using soy infant formula are to feed infants who are allergic to dairy products or are intolerant of lactose, galactose, or cow-milk protein (Tuohy, 2003). In elderly, reason is potential health benefit of soybean isoflavones in protection of age-related diseases, including osteoporosis (Setchell, 1998).

Structurally, soybean isoflavones genistein and daidzein are polyphenolic compounds, similar to estradiol-17 β and bind with a weaker potency to both types of ERs, with higher affinity for ER β (Kuiper et al., 1998). Despite the numerous beneficial effects of soy isoflavones, epidemiological and experimental data also exist showing an adverse effect on human health, namely on reproductive and thyroid axis. The association between high soy isoflavones intake and goitrogenesis, as well as protective effect of adequate iodine intake, was reported both in humans (Chorazy et al.1995; Van Wyk et al., 1959) and in different animal models (Ikeda et al., 2000; Kimura et al., 1976; McCarrison, 1933).

Therefore, besides the direct beneficial effect of soybean phytoestrogens on bone tissue, isoflavones may also act indirectly, through endocrine disruption and interference with HPT axis. Most researchers who examined osteoprotective potential of isoflavones did not include in their research examining of the thyroid status. We will address that aspect in this subchapter.

5.3.1 Phytoestrogens, thyroid hormones and skeletal development

Normal thyroid function in childhood is essential for development of endochondral and intramembranous bone, for normal linear growth, as well as for establishing peak bone mass. Hypothyroidism in children causes growth arrest, delayed bone maturation, and epiphyseal dysgenesis, while T₄ replacement results in rapid catch-up growth (Basset & Williams, 2003). Exposure to soybean isoflavones during development may alter thyroid hormone concentrations and disturb feedback regulation of HPT axis, and these effects can be more serious than in the adults.

Soy infant formula is fed to infants as a replacement for human milk, or as an alternative to cow milk formula. Genistein is the predominant isoflavone found in soy infant formula (58-67%), followed by daidzein (29-34%) and glycitein (5-8%) and infants fed soy infant formula have higher daily intakes of genistein and other isoflavones than other populations (Patisaul & Jefferson, 2010). The question of whether or not soy infant formula is safe has been widely debated for more than a decade, and early epidemiological studies demonstrated that infants fed adapted soy formula without iodine supply were hypothyroid (Van Wyk et al., 1959). This effect was eliminated by supplementing commercial soy infant formulas with iodine, or by

switching to cow milk (Chorazy et al., 1995). Today, soy formula is regularly supplied with iodine and a more recent study demonstrated no significant changes in the serum level of bone alkaline phosphatase, osteocalcin, intact PTH, and the urinary levels of the markers of bone metabolism in children (mean age of 37 months) fed with soy formula (Giampietro et al., 2004). However, infants with congenital hypothyroidism fed with iodine supplemented diet still need higher doses of L-thyroxine (Jabbar et al., 1997). This finding is of particular importance, keeping in mind that the consequence of congenital and juvenile acquired hypothyroidism is retardation of skeletal development and that the effects of T₄ replacement (achievement of predicted adult height) strongly depend on the duration of untreated hypothyroidism (Rivkees et al., 1988).

Soybean isoflavones may functionally disrupt the thyroid hormone (TH) system by influencing different steps such as synthesis, transport, action and metabolism of TH. Genistein and daidzein inhibit the activity of thyroid peroxidase (TPO), the key enzyme in the synthesis of thyroid hormones, both in vitro and in vivo (Chang & Doerge, 2000; Divi et al., 1997; Doerge & Chang, 2002). Besides the inhibitory effects of isoflavones on TPO, iodine deficiency is important risk factor for thyroid dysfunction and goiter development, both in humans and in rats. An adequate iodine supply is a way to prevent goitrogenic effects of soy bean isoflavones, especially in the high-risk group of patients with congenital hypothyroidism. Besides the serum concentrations of TH, biological activity of T_3 on bone tissue is determined by the membrane transporters of TH, local expression and activity of deiodinase enzymes and receptors for TSH and TH. Polymorphisms in above mentioned genes are associated with important chronic skeletal diseases, including osteoporosis and osteoarthritis (Andersen et al., 2002, 2003; Peeters et al., 2006).

Entry of T_3 and T_4 into target cells is determined by the active uptake of free hormones by specific cell membrane transporters: monocarboxylate transporter-8 (MCT8), MCT10 and organic acid transporter protein-1c1 (OATP1c1) (van der Deure et al., 2010). MCT8 is expressed in growth plate chondrocytes, bone forming osteoblasts and bone resorbing osteoclasts at all stages of cell differentiation, and its expression is regulated by thyroid status (Capelo et al., 2009), although its functional importance is still unclear. It seems that OATP1c1 is not expressed in the mouse skeleton (Capelo et al., 2009), but there are still no data regarding expression of MCT10. Tyrosine kinase inhibitors sunitinib and imatinib inhibit MCT8 – mediated iodothyroinine transport (Schweizer et al., 2010), but there are still no data regarding possible effects of genistein, which is a potent thyrosine kinase inhibitor as well, on cellular transport of TH.

Deiodinase (Dio) enzymes determine the intracellular levels of bioactive T3 and thus cellspecific gene expression. Expression of deiodinases is tissue specific: Dio 1 enzyme is not expressed in bone, while Dio 2 plays an important role in local regulation of thyroid hormone signaling during fetal bone development. In the adult skeleton Dio 2 activity is restricted to osteoblasts (Williams et al., 2008). Dio 2 expression and activity are inhibited by high concentrations of substrate (T₄) and thus are maximal in hypothyroidism and suppressed in thyrotoxicosis. Locally regulated activity of Dio 2 in osteoblasts maintains intra-cellular T₃ concentrations constant over the euthyroid range and preserves optimal bone mineralization. Inactivating deiodinase type 3 (Dio 3) is expressed in the skeleton, although the highest levels of enzyme activity occur in growth plate chondrocytes prior to weaning (Yen, 2001). Genistein inhibit both Dio 1 and Dio 2 activity in vitro (Mori et al., 1996), but the physiological importance of this mechanism is still unclear.

746

Based on analyses of rare monogenic diseases and the results of animal studies, it was proposed that T_3 play a key role in bone development, while TSH is not required for normal skeletal development (Bassett et al., 2008). T_3 enters the nucleus and binds to its nuclear receptors (TR). There are three functional TRs: TRa1, TR β 1 and TR β 2, encoded by the THRA and THRB genes. These receptors act as hormone inducible transcription factors that regulate expression of T_3 -responsive target genes (Yen, 2001). Both TRa1 and TR β 1 isoforms are expressed in bone and TRa1 levels are at least 10-fold greater than TR β 1. These findings support the opinion that TRa1 is the principal mediator of T_3 action in bone (Bassett & Williams, 2009; O'Shea et al., 2003).

In vitro experiments demonstrated that effects of T_3 in osteoblastic cell lines and primary osteoblast cultures depend on species, cell type, anatomic origin, differentiation phase and duration of the treatment. T_3 was reported to increase expression of osteocalcin, osteopontin, type I collagen, alkaline phosphatase, IGF-I and its regulatory binding proteins IGF1BP-2 and -4 (Milne et al., 2001; Pereira et al., 1999; Varga et al., 2004). Therefore, T_3 may exert its stimulatory effect on osteoblasts via complex pathways involving many growth factors and cytokines.

5.3.2 Phytoestrogens, thyroid hormones and osteoporosis prevention

Similar to osteoporosis, thyroid diseases are much more common in elderly women than in men and is associated with significant morbidity if left untreated (Schindler 2003; Suchartwatnachai et al., 2002). Still, this fact does not imply a causal relationship between the two diseases and many patients may independently develop both. Hypothyroidism occurs in 10% of females and 2% of males in patients older than 60 years. The prevalence of hyperthyroidism in the elderly is approximately 2% (Maugeri et al., 1996), though other authors reported that 10 to 15% of elderly patients were hyperthyroid (Kennedy & Caro, 1996). Thyrotoxicosis increase risk in developing secondary osteoporosis (Amashukeli et al., 2010; Lakatos, 2003).

Thyroid hormones play a significant role in maintaining adult bone homeostasis. Results of clinical and experimental studies are consistent and demonstrate that hypothyroid state slows down bone turnover and affect overall gain in bone mass and mineralization. By contrast, bone resorption and formation are accelerated in hyperthyroidism, while the remodeling cycle is shortened (Davies et al., 2005). Increased bone turnover and osteoporosis in thyrotoxicosis are attributed to the thyroid hormone excess and are not a consequence of deficient TSH receptor (TSHR) signaling. However, TSH may play a direct role in regulation of bone turnover, since TSH receptor was identified in osteoblasts. The experiment with ovariectomized rats, which were treated with low doses of TSH (insufficient to alter serum T_{3} , T_{4} or TSH levels), demonstrated that TSH treatment prevented bone loss and increased bone mass (Sampath et al., 2007; Sun et al., 2008). Although the TSHR is expressed in osteoblasts, current data from in vitro studies are contradictory and suggest that TSH may enhance, inhibit or have no effect on osteoblast differentiation and function (Bassett et al., 2008).

Prevention and treatment of osteoporosis involve Ca and vitamin D supplementation, as well as different drug therapy approaches, which include bisphosphonate, salmon CT and estrogen or androgen replacement therapy for menopausal women and andropausal men, respectively. In addition, in recent years, numerous discussions on safety and benefit of synthetic steroids (both estrogens and androgens) favor the trend towards consumption of "green" natural "phytosteroids" or "phyto-selective modulator of ERs". That is why nutritional supplements and concentrated extracts containing purified soybean phyto-SERMs genistein and daidzein are increasingly used as alternative therapy for osteoporosis and other age-related diseases in both sexes (Ramos, 2007; Setchell, 1998; Tham et al., 1998). However, all these treatments may affect thyroid function as well.

Not so many researchers have tried to link effects of supplementation or drug treatment on bone metabolism with modulation of thyroid hormone levels. Rodents are considered useful models for thyroid studies, even though significant differences between rodent and human thyroid physiology have been reported (Choksi et al., 2003; Poirier et al., 1999). Rat thyrocytes are characterized by abundant granular endoplasmic reticulum, well developed Golgi, prominent lysosomes, luminal (apical) microvilli, small mitochondria, and round nuclei with homogeneous chromatin (Fig. 9).

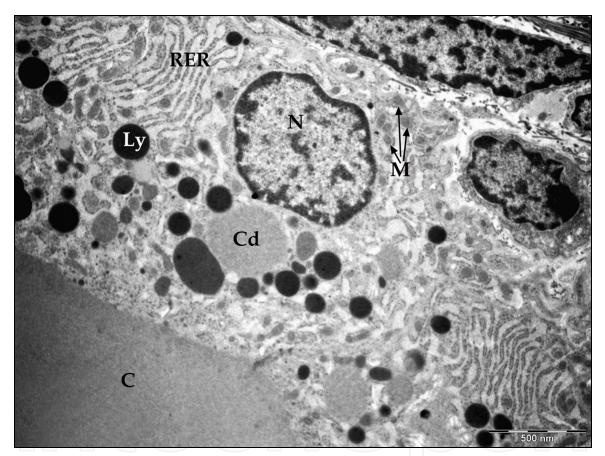


Fig. 9. Ultrastructure of thyroid follicular cell; nucleus (N), mitochondria (M), rough endoplasmatic reticulum (RER), lysosomes (Ly), colloidal droplets (Cd), colloid (C); unpublished image of Šošić-Jurjević et al.

In our laboratory we demonstrated that chronic Ca administration to middle-aged female rats significantly decreased the volume density of the thyroid follicular epithelium, epithelium's height and the index of activation rate, which are morphometric parameters of TH synthetic and secretory potential of thyrocytes (Šošić -Jurjević et al., 2002). Consistent with histomorphometric changes, reduced serum levels of total T_4 and T_3 were detected (Šošić -Jurjević et al., 2006). At the same time, we determined significant decrease of serum osteocalcin and urinary Ca, as biochemical parameters of reduced bone turnover after Ca treatment (unpublished data). In vitro studies with FTRL-5 cells demonstrated that Ca did not affect the morphology of these cells, but when administered together with TSH, it acted directly, by reducing the thyrotropin stimulatory effect (Gaberscek et al., 1998). Isoform VI of adenilyl cyclase, the enzyme crucial for TSH-induced activation of thyroid follicular cells, was found negatively modulated by Ca in human and dog thyroids (Vanvooren et al., 2000). Doses of Ca were chosen to mimic human exposure to high doses of Ca in treatment of osteoporosis. We can speculate that slowing down of thyroid hormone synthesis may be an indirect mechanism, which lead to decreased bone turnover detected after Ca treatment under our experimental conditions.

Sex steroids, estrogen and testosterone, play an important role in bone physiology and pathology. Endogenous estrogens are regularly produced in bone via aromatase enzyme activity, and exert their effects through ER, which are also detected in male bones (Carani et al., 1997; Grumbach & Auchus, 1999; Korach, 1994). Bone cells are sensitive to both estrogens and androgens, and aromatase inhibition causes similar degree of osteoporosis in male animals as orchidectomy (Vanderschueren et al., 1998).

There is a close relationship between sex steroids and thyroid function. Epidemiological studies suggest that the use of estrogens may contribute to the pathogenesis of thyroid tumors (Ron et al., 1987). Experimental studies on rodents demonstrated numerous sexrelated differences in thyroid function and, in general, adult male rodents have higher levels of TSH than females associated with lower T_4 and higher plasma levels of T_3 (Capen, 1997). The results related to treatment effects of sex steroids on different set points of thyroid function are inconsistent and depend on experimental conditions: type of experimental animal, animal's age and applied dose (Chen & Wallfish, 1978; Henderson et al., 1982; Sekulić et al., 2007). Our previous results demonstrated an inhibitory effect of pharmacologic doses of estradiol (previously used in human studies for treatment of osteoporosis) on thyroid follicular cells in ovariectomized young adult and ovarium-intact young and middle-aged rats, (Sekulić et al., 2006; Šošić -Jurjević et al., 2005, 2006a), as well as after treatments of orchidectomized 16-month-old rat males with 10 times lesser dose of estradiol dipropionate (Sekulić et al., 2010). We choose the dose of estradiol in the experiment which was previously reported to prevent bone loss in males (Fitts et al., 2001; Vanderput et al., 2001). Consistent with literature data, we also detected decreased serum osteocalcin levels, accompanied by decreased urinary Ca concentration in Orx rats treated with EDP (unpublished data). Contrary to effects of estradiol, testosterone treatment of castrated middle-aged males moderately increased serum TSH and total T4 levels (Sekulić et al., 2010), but similarly to estradiol treatment, decreased both serum osteocalcin levels and urinary Ca concentration (unpublished data). Therefore, it seems that the direct effect of sex steroids on bone tissue is more relevant for the net result of replacement therapy on bone protection then the indirect effect, mediated through modulation of thyroid function.

Direct negative effect of isoflavones on thyroid hormone synthesis, by significant blocking of TPO activity (more than 60%), has been well described. Genistein and daidzein were demonstrated to block both TPO-catalyzed reactions: iodination of thyrosine residues of Tg, and T₄ formation by coupling reactions, but this effect was eliminated by iodine (Chang & Doerge, 2000; Divi et al., 1997; Doerge & Chang 2002). Despite significant inactivation of this enzyme, serum thyroid hormone levels were unaffected by isoflavone treatments in young adult rats of both sexes. The authors supposed that soy could cause goiter, but only in animals or humans consuming diets marginally adequate in iodine, or who were predisposed to develop goiter. Most other authors, who performed their studies on young adult animals of both sexes, also reported that soy or isoflavones alone, in the absence of

other goitrogenic stimulus, did not affect thyroid weights, histopathology and the serum levels of TSH and thyroid hormones (Chang & Doerge, 2000; Schmutzler et al., 2004). The thyroid function becomes impaired with aging in rodents, and the number of thyroid dysfunction increase in elderly population (Donda & Lemarchand-Béraud, 1989; Reymond et al., 1992). We were the first who demonstrated that therapeutic doses of both genistein and daidzein induce hypertrophy of Tg-immunopositive follicular epithelium and colloid depletion (Fig. 10), and reduce the level of serum thyroid hormones, accompanied by

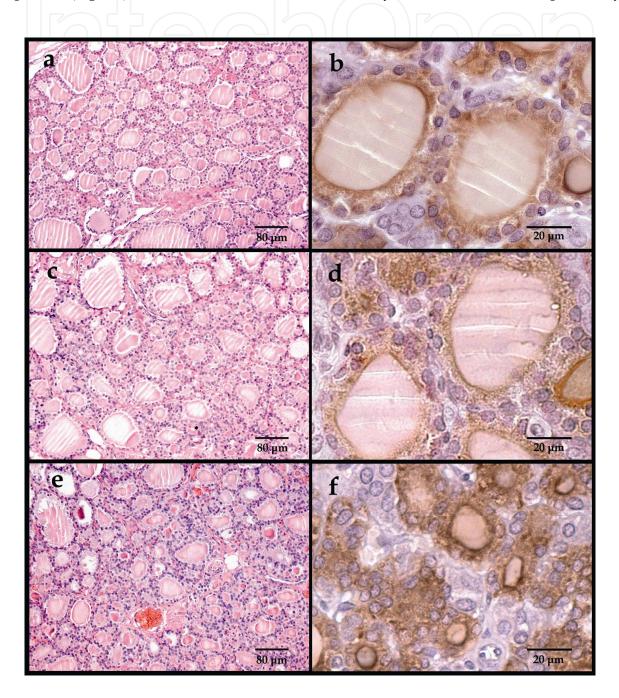


Fig. 10. Thyroid gland tissue of control (a, b), orchidectomized (c, d) and orchidectomized rats treated with daidzein (e, f); hematoxylin-eosin and immuno-staining for thyroglobulin; unpublished image of Šošić-Jurjević et al.

increased serum TSH, in orchidectomized (Orx) middle-aged male rats fed a iodinesufficient soy-free diet (Šošić -Jurjević et al., 2010). Our research team obtained that both genistein and daidzein increased bone mass following orchidectomy of middle-aged males (Filipovic et al., 2010 and unpublished data). Therefore, decreased serum level of TH might contribute to the detected increase in trabecular bone mass, and decrease in bone turnover in aged male orchidectomized rat model.

6. Conclusion

Phytoestrogens have the potential to maintain bone health. Owing to their properties, these plant-derived non-steroidal compounds have a potential beneficial role in delaying or preventing osteoporosis. Therefore, they have attracted much attention as alternatives to HRT. As SERM, phytoestrogens may generate a bone protective effect via stimulation of osteoblastic bone formation and inhibition of osteoclastic bone resorption. Proposed molecular mechanisms are based on their ER-mediated effects. In addition to direct action, phytoestrogens can affect bone structure indirectly, by stimulating or inhibiting the synthesis of certain hormones, i.e. through increased synthesis of CT from thyroid C cells, as well as reduction of PTH and thyroid hormone levels.

7. Acknowledgment

This work was supported by the Ministry of Education and Science of the Republic of Serbia, Grant No. 173009. The authors express their gratitude to the late Dr Dana Brunner for her guidance and contribution, to Mrs. Anna Nikolić and Mr. Kristijan Jurjević for assistance with English manuscript preparation.

8. References

- Abu, EO., Bord, S., Horner, A., Chatterjee, VK. & Compston, JE. (1997). The expression of thyroid hormone receptors in human bone. *Bone*, Vol. 21, pp. 137-142
- Adlercreutz, H. (1998). Evolution, nutrition, intestinal microflora, and prevention of cancer: a hypothesis. *Proc Soc Exp Biol Med*, Vol. 217, pp. 241–246
- Amashukeli, M., Giorgadze, E., Tsagareli, M., Nozadze, N. & Jeiranashvili, N. (2010). The impact of thyroid diseases on bone metabolism and fracture risk. *Georgian Med* News, Vol. 184-185, pp. 34-39
- Andersen, S., Bruun, NH., Pedersen, KM. & Laurberg, P. (2003). Biologic variation is important for interpretation of thyroid function tests. *Thyroid*, Vol. 13, pp. 1069-1078
- Andersen, S., Pedersen, KM., Bruun, NH. & Laurberg, P. (2002). Narrow individual variations in serum T(4) and T(3) in normal subjects: a clue to the understanding of subclinical thyroid disease. *J Clin Endocrinol Metab*, Vol. 87, pp. 1068-1072
- Anderson, JJ., Chen, X., Boass, A., Symons, M., Kohlmeier, M., Renner, JB. & Garner, SC. (2002). Soy isoflavones: no effects on bone mineral content and bone mineral density in healthy, menstruating young adult women after one year. J Am Coll Nutr, Vol. 21, pp. 388–393
- Aoki, K., Didomenico, E., Sims, NA., Mukhopadhyay, K., Neff, L., Houghton, A., Amling, M., Levy, JB., Horne, WC. & Baron, R. (1999). The tyrosine phosphatase SHP-1 is a

negative regulator of osteoclastogenesis and osteoclast resorbing activity: Increased resorption and osteopenia in mev/mev mutant mice. Bone, Vol. 25, pp. 261-267

- Arjmandi, BH., Khalil, DA. & Hollis, BW. (2002). Soy protein: its effects on intestinal calcium transport, serum vitamin D, and insulin-like growth factor-I in ovariectomized rats. *Calcif Tissue Int*, Vol. 70, pp. 483-487
- Arjmandi, BH., Lucas, EA., Khalil, DA., Devareddy, L., Smith, BJ., McDonald, J., Arquitt, AB., Payton, ME. & Mason, C. (2005). One year soy protein supplementation has positive effects on bone formation markers but not bone density in postmenopausal women. *Nutr J*, Vol. 4, pp. 8
- Arts, J., Kuiper, GG., Janssen, JM., Gustafsson, JA., Lowik, CW., Pols, HA. & van Leeuwen, JP. (1997). Differential expression of estrogen receptors alpha and beta mRNA during differentiation of human osteoblast SV-HFO cells. *Endocrinology*, Vol. 138, pp. 5067- 5070
- Atmaca, A., Kleerekoper, M., Bayraktar, M. & Kucuk, O. (2008). Soy isoflavones in the management of postmenopausal osteoporosis. *Menopause*, Vol. 15, pp. 748–757
- Bahr, JM., Nakai, M., Rivera, A., Walsh, J., Evans, GL., Lotinun, S., Turner, RT., Black, M. & Jeffery, EH. (2005). Dietary soy protein and isoflavones: minimal effects on bone and no effect on the reproductive tract of sexually mature ovariectomized Sprague-Dawley rats. *Menopause*, Vol. 12, pp. 165-173
- Bassett, JH. & Williams, GR. (2003). The molecular actions of thyroid hormone in bone. *Trends Endocrinol Metab*, Vol. 14, pp. 356-164
- Bassett, JH., Williams, AJ., Murphy, E., Boyde, A., Howell, PG., Swinhoe. R., Archanco, M., Flamant, F., Samarut, J., Costagliola, S., Vassart, G., Weiss, RE., Refetoff, S. & Williams, GR. (2008). A lack of thyroid hormones rather than excess thyrotropin causes abnormal skeletal development in hypothyroidism. *Mol Endocrinol*, Vol. 22, pp. 501-512
- Bassett, JH. & Williams, GR. (2009). The skeletal phenotypes of TRalpha and TRbeta mutant mice. *J Mol Endocrinol*, Vol. 42, pp. 269-282
- Bianco, P., Riminucci, M., Gronthos, S. & Robey, PG. (2001). Bone marrow stromal stem cells: nature, biology, and potential applications. *Stem Cells*, Vol. 19, pp. 180-192
- Blair, HC., Jordan, SE., Peterson, TG. & Barnes, S. (1996). Variable effects of tyrosine kinase inhibitors on avian osteoclastic activity and reduction of bone loss in ovariectomized rats. *J Cell Biochem*, Vol. 61, pp. 629-637
- Blum, SC., Heaton, SN., Bowman, BM., Hegsted, M. & Miller, SC. (2003). Dietary soy protein maintains some indices of bone mineral density and bone formation in ovariectomized rats. *J Nutr*, Vol. 133, pp. 1244-1249
- Branca, F. (2003). Dietary phyto-oestrogens and bone health. *Proc Nutr Soc*, Vol. 62, pp. 877– 887
- Brink, E., Coxam, V., Robins, S., Wahala, K., Cassidy, A. & Branca, F. (2008). Long-term consumption of isoflavone-enriched foods does not affect bone mineral density, bone metabolism, or hormonal status in early postmenopausal women: a randomized, double-blind, placebo controlled study. *Am J Clin Nutr*, Vol. 87, pp. 761–770
- Canalis, E., McCarthy, T. & Centrella, M. (1988). Growth factors and the regulation of bone remodeling. *J Clin Invest*, Vol. 81, pp. 277-281

- Canalis, E., Pash, J. & Varghese, S. (1993). Skeletal growth factors. *Crit Rev Eukaryot Gene Expr*, Vol. 3, pp. 155-166
- Canalis, E., Pash, J., Gabbitas, B., Rydziel, S. & Varghese, S. (1993a). Growth factors regulate the synthesis of insulin-like growth factor-I in bone cell cultures. *Endocrinology*, Vol. 133, pp. 33-38
- Capelo, LP., Beber, EH., Fonseca, TL. & Gouveia, CH. (2009). The monocarboxylate transporter 8 and L-type amino acid transporters 1 and 2 are expressed in mouse skeletons and in osteoblastic MC3T3-E1 cells. *Thyroid*, Vol. 19, pp. 171-178
- Capen, C. (1997). Mehanicistic data and risk assessment of selected toxic end points of the thyroid gland. *Toxicologic Pathology*, Vol. 25, pp. 39-48
- Carani, C., Qin, K., Simoni, M., Faustini-Fustini, M., Serpente, S., Boyd, J., Korach, KS. & Simpson, ER. (1997). Effect of testosterone and estradiol in a man with aromatase deficiency. N Engl J Med, Vol. 337, pp. 91-95
- Chang, HC. & Doerge, DR. (2000). Dietary genistein inactivates rat thyroid peroxidase in vivo without an apparent hypothyroid effect. *Toxicol Appl Pharmacol*, Vol. 168, pp. 244-252
- Chen, D., Zhao, M. & Mundy, GR. (2004). Bone morphogenetic proteins. *Growth Factors*, Vol. 22, 233-241
- Chen, HJ. & Walfish, PG. (1978). Effects of estradiol benzoate on thyroid-pituitary function in female rats. *Endocrinology*, Vol. 103, pp. 1023-1030
- Chen, JJ. & Chang, HC. (2007). By modulating androgen receptor coactivators, daidzein may act as a phytoandrogen. *Prostate*, Vol. 67, pp. 457-462
- Chen, WF. & Wong, MS. (2006). Genistein modulates the effects of parathyroid hormone in human osteoblastic SaOS-2 cells. *Br J Nutr*, Vol. 95, pp. 1039-1047
- Chen, XW., Garner, SC., Quarles, LD. & Anderson, JJB. (2003). Effects of genistein on expression cell of bone markers during MC3T3-E1 osteoblastic differentiation. J Nutr Biochem, Vol. 14, pp. 342–349
- Choksi, NY., Jahnke, GD., St Hilaire, C. & Shelby, M. (2003). Role of thyroid hormones in human and laboratory animal reproductive health. Birth Defects *Res B Dev Reprod Toxicol*, Vol. 68, pp. 479-491
- Chorazy, PA., Himelhoch, S., Hopwood, NJ., Greger, NG. & Postellon, DC. (1995). Persistent hypothyroidism in an infant receiving a soy formula: case report and review of the literature. *Pediatrics*, Vol. 96, pp. 148-150
- Collin-Osdoby, P., Rothe, L., Anderson, F., Nelson, M., Maloney, W. & Osdoby, P. (2001). Receptor activator of NF-kappa B and osteoprotegerin expression by human microvascular endothelial cells, regulation by inflammatory cytokines, and role in human osteoclastogenesis. *J Biol Chem*, Vol. 276, pp. 20659–20672
- Comelekoglu, U., Bagis, S., Yalin, S., Ogenler, O., Yildiz, A., Sahin, NO., Oguz, I. & Hatungil, R. (2007). Biomechanical evaluation in osteoporosis: ovariectomized rat model. *Clin Rheumatol*, Vol. 26, pp. 380-384
- Dang, ZC., Audinot, V., Papapoulos, SE., Boutin, JA. & Lowik, C. (2003). Peroxisome proliferator-activated receptor gamma (PPAR gamma) as a molecular target for the soy phytoestrogen genistein. J Biol Chem, Vol. 278, pp. 962–967

- Dang, ZC. & Lowik, C. (2004). The balance between concurrent activation of ERs and PPARs determines daidzein-induced osteogenesis and adipogenesis. J Bone Miner Res, Vol. 19, pp. 853–861
- Davies, J., Warwick, J., Totty, N., Philp, R., Helfrich, M. & Horton, M. (1989). The osteoclast functional antigen, implicated in the regulation of bone resorption, is biochemically related to the vitronectin receptor. *J Cell Biol*, Vol. 109, pp. 1817-1826
- Davies, TF, Ando, T., Lin, RY., Tomer, Y. & Latif, R. (2005). Thyrotropin receptor-associated diseases: from adenomata to Graves disease. *J Clin Invest*, Vol. 115, pp. 1972-1983
- Davison, S. & Davis, SR. (2003). Hormone replacement therapy: current controversies. *Clin Endocrinology*, Vol. 58, pp. 249–261
- DeWilde, A., Lieberherr, M., Colin, C. & Pointillart, A. (2004). A low dose of daidzein acts as an ER beta-selective agonist in trabecular osteoblasts of young female piglets. *J Cell Physiol*, Vol. 200, pp. 253–262
- Divi, RL., Chang, HC. & Doerge, DR. (1997). Anti-thyroid isoflavones from soybean: isolation, characterization, and mechanisms of action. *Biochem Pharmacol*, Vol. 54, pp. 1087-1096
- Doerge, DR. & Chang, HC. (2002). Inactivation of thyroid peroxidase by soy isoflavones, in vitro and in vivo. *J Chromatogr B Analyt Technol Biomed Life Sc*, Vol. 777, pp. 269-279
- Donda, A. & Lemarchand-Béraud, T. (1989). Aging alters the activity of 5'-deiodinase in the adenohypophysis, thyroid gland, and liver of the male rat. *Endocrinology*. Vol. 124, pp. 1305-1309
- Eriksen, EF., Hodgson, SF., Eastell, R., Cedel, SL., O'Fallon, WM. & Riggs, BL. (1990). Cancellous bone remodeling in type I (postmenopausal) osteoporosis: quantitative assessment of rates of formation, resorption, and bone loss at tissue and cellular levels. J Bone Miner Res, Vol. 5, pp. 311-319
- Erlandsson, MC., Islander, U., Moverare, S., Ohlsson, C. & Carlsten, H. (2005). Estrogenic agonism and antagonism of the soy isoflavone genistein in uterus, bone and lymphopoiesis in mice. *APMIS*, Vol. 113, pp. 317-323
- Farley, JR., Tarbaux, NM., Hall, SL., Linkhart, TA. & Baylink, DJ. (1988). The anti-boneresorptive agent calcitonin also acts in vitro to directly increase bone formation and bone cell proliferation. *Endocrinology*, Vol. 123, pp. 159–167
- Farley, JR., Hall, SL., Herring, S. & Tarbaux, NM. (1992). Two biochemical indices of mouse bone formation are increased, in vivo, in response to calcitonin. *Calcif Tissue Int*, Vol. 50, pp. 67–73
- Farley, J., Dimai, HP., Stilt-Coffing, B., Farley, P., Pham, T. & Mohan, S. (2000). Calcitonin increases the concentration of insulin-like growth factors in serum-free cultures of human osteoblast-line cells. *Calcif Tissue Int*, Vol. 67, pp. 247–254
- Filipović, B., Šošić -Jurjević, B., Manojlović-Stojanoski, M., Nestorović, N., Milošević, V. & Sekulić, M. (2002). The effect of ovariectomy on thyroid C cells of adult rats. Yugoslov Med Biohem, Vol. 21, pp. 345-350
- Filipović, B., Šošić -Jurjević, B., Nestorović, N., Manojlović Stojanoski, M., Kostić N., Milošević, V. & Sekulić M. (2003). The thyroid C cells of ovariectomized rats treated with estradiol. *Histochem Cell Biol*, Vol. 120, pp. 409-414

- Filipović, B., Šošić -Jurjević, B., Manojlović Stojanoski, M., Nestorović, N., Milošević V. & Sekulić M. (2005). The effect of chronic calcium treatment on thyroid C cells in ovariectomized rats. *Life Sci*, Vol. 77, pp. 121-129
- Filipović, B., Šošić -Jurjević, B., Ajdžanović, V., Trifunović, S., Manojlović Stojanoski, M., Ristić, N., Nestorović, N., Milošević, V. & Sekulić M. (2007) The effect of orchidectomy on thyroid C cells and bone histomorphometry in middle-aged rats. *Histochem Cell Biol*, Vol. 128, pp. 153–159
- Filipović, B., Šošić -Jurjević, B., Ajdzanović, V., Brkić, D., Manojlović-Stojanoski, M., Milosević, V. & Sekulić M. (2010). Daidzein administration positively affects thyroid C cells and bone structure in orchidectomized middle-aged rats. Osteoporos Int, Vol. 21, pp. 1609-1616
- Filipović B., Šošić -Jurjević B., Ajdžanović V., Pantelić J., Nestorović N. & Sekulić M. Estardiol effects the function of neuroendocrine C cells in orchidectomized middle-aged rat thyroid gland. (2010a). *The 7th International Congress of Neuroendocrinology*, p. 180, Rouen, France, July 11-15, 2010
- Fitts, JM., Klein, RM. & Powers, CA. (2001). Estrogen and tamoxifen interplay with T(3) in male rats: pharmacologically distinct classes of estrogen responses affecting growth, bone, and lipid metabolism, and their relation to serum GH and IGF-I. *Endocrinology*, Vol. 142, pp. 4223-4235
- Fonseca, D. & Ward, WE. (2004). Daidzein together with high calcium preserve bone mass and biomechanical strength at multiple sites in ovariectomized mice. *Bone*, Vol. 35, pp. 489-497
- Frost, HM. (1964). Dynamics of bone remodeling. In: *Frost HM (ed) Bone Biodynamics*. *Littel, Brown, Boston,* pp 315-333
- Fuller, K., Wong, B., Fox, S., Choi, Y. & Chambers, TJ. (1998). TRANCE is necessary and sufficient for osteoblast-mediated activation of bone resorption in osteoclasts. J Exp Med, Vol. 188, pp. 997–1001
- Gaberscek, S., Stiblar-Martincic, D. & Kalisnik, M. (1998). The influence of calcium on thyroid follicular cells FRTL-5 in vitro. *Folia Biol (Praha)*, Vol. 44, pp. 49-52
- Gao, YH. & Yamaguchi, M. (1999). Suppressive effect of genistein on rat bone osteoclasts: apoptosis is induced through Ca2+ signaling. *Biol Pharm Bull*, Vol. 22, pp. 805–809
- Gao, YH. & Yamaguchi, M. (2000) Suppressive effect of genistein on rat bone osteoclasts: involvement of protein kinase inhibition and protein tyrosine phosphatase activation. *Int J Mol Med*, Vol. 5, pp. 261-267
- Geppert, J., Baier, S., Zehn, N., Gouni-Berthold, I., Berthold, HK., Reinsberg, J. & Stehle, P. (2004). Short-term effects of high soy supplementation on sex hormones, bone markers, and lipid parameters in young female adults. *Eur J Nutr*, Vol. 43, pp. 100–108
- Giampietro, PG., Bruno, G., Furcolo, G., Casati, A., Brunetti, E., Spadoni, GL. & Galli, E. (2004). Soy protein formulas in children: no hormonal effects in long-term feeding. J Pediatr Endocrinol Metab, Vol. 17, pp. 191-196
- Globus, RK., Plouet, J. & Gospodarowicz, D. (1989). Cultured bovine bone cells synthesize basic fibroblast growth factor and store it in their extracellular matrix. *Endocrinology*, Vol. 124, pp. 1539-1547

- Gogakos, AI., Duncan Bassett, JH. & Williams, GR. (2010). Thyroid and bone. *Arch Biochem* Biophys, Vol. 503, pp. 129-136
- Grauer, A., Klein, P., Naveh-Many, T., Silver, J., Ziegler, R. & Raue, F. (1993). Diminished calcitonin secretion after ovariectomy without apparent reduction in calcitonin content in the rat. *Horm Metab Res*, Vol. 25, pp. 389–390
- Grumbach, MM. & Auchus, RJ. (1999). Estrogen: consequences and implications of human mutations in synthesis and action. *J Clin Endocrinol Metab*, Vol. 84, pp. 4677-4694
- Ham, KD. & Carlson CS. (2004). Effects of estrogen replacement therapy on bone turnover in subchondral bone and epiphyseal metaphyseal cancellous bone of ovariectomized cynomolgus monkeys. *J Bone Miner Res*, Vol. 19, pp. 823-829
- Henderson, KM., McNeilly, AS. & Swanston, IA. (1982). Gonadotrophin and steroid concentrations in bovine follicular fluid and their relationship to follicle size. *J Reprod Fertil*, Vol. 65, pp. 467-473
- Ho, SC., Woo, J., Lam, S., Chen, YM., Sham, A. & Lau, J. (2003). Soy protein consumption and bone mass in early postmenopausal Chinese women. *Osteoporos Int*, Vol. 14, pp. 835–842
- Hsu, HL., Lacey, DL., Dunstan, CR., Solovyev, I., Colombero, A., Timms, E., Tan, HL., Elliott, G., Kelley, MJ., Sarosi, I., Wang, L., Xia, XZ., Elliott, R., Chiu, L., Black, T., Scully, S., Capparelli, C., Morony, S., Shimamoto, G., Bass, MB. & Boyle, WJ. (1999). Tumor necrosis factor receptor family member RANK mediates osteoclast differentiation and activation induced by osteoprotegerin ligand. *Proc Nat Acad Sci*, Vol. 96, pp. 3540–3545
- Huang, YF., Cao, SM., Nagamani, M., Anderson, KE., Grady, JJ. & Lu, LJW. (2005). Decreased circulating levels of tumor necrosis factor-alpha in postmenopausal women during consumption of soy-containing isoflavones. *J Clin Endocr Metab*, Vol. 90, pp. 3956–3962
- Huang, HY., Yang, HP., Yang, HT., Yang, TC., Shieh, MJ. & Huang, SY. (2006). One-year soy isoflavone supplementation prevents early postmenopausal bone loss but without a dosedependent effect. *J Nutr Biochem*, Vol. 17, pp. 509–517
- Hunter, SJ., Schraer, H. & Gay, CV. (1989). Characterization of the cytoskeleton of isolated chick osteoclasts: effect of calcitonin. *J Histochem Cytochem*, Vol. 37, pp. 1529–1537
- Ikeda, T., Nishikawa, A., Imazawa, T., Kimura, S. & Hirose, M. (2000). Dramatic synergism between excess soybean intake and iodine deficiency on the development of rat thyroid hyperplasia. *Carcinogenesis*, Vol. 21, pp. 707-713
- Isaia, GC., Campagnoli, C., Mussetta, M., Massobrio, M., Salamono, G., Gallio, M. & Molinatti, GM. (1989). Calcitonin and lumbar bone mineral content during oestrogen-progesterone administration in postmenopausal women. *Maturitas*, Vol. 11, pp. 287–294
- Isaia, GC., Mussetta, M., Massobrio, M., Sciolla, A., Gallio, M. & Molinatti, GM. (1992). Influence of estrogens on calcitonin secretion. *J Endocrinol Invest*, Vol. 15, pp. 59–62
- Ishimi, Y., Miyaura, C., Ohmura, M., Onoe, Y., Sato, T., Uchiyama, Y., Ito, M., Wang, X., Suda, T. & Ikegami, S. (1999). Selective effects of genistein, a soybean isoflavone, on B-lymphopoiesis and bone loss caused by estrogen deficiency. *Endocrinology*, Vol. 140, pp. 1893-1900

- Ishimi, Y., Yoshida, M., Wakimoto, S., Wu, J., Chiba, H., Wang, X Takeda, K. & Miyaura C. (2002). Genistein, a soybean isoflavone, affects bone marrow lymphopoiesis and prevents bone loss in castrated male mice. *Bone*, Vol. 31, pp. 180-185
- Ito, N., Yamazaki, H., Nakazaki, M., Miyahara, T., Kozuka, H. & Sudo, H. (1987). Response of osteoblastic clonal cell line (MC3T3-E1) to [Asu]eel calcitonin at a specific cell density or differentiation stage. *Calcif Tissue Int*, Vol. 40, pp. 200–205
- Jabbar, MA., Larrea, J. & Shaw, RA. (1997). Abnormal thyroid function tests in infants with congenital hypothyroidism: the influence of soy-based formula. *J Am Coll Nutr*, Vol. 16, pp. 280-282
- Jia, TL., Wang, HZ., Xie, LP., Wang, XY. & Zhang, RQ. (2003). Daidzein enhances osteoblast growth that may be mediated by increased bone morphogenetic protein (BMP) production. *Biochem Pharmacol*, Vol. 65, pp. 709–715
- Kanis, JA., Aaron, JE., Evans, D., Thavarajah, M. & Beneton, M. (1990). Bone loss and agerelated fractures. *Exp Gerontol*, Vol. 25, pp. 289-296
- Katagiri, T. & Takahashi, N. (2002). Regulatory mechanisms of osteoblast and osteoclast differentiation. *Oral Dis*, Vol. 8, pp. 147–159
- Kennedy, JW. & Caro, JF. (1996). The ABC of managing hyperthyroidism in the older patient. *Geriatrics*, Vol. 51, pp. 22-32
- Khalil, DA., Lucas, EA., Smith, BJ., Soung, DY., Devareddy, L., Juma, S., Akhter, MP., Recker, R. & Arjmandi, BH. (2005). Soy isoflavones may protect against orchidectomy-induced bone loss in aged male rats. *Calcif Tissue Int.* Vol. 76, pp. 56– 62
- Khosla, S., Atkinson, EJ., Melton, LJ III. & Riggs, BL. (1997). Effects of age and estrogen status on serum parathyroid hormone levels and biochemical markers of bone turnover in women: a populationbased study. J Clin Endocrinol Metab, Vol. 82, pp. 1522–1527
- Kim, CH., Takai, E., Zhou, H., von Stechow, D., Müller, R., Dempster, DW. & Guo, XE. (2003). Trabecular bone response to mechanical and parathyroid hormone stimulation: the role of mechanical microenvironment. *J Bone Miner Res*, Vol. 18, pp. 2116-2125
- Kimura, S., Suwa, J., Ito, M. (1976). Sato, H. Development of malignant goiter by defatted soybean with iodine-free diet in rats. *Gann*, Vol. 67, pp. 763-765
- Korach, KS. (1994). Insights from the study of animals lacking functional estrogen receptor. *Science*, Vol. 266, pp. 1524-1527
- Kuiper, GG., Lemmen, JG., Carlsson, B., Corton, JC., Safe, SH., van der Saag, PT., van der Burg, B. & Gustafsson, JA. (1998). Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. *Endocrinology*, Vol. 139, pp. 4252–4263
- Kuiper, GG., Lemmen, JG., Carlsson, B., Corton, JC., Safe, SH., van der Saag, PT., van der Burg, B. & Gustafsson, JA. (1998). Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. *Endocrinology*, Vol. 139, pp. 4252–4263
- Kung, AWC. (1994). The effect of thyroid hormone on bone metabolism and osteoporosis. J Hong Kong Med Assoc, Vol. 46, pp. 247-251
- Lakatos, P. (2003). Thyroid hormones: beneficial or deleterious for bone? *Calcif Tissue Int*, Vol. 73, pp. 205-209
- Lee, CA. & Einhorn, T. (2001). In: Osteoporosis, edited by Marcus, Feldman & Kelsey, pp. 3-20

- Lee, YB., Lee, HJ., Kim, KS., Lee, JY., Nam, SY., Cheon, SH. & Sohn HS. (2004). Evaluation of the preventive effect of isoflavone extract on bone loss in ovariectomized rats. *Bioscience, Biotechnology and Biochemistry*, Vol. 68, pp. 1040-1045
- Li, BB. & Yu, SF. (2003). Genistein prevents bone resorption diseases by inhibiting bone resorption and stimulating bone formation. *Biol Pharm Bull*, Vol. 26, pp. 780–786
- Lindsay, R., Hart, DM., Aitken, JM., MacDonald, ED., Anderson, JB. & Clarke, AC. (1976). Long-term prevention of postmenopausal osteoporosis by oestrogen. *Lancet*, Vol. 1, pp. 1038–1041
- Lindsay, R., Hart, DM. & Clark, DM. (1984). The minimum effective dose of estrogen for prevention of postmenopausal bone loss. *Obstet Gynecol*, Vol. 63, pp. 759–763
- Loughlin, K. & Richie, J. (1997). Prostate cancer after exogenous testosterone treatment for impotence. *J Urology*, Vol. 157, pp.1845
- Lu, CC., Tsai, SC., Chien, EJ., Tsai, CL. & Wang, PS. (2000). Age-related differences in the secretion of calcitonin in male rats. *Metabolism*, Vol. 49, pp. 253–258
- Lydeking-Olsen, E., Beck-Jensen, JE., Setchell, KD. & Holm-Jensen, T. (2004). Soymilk or progesterone for prevention of bone loss: a 2 year randomized, placebo- ontrolled trial. *Eur J Nutr*. Vol. 43, pp. 246–257
- Maugeri, D., Salvatore Russo, M., Carnazzo, G., Di Stefano, F., Catanzaro, S., Campagna, S., Romano, G., Franze, C., Motta, M., Panebianco, P. (1996). Altered laboratory thyroid parameters indicating hyperthyroidism in elderly subjects. *Arch Gerontol Geriatr*, Vol. 22, pp. 145-153
- McCarrison, R. (1993). A Paper on FOOD AND GOITRE. Br Med J, Vol. 14, pp. 671-675
- Mei, J., Yeung, SS. & Kung, AW. (2001). High dietary phytoestrogen intake is associated with higher bone mineral density in postmenopausal but not premenopausal women. J Clin Endocrinol Metab, Vol. 86, pp. 5217-5221
- Milne, M., Quail, JM., Rosen, CJ. & Baran, DT. (2001). Insulin-like growth factor binding proteins in femoral and vertebral bone marrow stromal cells: expression and regulation by thyroid hormone and dexamethasone. J Cell Biochem, Vol. 81, pp. 229-240
- Mori, K., Stone, S., Braverman, LE. & Devito, WJ. (1996). Involvement of tyrosine phosphorylation in the regulation of 5'-deiodinases in FRTL-5 rat thyroid cells and rat astrocytes. *Endocrinology*, Vol. 137, pp. 1313-1318
- Mundy, GR. (1995). Local control of bone formation by osteoblasts. *Clin Orthop Relat Res*, Vol. 313, pp. 19-26
- Nakai, M., Cook, L., Pyter, LM., Black, M., Sibona, J., Turner, RT., Jeffery, EH. & Bahr, JM. (2005). Dietary soy protein and isoflavones have no significant effect on bone and a potentially negative effect on the uterus of sexually mature intact Sprague-Dawley female rats. *Menopause*, Vol. 12, pp. 291-298
- Nelson, HD., Humphrey, LL., Nygren, P., Teutsch, SM. & Allan, JD. (2002). Postmenopausal hormone replacement therapy: scientific review. *JAMA*, Vol. 288, pp. 872–881
- Newton, KM., LaCroix, AZ., Levy, L., Li, SS., Qu, P., Potter, JD. & Lampe, JW. (2006) Soy protein and bone mineral density in older men and women: a randomized trial. *Maturitas*, Vol. 55, pp. 270–277
- Okamoto, F., Okabe, K. & Kajiya, H. (2001). Genistein, a soybean isoflavone, inhibits inward rectifier K+ channels in rat osteoclasts. *Jap J Physiol*, Vol. 51, pp. 501–509

- Okura, A., Arakawa, H., Oka, H., Yoshinari, T. & Monden, Y. (1998). Effect of genistein on topoisomerase activity and on the growth of [val 12] Ha-ras transformed NIH 3T3 cells. *Biochem Biophys Res Commun*, Vol. 157, pp. 183-189
- Om, AS. & Shim, JY. (2007). Effect of daidzein, a soy isoflavone, on bone metabolism in Cdtreated ovariectomized rats. *Acta Biochim Pol*, Vol. 54, pp. 641-646
- Onoe, Y., Miyaura, C., Ohta, H., Nozawa, S. & Suda, T. (1997). Expression of estrogen receptor in rat bone. *Endocrinology*, Vol. 138, pp. 4509-4512
- O'Shea, PJ., Harvey, CB., Suzuki, H., Kaneshige, M., Kaneshige, K., Cheng, SY. & Williams, GR. (2003). A thyrotoxic skeletal phenotype of advanced bone formation in mice with resistance to thyroid hormone. *Mol Endocrinol*, Vol. 17, pp. 1410-1424
- Pan, W., Quarles, LD., Song, LH., Yu, YH., Jiao, C., Tang, HB., Jiang, CH., Deng, HW., Li, YJ., Zhou, HH. & Xiao, ZS. (2005). Genistein stimulates the osteoblastic differentiation via NO/cGMP in bone marrow culture. J Cell Biochem, Vol. 94, pp. 307–316
- Pantelić, J., Filipović, B., Šošić -Jurjević, B., Medigović, I. & Sekulić, M. (2010). Effects of testosterone and estradiol treatment on bone histomorphometry in orchidectomized middle-aged rats. *Proceedings of 4th Serbian Congress for Microscopy*, pp. 143-144, Belgrade, Serbia, October 11-12, 2010
- Patisaul, HB. & Jefferson, W. (2010). The pros and cons of phytoestrogens. *Front Neuroendocrinol*, Vol. 31, pp. 400-419
- Peeters, RP., van der Deure, WM. & Visser, TJ. (2006). Genetic variation in thyroid hormone pathway genes; polymorphisms in the TSH receptor and the iodothyronine deiodinases. *Eur J Endocrinol*, Vol. 155, pp. 655-662
- Pereira, RC., Jorgetti, V. & Canalis, E. (1999). Triiodothyronine induces collagenase-3 and gelatinase B expression in murine osteoblasts. *Am J Physiol*, Vol. 77, pp. E496-E504
- Picherit, C., Bennetau-Pelissero, C., Chanteranne, B., Lebecque, P., Davicco, MJ., Barlet, JP. & Coxam, V. (2001). Soybean isoflavones dose-dependently reduce bone turnover but do not reverse established osteopenia in adult ovariectomized rats. *J Nutr*, Vol. 131, pp. 723-728
- Poirier, LA., Doerge, DR., Gaylor, DW, Miller, MA, Lorentzen, RJ., Casciano, DA., Kadlubar, FF. & Schwetz, BA. (1999). An FDA review of sulfamethazine toxicity. *Regul Toxicol Pharmacol*, Vol. 30 pp. 217-222
- Ramos, S. (2007). Effects of dietary flavonoids on apoptotic pathways related to cancer chemoprevention. *J Nutr Biochem*, Vol. 18, pp. 427-442
- Rassi, CM., Lieberherr, M., Chaumaz, G., Pointillart, A. & Cournot, G. (2002). Downregulation of osteoclast differentiation by daidzein via caspase 3. *J Bone Miner Res*, Vol. 17, pp. 630-638
- Register, TC., Jayo, MJ. & Anthony, MS. (2003). Soy phytoestrogens do not prevent bone loss in postmenopausal monkeys. *J Clin Endocrinol Metab*, Vol. 88, pp. 4362-4370
- Rehman, HU. & Masson, EA. (2005). Neuroendocrinology of female aging. *Gender Medicine*, Vol. 2, pp. 41-56
- Reinholt, FP., Hultenby, K., Oldberg, A. & Heinegård, D. (1990). Osteopontin--a possible anchor of osteoclasts to bone. *Proc Natl Acad Sci U S A*, Vol. 87, pp. 4473-4475

- Ren, P., Ji, H., Shao, Q., Chen, X., Han, J. & Sun, Y. (2007).Protective effects of sodium daidzein sulfonate on trabecular bone in ovariectomized rats. *Pharmacology*, Vol. 79, pp. 129-136
- Reymond, F., Dénéréaz, N. & Lemarchand-Béraud, T. (1992). Thyrotropin action is impaired in the thyroid gland of old rats. *Acta Endocrinol (Copenh)*, Vol. 126, pp. 55-63
- Rivkees, SA., Bode, HH. & Crawford, JD. (1988). Long-term growth in juvenile acquired hypothyroidism: the failure to achieve normal adult stature. *N Engl J Med*, Vol. 318, pp. 599-602
- Rizzoli, R., Poser, J. & Bürgi, U. (1986). Nuclear thyroid hormone receptors in cultured bone cells. *Metabolism*, Vol. 35, pp. 71-74
- Roef, G., Lapauw, B., Goemaere, S., Zmierczak, H., Fiers, T., Kaufman, J.M. & Taes Y. (2011). Thyroid hormone status within the physiological range affects bone mass and density in healthy men at the age of peak bone mass. *Eur J Endocrinol*, Vol. 164, pp. 1027-1034
- Ron, E., Kleinerman, RA., Boice, JDJr., LiVolsi, VA., Flannery, JT. & Fraumeni, JFJr. (1987). A population-based case-control study of thyroid cancer. J Natl Cancer Inst, Vol. 79, pp. 1-12
- Rydziel, S., Shaikh, S. & Canalis, E. (1994). Platelet-derived growth factor-AA and -BB (PDGF-AA and -BB) enhance the synthesis of PDGF-AA in bone cell cultures. *Endocrinology*, Vol. 134, pp. 2541-2546
- Sakai, K., Yamada, S. & Yamada, K. (2000). Effect of ovariectomy on parafollicular cells in the rat. *Okajimas Folia Anat Jpn*, Vol. 76, pp. 311–319
- Sampath, TK., Simic, P., Sendak, R., Draca, N., Bowe, AE., O'Brien, S., Schiavi, SC., McPherson, JM. & Vukicevic, S. (2007). Thyroid-stimulating hormone restores bone volume, microarchitecture, and strength in aged ovariectomized rats. J Bone Miner Res, Vol. 22, pp. 849-859
- Schindler, AE. (2003). Thyroid function and postmenopause. *Gynecol* Endocrinol, Vol. 17, pp. 79-85
- Schmutzler, C., Hamann, I., Hofmann, PJ., Kovacs, G., Stemmler, L., Mentrup, B., Schomburg, L., Ambrugger, P., Grüters, A., Seidlova-Wuttke, D., Jarry, H., Wuttke, W. & Köhrle, J. (2004). Endocrine active compounds affect thyrotropin and thyroid hormone levels in serum as well as endpoints of thyroid hormone action in liver, heart and kidney. *Toxicology*, Vol. 205, pp. 95-102
- Schweizer, U., Braun, D., Köhrle, J. & Hershman J. (2010). Tyrosine kinase inhibitors non competitively inhibit MCT8-mediated iodothyronine transport. 14th International Thyroid Congress, Paris, 11-16 September, LB-12
- Sekulić, M., Šošić -Jurjević, B., Filipović, B., Milošević, V., Nestorović, N. & Manojlović-Stojanoski, M. (2005). The effects of synthetic salmon calcitonin on thyroid C and follicular cells in adult female rats. *Folia Histochem Cytobiol*. Vol. 43, pp. 103-108
- Sekulić, M., Šošić -Jurjević, B., Filipović, B., Manojlović-Stojanoski, M. & Milosević, V. (2006). Immunoreactive TSH cells in juvenile and peripubertal rats after estradiol and human chorionic gonadotropin treatment. *Acta Histochem*, Vol. 108, pp. 117-123
- Sekulić, M., Šošić -Jurjević, B., Filipović, B., Nestorović, N., Negić, N., Stojanoski, MM. & Milosević, V. (2007). Effect of estradiol and progesterone on thyroid gland in pigs: a

histochemical, stereological, and ultrastructural study. *Microsc Res Tech*, Vol. 70, pp. 44-49

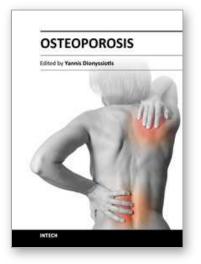
- Sekulić, M., Šošić -Jurjević, B., Filipović, B., Ajdzanović, V., Pantelić, J., Nestorović, N., Manojlović-Stojanoski, M. & Milosević V. (2010). Testosterone and estradiol differently affect thyroid structure and function i orchidectomized middle—aged rats. 14th International Thyroid Congress, Paris p. 0302
- Sendak, RA., Sampath, TK. & McPherson, JM. (2007). Newly reported roles of thyroidstimulating hormone and follicle-stimulating hormone in bone remodelling. *Int Orthop*, Vol. 31, pp. 753-757
- Setchell, KD. (1998). Phytoestrogens: the biochemistry, physiology, and implications for human health of soy isoflavones. *Am J Clin* Nutr, Vol. 68, pp. 1333S-1346S
- Setchell, KDR., Brown, NM. & Lydeking-Olsen, E. (2002). The clinical importance of the metabolite equolVa clue to the effectiveness of soy and its isoflavones. *J Nutr*, Vol. 132, pp. 3577-3584
- Šošić -Jurjević, B., Filipović, B., Nestorović, N., Lovren, M. & Sekulić, M. (2002), Effect of calcium on structural and morphometric features of thyroid gland tissue in middle-aged rat females. *Jug Med Biochem*, Vol. 21, pp. 261-267
- Šošić -Jurjević, B., Filipović, B., Milosević, V., Nestorović, N., Manojlović-Stojanoski, M., Brkić, B. & Sekulić, M. (2005). Chronic estradiol exposure modulates thyroid structure and decreases T4 and T3 serum levels in middle-aged female rats. *Horm Res*, Vol. 63, pp. 48-54
- Šošić -Jurjević, B., Filipović, B., Milosević, V., Nestorović, N., Negić, N. & Sekulić, M. (2006a). Effects of ovariectomy and chronic estradiol administration on pituitary-thyroid axis in adult rats. *Life Sci*, Vol. 79, pp. 890-897
- Šošić -Jurjević, B., Filipović, B., Stojanoski-Manojlović, M. & Sekulić, M. (2006). Calcium administration decreases thyroid functioning in middle-aged female rats Archives of Biological Sciences, Vol. 58, pp. 31-32
- Šošić -Jurjević, B., Filipović, B., Ajdzanović, V., Savin, S., Nestorović, N., Milosević, V. & Sekulić, M. (2010). Suppressive effects of genistein and daidzein on pituitarythyroid axis in orchidectomized middle-aged rats. *Exp Biol Med (Maywood)*, Vol. 235, pp. 590-598
- Soung, DY., Devareddy, L., Khalil, DA., Hooshmand, S., Patade, A., Lucas, EA. & Arjmandi, BH. (2006). Soy affects trabecular microarchitecture and favorably alters select bone-specific gene expressions in a male rat model of osteoporosis. *Calcif Tissue Int*, Vol. 78, pp. 385–391
- Suchartwatnachai, C., Thepppisai, U. & Jirapinyo, M. (2002). Screening for hypothyroidism at a menopause clinic. *Int J Gynaecol Obstet*, Vol. 77, pp. 39-40
- Sugimoto, E. & Yamaguchi, M. (2000). Anabolic effect of genistein in osteoblastic MC3T3-E1 cells. *Int J Mol Med*, Vol. 5, pp. 515-520
- Sugimoto, E. & Yamaguchi, M. (2000a). Stimulatory effect of Daidzein in osteoblastic MC3T3-E1 cells. *Biochem Pharmacol*, Vol. 59, pp. 471-475
- Suh, KS., Koh, G., Park, CY., Woo, JT., Kim, SW., Kim, JW., Park, IK. & Kim, YS. (2003). Soybean isoflavones inhibit tumor necrosis factor-alpha-induced apoptosis and the production of interleukin-6 and prostaglandin E-2 in osteoblastic cells. *Phytochemistry*, Vol. 63, pp. 209–215

- Sun, L., Vukicevic, S., Baliram, R., Yang, G., Sendak, R., McPherson, J., Zhu, L.L., Iqbal, J., Latif, R., Natrajan, A., Arabi, A., Yamoah, K., Moonga, BS., Gabet, Y., Davies, TF., Bab, I., Abe, E., Sampath, K. & Zaidi, M. (2008). Intermittent recombinant TSH injections prevent ovariectomy-induced bone loss. *Proc Natl Acad Sci USA*, Vol. 105, pp. 4289-4294
- Teitelbaum, SL. (2000). Bone resorption by osteoclasts. Science, Vol. 289, pp. 1504-1508
- Tham, DM., Gardner, CD. & Haskell, WL. (1998). Clinical review 97: potential health benefits of dietary phytoestrogens: a review of the clinical, epidemiological, and mechanistic evidence. *J Clin Endocrinol Metab*, Vol. 83, pp. 2223–2235
- Theoleyre, S., Wittrant, Y., Tat, SK., Fortun, Y., Redini, F. & Heymann, D. (2004). The molecular triad OPG/RANK/RANKL: involvement in the orchestration of pathophysiological bone remodeling. *Cytokine Growth Factor Rev*, Vol. 15, pp. 457– 475
- Trisomboon, H., Malaivijitnond, S., Suzuki, J., Hamada, Y., Watanabe, G. & Taya, K. (2004). Long-term treatment effects of Pueraria mirifica phytoestrogens on parathyroid hormone and calcium levels in aged menopausal cynomolgus monkeys. J Reprod Dev, Vol. 50, pp. 639-645
- Tuohy, PG. (2003). Soy infant formula and phytoestrogens. J Paediatr Child Health, Vol. 39, pp. 401-405
- Udagawa, N., Takahashi, N., Jimi, E., Matsuzaki, K., Tsurukai, T., Itoh, K., Nakagawa, N., Yasuda, H., Goto, M., Tsuda, E., Higashio, K., Gillespie, MT., Martin, TJ. & Suda, T. (1999). Osteoblasts/stromal cells stimulate osteoclast activation through expression of osteoclast differentiation factor/RANKL but not macrophage colony-stimulating factor. *Bone*, Vol. 25, pp. 517–523
- Väänänen, HK., Zhao, H., Mulari, M. & Halleen JM. (2000). The cell biology of osteoclast function. *J Cell Sci*, Vol. 113, pp. 377-381
- van der Deure, WM., Peeters, RP. & Visser, TJ. (2010). Molecular aspects of thyroid hormone transporters, including MCT8, MCT10, and OATPs, and the effects of genetic variation in these transporters. *J Mol Endocrinol*, Vol. 44, pp. 1-11
- Van Wyk, JJ., Arnold, MB., Wynn, J. & Pepper, F. (1959). The effects of a soybean product on thyroid function in humans. *Pediatrics*, Vol. 24, pp. 752-760
- Vandenput, L., Ederveen, AG., Erben, RG., Stahr, K., Swinnen, JV., Van Herck, E., Verstuyf, A., Boonen, S., Bouillon, R. & Vanderschueren, D. (2001). Testosterone prevents orchidectomy-induced bone loss in estrogen receptor-alpha knockout mice. *Biochem Biophys Res Commun*, Vol. 285, pp. 70-76
- Vanderschueren, D., Van Herck, E., Suiker, AMH., Visser, WJ., Schot, LPC. & Bouillon, R. (1992). Bone and mineral metabolism in aged male rats: short- and long-term effects of androgen deficiency. *Endocrinology*, Vol. 130, pp. 2906–2916
- Vanderschueren, D., Boonen, S. & Bouillon, R. (1998). Action of androgens versus estrogens in male skeletal homeostasis. *Bone*, Vol. 23, pp. 391-394
- Vanvooren, V., Allgeier, A., Cosson, E., Van Sande, J., Defer, N., Pirlot, M., Hanoune, J. & Dumont, JE. (2000). Expression of multiple adenylyl cyclase isoforms in human and dog thyroid. *Mol Cell Endocrinol*, Vol. 170, pp. 185-196

- Varga, F., Spitzer, S. & Klaushofer, K. (2004). Triiodothyronine (T3) and 1,25dihydroxyvitamin D3 (1,25D3) inversely regulate OPG gene expression in dependence of the osteoblastic phenotype. *Calcif Tissue Int*, Vol. 74, pp. 382-387
- Villa, I., Dal, Fiume, C., Maestroni, A., Rubinacci, A., Ravasi, F. & Guidobono, F. (2003). Human osteoblast-like cell proliferation induced by calcitonin-related peptides involves PKC activity. *Am J Physiol Endocrinol Metab*, Vol. 284, pp. E627–E633
- Wangen, KE., Duncan, AM., Merz-Demlow, BE., Xu, X., Marcus, R., Phipps, WR. & Kurzer, MS. (2000). Effects of soy isoflavones on markers of bone turnover in premenopausal and postmenopausal women. J Clin Endocr Metab, Vol. 85, pp. 3043–3048
- Watanabe, K., Takekoshi, S. & Kakudo, K. (1992). Effects of ipriflavone on calcitonin synthesis in C cells of the rat thyroid. *Calcif Tissue Int*, Vol. 51, pp. S27–S29
- Weiner, KX. & Dias, JA. (1990). Protein synthesis is required for testosterone to decrease ornithine decarboxylase messenger RNA levels in rat Sertoli cells. *Mol Endocrinol*, Vol. 4, pp. 1791–1798
- Williams, AJ., Robson, H., Kester, MH., van Leeuwen, JP., Shalet, SM., Visser, TJ. & Williams, GR. (2008). Iodothyronine deiodinase enzyme activities in bone. *Bone*, Vol. 43, pp. 126-134
- Williams, JP., Jordan, SE., Barnes, S. & Blair, HC. (1998). Tyrosine kinase inhibitor effects on avian osteoclastic acid transport. *Am J Clin Nutr*, Vol. 68, pp. 1369S-1374S
- Wong, C., Lai, T., Hilly, JM., Stewart, CE. & Farndon, JR. (2002). Selective estrogen receptor modulators inhibit the effects of insulin-like growth factors in hyperparathyroidism. *Surgery*, Vol. 132, pp. 998-1006
- Wu, J., Wang, XX., Chiba, H., Higuchi, M., Takasaki, M., Ohta, A. & Ishimi, Y. (2003). Combined intervention of exercise and genistein prevented and rogen deficiencyinduced bone loss in mice. *J Appl Physiol*, Vol. 94, pp. 335–342
- Wu, J., Wang, X., Chiba, H., Higuchi, M., Nakatani, T., Ezaki, O., Cui, H., Yamada, K. & Ishimi, Y. (2004). Combined intervention of soy isoflavone and moderate exercise prevents body fat elevation and bone loss in ovariectomized mice. *Metabolism*, Vol. 53, pp. 942-948
- Wu, J., Oka, J., Tabata, I., Higuchi, M., Toda, T., Fuku, N., Ezaki, J., Sugiyama, F., Uchiyama, S., Yamada, K. & Ishimi, Y. (2006). Effects of isoflavone and exercise on BMD and fat mass in postmenopausal Japanese women: a 1-year randomized placebo-controlled trial. *J Bone Miner Res*, Vol. 21, pp. 780–789
- Yamagishi, T., Otsuka, E. & Hagiwara, H. (2001). Reciprocal control of expression of mRNAs for osteoclast differentiation factor and OPG in osteogenic stromal cells by genistein: evidence for the involvement of topoisomerase II in osteoclastogenesis. *Endocrinology*, Vol. 142, pp. 3632-3637
- Yamaguchi, M. & Sugimoto, E. (2000). Stimulatory effect of genistein and daidzein on protein synthesis in osteoblastic MC3T3-E1 cells: activation of aminoacyl-tRNA synthetase. *Mol Cell Biochem*, Vol. 214, pp. 97-102
- Yen, PM. (2001). Physiological and molecular basis of thyroid hormone action. *Physiol Rev*, Vol. 81, pp. 1097-1142

Zaidi, M., Datta, HK., Moonga, BS. & MacIntyre, I. (1990). Evidence that the action of calcitonin on rat osteoclasts is mediated by two G proteins acting via separate post-receptor pathways. *J Endocrinol*, Vol. 126, pp. 473–481





Osteoporosis Edited by PhD. Yannis Dionyssiotis

ISBN 978-953-51-0026-3 Hard cover, 864 pages Publisher InTech Published online 24, February, 2012 Published in print edition February, 2012

Osteoporosis is a public health issue worldwide. During the last few years, progress has been made concerning the knowledge of the pathophysiological mechanism of the disease. Sophisticated technologies have added important information in bone mineral density measurements and, additionally, geometrical and mechanical properties of bone. New bone indices have been developed from biochemical and hormonal measurements in order to investigate bone metabolism. Although it is clear that drugs are an essential element of the therapy, beyond medication there are other interventions in the management of the disease. Prevention of osteoporosis starts in young ages and continues during aging in order to prevent fractures associated with impaired quality of life, physical decline, mortality, and high cost for the health system. A number of different specialties are holding the scientific knowledge in osteoporosis. For this reason, we have collected papers from scientific departments all over the world for this book. The book includes up-to-date information about basics of bones, epidemiological data, diagnosis and assessment of osteoporosis, secondary osteoporosis, pediatric issues, prevention and treatment strategies, and research papers from osteoporotic fields.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Branko Filipović and Branka Šošić-Jurjević (2012). The Phytoestrogens, Calcitonin and Thyroid Hormones: Effects on Bone Tissue, Osteoporosis, PhD. Yannis Dionyssiotis (Ed.), ISBN: 978-953-51-0026-3, InTech, Available from: http://www.intechopen.com/books/osteoporosis/the-phytoestrogens-calcitonin-and-thyroid-hormones-effects-on-bone-tissue

INTECH

open science | open minds

InTech Europe

University Campus STeP Ri Slavka Krautzeka 83/A 51000 Rijeka, Croatia Phone: +385 (51) 770 447 Fax: +385 (51) 686 166 www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai No.65, Yan An Road (West), Shanghai, 200040, China 中国上海市延安西路65号上海国际贵都大饭店办公楼405单元 Phone: +86-21-62489820 Fax: +86-21-62489821 © 2012 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the <u>Creative Commons Attribution 3.0</u> <u>License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen